

Chemical composition, actividad larvicide, insecticide and repellent of essential oil

Aedes aegypti

Composição química, atividade larvicida, inseticida e repelente e larvicida de óleos essenciais frente ao *Aedes aegypti*

Composición química, actividad larvicia, insecticida y repelente de los aceites esenciales contra *Aedes aegypti*

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Antônia Laires da Silva Santos

ORCID: <https://orcid.org/0000-0001-6233-6173>
Federal University of Piauí, Brazil
E-mail: serialufpi@gmail.com

Felipe Pereira da Silva Santos

ORCID: <https://orcid.org/0000-0001-9079-952X>
Federal University of Piauí, Brazil
E-mail: felipe.p.santos1@hotmail.com

Alyandra de Sousa Nascimento

ORCID: <https://orcid.org/0000-0001-5059-7595>
Federal University of Piauí, Brazil
E-mail: alyandra.ufpi@gmail.com

Layana Karine Farias Lima

ORCID: <https://orcid.org/0000-0002-1431-2422>
Federal University of Piauí, Brazil
E-mail: layana_farias@hotmail.com

Lucas Mendes Feitosa Dias

ORCID: <https://orcid.org/0000-0002-8706-9945>
Federal University of Piauí, Brazil
E-mail: lucas.mendes1610@hotmail.com

Geovanna Trajano Oliveira da Silva
ORCID: <https://orcid.org/0000-0003-2549-6703>
Federal University of Piauí, Brazil
E-mail: geovannatrajano@gmail.com

Mahendra Rai

ORCID: <https://orcid.org/0000-0001-7645-9801>
University Sant. Gadge Baba Amravati, India
E-mail: indobraz77@gmail.com

Christiane Mendes Feitosa

ORCID: <https://orcid.org/0000-0001-8013-1761>
Federal University of Piauí, Brazil
E-mail: chistiane@ufpi.edu.br

Abstract

Dengue is transmitted by the mosquito *Aedes aegypti*, which is controlled by insecticides and repellents. The plant-based essential oils are used as excellent repellents. Therefore, the present review is essential for understanding the efficacy of essential oils against mosquitoes. The articles related to essential oils were searched from 2014 to 2022 in Pubmed, Science Direct, LILACS, Scielo, and Google Scholar using the following keywords-essential oil, *Aedes aegypti*, and repellents. A total of 280 plants were extracted for EOs from 33 families. The most plants belong to Lamiaceae (45.4%), followed by Myrtaceae (38.3%) and Rutaceae (28.4%). The metabolites in EOs with the maximum repellent/larvicidal activity include β - caryophyllene, α - pinene, 1,8 - cineol, linalool, and eugenol, the lethal concentrations ranged from 40 to 120 ppm. These results support the view that essential oils are promising in the formulation of repellents, larvicides, insecticides, and pesticides.

Keywords: *Aedes aegypti*; Insecticide; Essential oil; Larvicidal; Repellent.

Resumo

A dengue transmitida pelo mosquito *Aedes aegypti* é controlada por larvicida, inseticida e repelente. Os óleos essenciais à base de plantas são usados como excelentes repelentes. Portanto, a presente revisão é essencial para o entendimento da eficácia dos óleos essenciais contra os mosquitos. Os artigos relacionados a óleos essenciais foram

pesquisados de 2014 a 2022 no Pubmed, Science Direct, LILACS, Scielo e Google Scholar usando as seguintes palavras-chave: óleo essencial, *Aedes aegypti* e repelentes. Um total de 280 plantas foram extraídas para EOs de 33 famílias. A maioria das plantas pertence a Lamiaceae (45,4%), seguida por Myrtaceae (38,3%) e Rutaceae (28,4%). Os metabólitos em OE com atividade repelente / larvicida máxima incluem β-cariofileno, α-pineno, 1,8-cineol, linalol e eugenol, as concentrações letais variaram de 40 a 120 ppm. Esses resultados apóiam a visão de que os óleos essenciais são promissores na formulação de repelentes, larvicidas, inseticidas e pesticidas.

Palavras-chave: *Aedes aegypti*; Inseticida; Óleo essencial; Larvicida; Repelente.

Resumen

El dengue es transmitido por el mosquito *Aedes aegypti*, que es controlado por insecticidas y repelentes. Los aceites esenciales de origen vegetal se utilizan como excelentes repelentes. Por lo tanto, la presente revisión es esencial para comprender la eficacia de los aceites esenciales contra los mosquitos. Los artículos relacionados con los aceites esenciales se buscaron entre 2014 y 2022 en Pubmed, Science Direct, LILACS, Scielo y Google Scholar utilizando las siguientes palabras clave: aceite esencial, *Aedes aegypti* y repelentes. Se extrajeron un total de 280 plantas para OE de 33 familias. La mayoría de plantas pertenecen a Lamiaceae (45,4%), seguidas de Myrtaceae (38,3%) y Rutaceae (28,4%). Los metabolitos en los AE con la máxima actividad repelente / larvicida incluyen β-cariofileno, α-pineno, 1,8-cineol, linalol y eugenol; las concentraciones letales oscilaron entre 40 y 120 ppm. Estos resultados apoyan la opinión de que los aceites esenciales son prometedores en la formulación de repelentes, larvicidas, insecticidas y pesticidas.

