Biocompatibility and biomineralization of the experimental nanoparticulate mineral trioxide aggregate (MTA)

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Abstract
To investigate the tissue response and the biomineralization ability of the experimental nanoparticulate mineral trioxide aggregate compared to grey MTA and Fillapex MTA. Polyethylene tubes containing materials or empty tubes for control were inserted into the subcutaneous tissues of 30 rats. After 7, 15, 30, 60, and 90 days, the rats were killed and the tubes were removed for analysis using hematoxylin-eosin staining, von Kossa staining, and under polarized light. Inflammation was graded through a score system; the biomineralization ability was recorded as present or absent. The results were statistically analyzed using the Kruskal-Wallis test (p<0.05). On days 7 and 15 there was a significant difference between the Nano MTA (median score of 3) and MTA Fillapex groups (median score of 4), being MTA Fillapex the material with the highest number of inflammatory cells. At 30, 60, and 90 days there was no difference between the Nano MTA, Grey MTA, and MTA Fillapex groups. All materials induced the formation of mineralized tissue in all experimental periods. Nano MTA showed biocompatibility and biomineralization similar to grey MTA Angelus.

Keywords: Materials testing; Nanoparticles; Endodontics; Root canal therapy.
1. Introduction

Biocompatible materials able to induce the development of biomineralized tissue are of great interest in endodontic clinical applications such as root perforations (Hashem & Hassani, 2008), root-end fillings in apical surgeries (Baek, Plenk, & Kim, 2005), pulp capping (Farsi, Alamoudi, Balto, & Al Mushayt, 2006), apical plug in apexifications (Simon, Rilliard, Berdal, & Machtou, 2007), and as a coronal barrier in revascularization (Moreno-Hidalgo, Caleza-Jimenez, Mendoza-Mendoza, & Iglesias-Linares, 2014). All of these procedures imply contact with living tissues and body fluids, an environment that may affect the physical aspects of the employed material (Nekooefar, Stone, & Dummer, 2010).

Materials based on mineral trioxide aggregate (MTA) are the gold standard for the aforementioned cases (Parirokh, Torabinejad, & Dummer, 2018; Torabinejad, Parirokh, & Dummer, 2018). These materials with a composition largely based on Portland cement components exhibit a hydrophilic nature, enabling its application even in the presence of moisture, once the main elements are tricalcium and dicalcium silicate, tricalcium aluminate, tricalcium oxide, and radiopacifying agents, such as bismuth oxide (J. Camilleri, 2008).

Although it is a material of excellent biological properties, some physicochemical and working properties are lacking. As a result, infinite modifications and preparations have been developed and tested to circumvent these drawbacks. However, none of these new modifications presents powders in a nano-sized, that is particles with 1/1,000 of a micron. Nano-sized particles are of interest because of the high surface-to-volume ratio versus micron-sized particles. The surface-to-volume ratio changes from less than 10 % for micron-sized particles to more than 50 % for nanoparticles, which is essential because a higher surface-to-volume ratio can dramatically increase reactivity, such as hydration and MTA microhardness (Josette Camilleri, 2014).

Hence, this study evaluated a nanoparticulate mineral trioxide aggregate (Nano MTA), analyzing its in vivo reaction in the subcutaneous tissue of rats, and its power to induce mineralization assessed by von Kossa staining and structures
birefringent to polarized light. Grey MTA and MTA Fillapex were used for comparison.

2. Methodology

Animals

Thirty male Wistar rats aged 3 months were used in this study. Sample size estimates were based on data from previous studies using six animals per group (de Azevedo Queiroz et al., 2018; Viola et al., 2012). They were arranged into five groups, each group corresponding to an experimental time: 7, 15, 30, 60 and 90 days. The rats were kept in temperature-controlled housings receiving water and diet ad libitum. Animal care was provided according to the Ethics Committee of Araçatuba School of Dentistry-UNESP for animal use, which was approved before the beginning of the experiments (FOA no. 374-2017) We performed the study following the Animal Research Reporting In Vivo Experiment (ARRIVE) guidelines.