Palabras clave: *Aedes aegypti*; Insecticida; Aceite esencial; Larvicida; Repelente.

1. Introduction

Aedes aegypti (Diptera: Culicidae) is the main vector responsible for diseases such as Dengue, Chikungunya and Zika. Such diseases have contributed to the increase in significant human morbidity and mortality in many countries with different economic, climatic and social characteristics. Mosquito control and personal protection are currently the most important measures to control the diseases caused. The use of larvicides, repellants and insecticides is a more practical and economical way to prevent the transmission of these diseases to human beings. Although synthetic insecticides and repellants are still the main means of protecting crops, the use of alternative methods has increased, due to the current need to overcome problems such as resistance and reducing the risks of environmental contamination caused by non-biodegradable synthetic products; one of these searches is based on plants (Pichersky & Gershenson, 2002; Drewes et al., 2006).

The use of substances extracted from plants (Essential Oils – EOs) with repellent and insecticidal action, originated in China, later expanding to the West. Due to the difference in volatility between the components of the liquid, distillation is the main process for separating these constituents, as it is based on the difference in composition of the constituents in the liquid and vapor phase in equilibrium (Almeida, 2005; Latyki, 2017).

During World War II some synthetic repellants were developed to protect humans from mosquito bites, with DEET (N,N-diethyl-m-toluamide) not only being a broad-looking repellent, but also the most effective and persistent on the skin. Herbal repellants, even with comparable or even better activities, tend to be short-lived and their effectiveness depends on their volatility, which makes synthetics more effective and more durable than natural products (Dethier, 1956; Isman, 2006; Nerio et al., 2010).

With 46,781 plant species, Brazil has the richest flora in the world (about 19% of the world's flora). In number of species, Legumes (2,144), Myrtaceae (1,038) and Rubiaceae (1,000) are the largest families, with about 750 and 500 species being also notable for Apocynaceae and Lamiaceae, respectively (Giulietti et al., 2005; Flora of Brazil, 2020). In recent years, these families have been intensively studied regarding their morphology, anatomy, cytogenetics, ecology and mainly phytochemistry, as they make a significant contribution to the drug industry and a vast number of products and extracts with high pharmacological and therapeutic activities (Figuereido et al., 2007).

Larvicidal, repellent and insecticide properties have been targets for the development of new products, as since 2008 Brazil has become the largest consumer of pesticides in the world, using around 700 thousand tons of synthetic products.

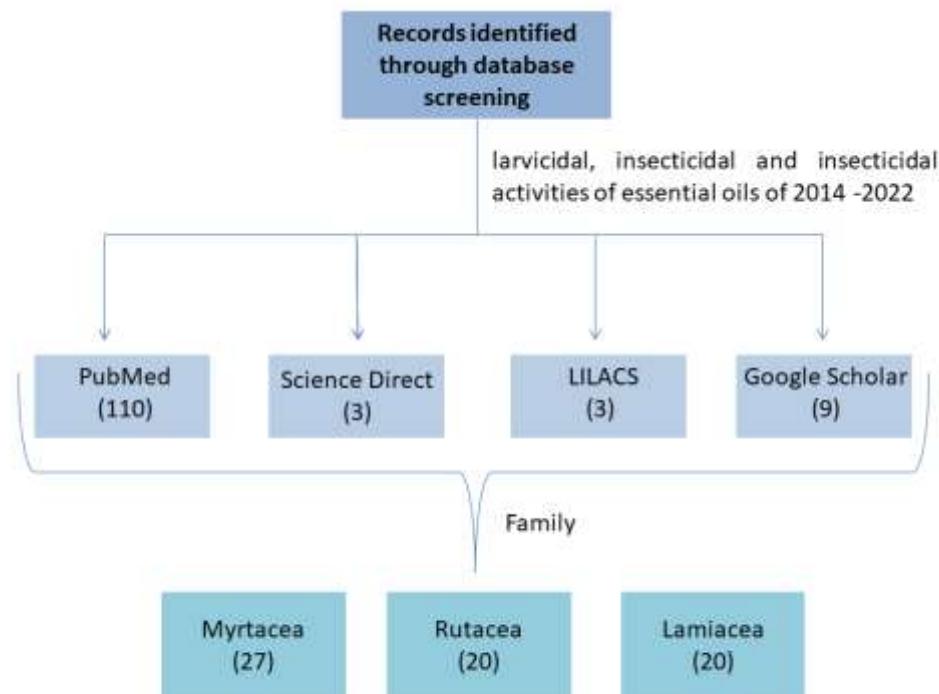
Significant restructuring in the industrial sector due to the growth in the number of active substances extracted from plants present in the market has contributed for this change to happen. Because nature has several substances that play a defensive role, among these substances we can mention: pyrethroids, extracted from chrysanthemum (*Chrysanthemum c.*) (Trev.), nicotine from *Nicotiana tabacum L.*, rotenone, extracted from *Derris sp.*, *Lonchocarpus sp.* and azadirachtin, isolated from *Azadirachta indica A. Juss.* Monoterpenes and sesquiterpernes also stand out (Ootani, 2010; Morais & Marinho, 2016).