Surgical procedures

Polyethylene tubes (Abbott Labs, São Paulo, Brazil) with a 1.0-mm internal diameter, 1.6-mm outside diameter, and 10.0-mm length were filled with Nano MTA (UNESP, Ilha Solteira, Brazil), grey MTA (Angelus, Londrina, Brazil), MTA Fillapex (Angelus) prepared according to the company's guidelines. Empty tubes were used as control. The surgical procedure was performed according to previous studies (L. Cosme-Silva et al., 2019). The rats were anesthetized with ketamine (87 mg/kg Francotar; Virbac do Brasil, Roseira, Brazil) and xylazine (13 mg/kg Rompum; Bayer S A, São Paulo, Brazil) administered intramuscularly (Dal-Fabbro et al., 2019); their dorsal part was shaved, the skin disinfected with 5% iodine solution, and a 2.0-cm incision was made in a head-to-tail direction with a #15 Bard-Parker blade (BD, Franklin Lakes, USA). The skin was reflected to create four pockets around the incision into which the tubes filled with each material and the empty tube were randomly implanted. The skin was closed with a 4.0 silk suture (Ethicon 4.0-Johnson & Johnson, São Paulo, Brazil).

Histological analysis

At the 7, 15, 30, 60, and 90 days after implantation, the rats were euthanized by an excessive dose of anaesthetic solution: sodium thiopental (Thiopentax; Cristalia, Itapira, Brazil), and the tubes with surrounding tissue were removed, fixed in 10% formalin solution (pH = 7.0), cut transversely in halves, and one half processed with paraffin (Leopoldo Cosme-Silva, Renan Dal-Fabbro, et al., 2019). Serial sections 5-mm thick were obtained and stained with hematoxylin-eosin, whereas 10-mm sections were processed by von Kossa staining or directly examined under polarized light. Histological slices were evaluated under light-field illumination through a microscope with 400x magnification (DM 4000 B; Leica, Wetzlar, Germany) by a single calibrated investigator. Haematoxylin-eosin-stained inflammatory cells close to the material was scored as follows: 1, no or few inflammatory cells and no reaction; 2, less than 25 cells and mild reaction; 3, between 25 and 125 cells and moderate reaction; and 4, 125 or more cells and severe reaction (Leopoldo Cosme-Silva, Francine Benetti, et al., 2019). Biomineralization were assessed through von Kossa and birefringent structures to the polarized light, being recorded as present or absent (Cosme-Silva et al., 2020; Sales et al., 2021).

Statistical analysis

Using the GraphPad Prism (version 8.0) software (GraphPad Software Inc, La Jolla, USA), the Kruskal-Wallis test was performed, followed by Dunn's test. Values of p < 0.05 were considered significant.
3. Results and Discussion

Histological Analysis

Representative images of the groups can be observed in Figure 1. On days 7 and 15 there was a significant difference between the groups Nano MTA (median score of 3) and MTA Fillapex (median score of 4), being MTA Fillapex the material with the highest number of inflammatory cells in the fibrous capsule (p < 0.05). Compared to the gold standard (Grey MTA Angelus), Nano MTA demonstrated a similar inflammatory reaction (p > 0.05). After 15 days, it was possible to observe a reduction on the inflammatory infiltrate in all groups, until they reach a mild inflammatory response at 90 days. At 30, 60 and 90 days there was no difference between the groups Nano MTA, Grey MTA, and MTA Fillapex (p > 0.05). Regarding the biomineralization, with exception of the control group, all materials induced the formation of mineralized tissue in all experimental periods, evidenced by the presence of structures darkly stained by the Von Kossa technique and the birefringent structures observed under polarized light Figure 2, Table 1.

Table 1: Inflammatory Score and Biomineralization Ability (%) of all groups. *Different letters indicate statistical difference among the materials in the same time period (P < 0.05).

<table>
<thead>
<tr>
<th>Time / P value</th>
<th>Groups</th>
<th>Inflammatory SCORE</th>
<th>Median</th>
<th>Biomineralization ability (%)</th>
</tr>
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<tbody>
<tr>
<td>7 days / p = 0.01</td>
<td>Nano MTA A</td>
<td>0 0 5 1</td>
<td>3</td>
<td>100</td>
</tr>
<tr>
<td></td>
<td>Grey MTA A</td>
<td>0 0 5 1</td>
<td>3</td>
<td>100</td>
</tr>
<tr>
<td></td>
<td>MTA Fillapex B</td>
<td>0 0 0 6</td>
<td>4</td>
<td>100</td>
</tr>
<tr>
<td></td>
<td>Control AB</td>
<td>0 0 4 2</td>
<td>3</td>
<td>0</td>
</tr>
<tr>
<td>15 days / p = 0.01</td>
<td>Nano MTA A</td>
<td>0 3 3 0</td>
<td>2.5</td>
<td>100</td>
</tr>
<tr>
<td></td>
<td>Grey MTA A</td>
<td>0 3 3 0</td>
<td>2.5</td>
<td>100</td>
</tr>
<tr>
<td></td>
<td>MTA Fillapex B</td>
<td>0 0 2 4</td>
<td>4</td>
<td>100</td>
</tr>
<tr>
<td></td>
<td>Control A</td>
<td>0 3 3 0</td>
<td>2.5</td>
<td>0</td>
</tr>
<tr>
<td>30 days / p = 0.29</td>
<td>Nano MTA A</td>
<td>1 5 0 0</td>
<td>2</td>
<td>100</td>
</tr>
<tr>
<td></td>
<td>Grey MTA A</td>
<td>1 5 0 0</td>
<td>2</td>
<td>100</td>
</tr>
<tr>
<td></td>
<td>MTA Fillapex A</td>
<td>0 4 2 0</td>
<td>2</td>
<td>100</td>
</tr>
<tr>
<td></td>
<td>Control A</td>
<td>2 3 1 0</td>
<td>2</td>
<td>0</td>
</tr>
<tr>
<td>60 days / p = 0.78</td>
<td>Nano MTA A</td>
<td>2 4 0 0</td>
<td>2</td>
<td>100</td>
</tr>
<tr>
<td></td>
<td>Grey MTA A</td>
<td>3 3 0 0</td>
<td>1.5</td>
<td>100</td>
</tr>
<tr>
<td></td>
<td>MTA Fillapex A</td>
<td>2 3 1 0</td>
<td>2</td>
<td>100</td>
</tr>
<tr>
<td></td>
<td>Control A</td>
<td>3 3 0 0</td>
<td>1.5</td>
<td>0</td>
</tr>
<tr>
<td></td>
<td>4</td>
<td>2</td>
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</tr>
<tr>
<td>Nano MTA ^</td>
<td>4</td>
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<td>Grey MTA ^</td>
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<tr>
<td>MTA Fillapex</td>
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<td>3</td>
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<td>0</td>
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<tr>
<td>Control ^</td>
<td>3</td>
<td>3</td>
<td>0</td>
<td>0</td>
</tr>
</tbody>
</table>

90 days / \( p = 0.88 \)

Source: Authors.

**Figure 1:** Representative images of the subcutaneous tissue reactions in the Nano MTA (A, E), Grey MTA (B, F), MTA Fillapex (C, G) and Control groups (D, H). Presence of moderate inflammatory cell infiltration (orange arrows) in (A, B, D) and severe (black arrow) in (C) at 7 days. Presence of mild inflammatory cell infiltration (E, F, H) and moderate/severe (G) at 30 days. (A-H) Hematoxylin-eosin staining, 100X.

Source: Authors.

**Figure 2:** Representative images of the subcutaneous tissue reactions in the Nano MTA (I, M), Grey MTA (J, N), MTA Fillapex (K, O) and Control groups (L, P). Absence of positive structures for von Kossa and absence of birefringent structures to polarized light (L, P) for the Control group at all time periods (90 days in the figure). All the other three materials showed Von Kossa-positive staining (I, J, K) and granulations birefringent to polarized light (M, N, O) near the tube opening at all time periods (90 days in the figure). (A-H) Hematoxylin-eosin staining, 100X; (I-L) staining according to the von Kossa technique, 100X; (M-P) polarized light visualization, 100X.

Source: Authors.
Mineral trioxide aggregate (MTA) is the primary hydraulic calcium silicate cement patented for endodontic applications and the most well known and most thoroughly investigated of all the hydraulic calcium cements available to date. It is a clinker-derived Portland cement composed of different phases, including tricalcium silicate, dicalcium silicate, tricalcium aluminate, tetracalcium aluminoferrite and calcium sulphate, as well as bismuth oxide as a radiopacifier (Josette Camilleri, 2014).

The placement of the materials into the subcutaneous tissue of rats is considered a standardized and valid test for biocompatibility (Olsson, Sliwkowski, & Langeland, 1981), analyzing the inflammatory tissue response through hematoxylin-eosin staining (Bueno et al., 2018), within time periods of the endorsed standard practices for biological evaluation of dental materials ("Recommended standard practices for biological evaluation of dental materials. Federation Dentaire International, Commission of Dental Materials, Instruments, Equipment and Therapeutics,” 1980), being used widely across the world. The polyethylene tubes delimit the dispersion of the material to a restricted area, enabling comparing the dispersion of the material in the tooth apex to the periapical tissues.

Biocompatible materials are not inert but should not cause an unacceptable physiological response in the host when compared to other biocompatible materials already documented (J. Camilleri, 2008). The results observed with Nano MTA were similar to the grey MTA and control groups. A moderate chronic inflammatory response at days 7 and 15, reducing over time. Positive von Kossa areas and birefringent structures under polarized light were also observed in all groups, except for the Control, showing that the material stimulated the formation of mineralized tissue in subcutaneous tissues of rats probably via the calcium carbonate formation from the calcium of the material and carbon dioxide from the surrounding tissue (Holland et al., 1999).

One possible reason for the adequate biological properties of the Nano MTA material containing is their high similarity to the conventional MTA composition, differing only in particle size. This change in particle size can play an important role in physical and chemical properties, leading to an increased surface area of powder that can reduce the setting time and increase the microhardness even at lower pH values after hydration. This fast setting prevents washout or dislodgement of MTA cement in clinical use. The bioactivity of these materials is due to the hydration of the calcium silicate leading to by-products formation. When in contact with the tissue, materials based on calcium silicate form calcium hydroxide and release calcium and hydroxyl ions promoting the increase of pH. The alkaline pH and the release of calcium ions initially promote a tissue inflammatory response, and later, these ions react with the carbon dioxide present in the tissue, giving rise to calcite crystals, which in turn are related to the decrease of inflammation and deposition of mineralized structures promoting repair (Cintra et al., 2017; Gomes-Filho et al., 2009; Holland et al., 1999).

Every new material needs to be tested prior to being launched to the market. The sample of the Nano MTA used in this study was donated by a researcher from the São Paulo State University (UNESP), Ilha Solteira, Brazil. As in any preclinical experimental study, the results of the present investigation should be read with caution since the methodology used has limitations to its application in humans. The use of animals allows investigating different materials under controlled laboratory conditions prior to their use in humans (Browne, 1994). Although these results do not reflect a complete analysis of the reactions that occur in the human conditions, they are significant for the preliminary assessment of the biocompatibility and biomineralization of the tested material.

Considering that Nano MTA presented a biocompatibility and biomineralization capability similar to grey MTA, it seems to be a promising alternative for root canal treatment due to alleged high chemical physical properties as a nanoparticulate material. However, more studies are required to reinforce its physical and biological characteristics.
4. Conclusion

Nano MTA exhibited biocompatibility and biomineralization ability comparable to grey MTA Angelus.

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References