Aiming at a better assessment of the larvicidal and repellent activities of substances extracted from EOs against the *Aedes aegypti* mosquito, this review focuses on the evaluation of the main sources of EOs and main compounds that have activities against larvae and mosquitoes in the period between 2014 to 2022.

2. Methodology

In this study, an integrative review was performed from 2014 to 2022 using Web-based databases, including PubMed, Science Direct, LILACS and Google Scholar using the following keywords: essential oil, *Aedes aegypti*, larvicidal, insecticidal and repellents (Figure 1 and Table 1).

Figure 1: Article search and selection strategy.



Source: Authors.

Table 1: Works selection.

Platform	Found works	Selected works
PubMed	110	56
Science Direct	3	3
LILACS	3	2
Google Scholar	9	6

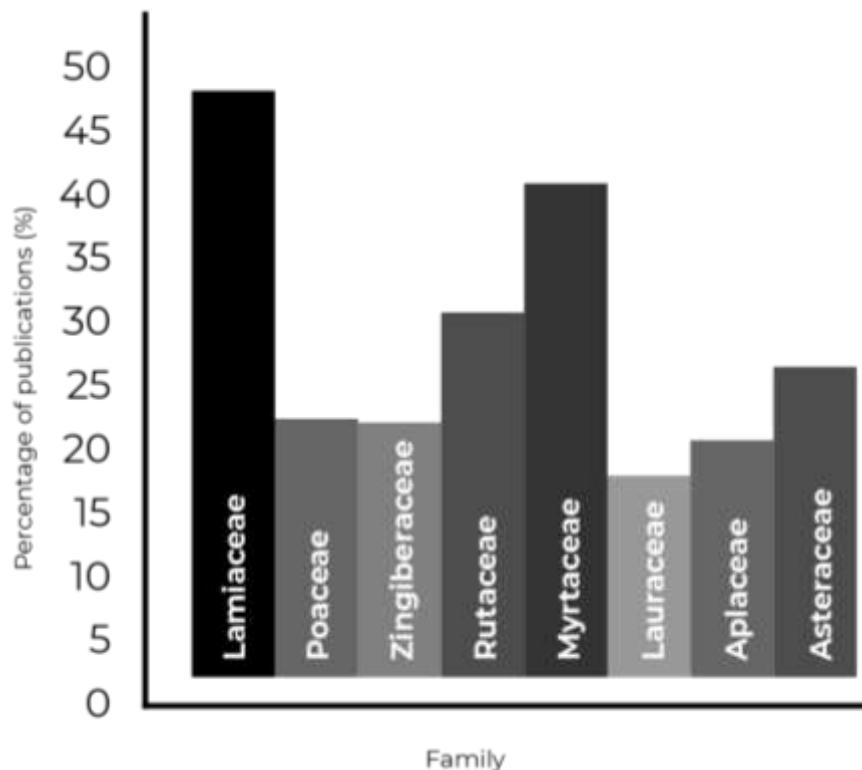
Source: Prepared by the research authors (2022).

3. Results and Discussion

3.1 Botanical Description

About 280 EOs extracted from 33 different families have shown repellency against *Aedes aegypti*. Among the EO-producing plants, some families such as Lamiaceae (45.4%), Myrtaceae (38.3%) and Rutaceae (28.4%) were widely studied (Graph 1).

Graph 1. Families with the highest number of published works between 2014 and 2022.



Source: Authors.

The Lamiaceae (Figure 2) family, also known as *Labiatae*, is composed of several plant species of economic and medicinal interest. It covers 258 genera and 7,193 species, which can be found in both tropical and temperate regions, and in Brazil, there are on average 23 genera and 232 native species (Carréra et al., 2015; Trindade et al., 2016). Because they have important biosynthesized compounds due to secondary metabolism, the Lamiaceae family is known for its biological activities already reported in the literature and for its use as a spice (Lima & Cardoso, 2007).

Figure 2. Species of the Lamiaceae family (A - *Mentha longifolia*; B - *Lamium purpureum*; C - *Scutellaria galericulata*; D - *Ajuga reptans* and E - *Ocimum basilicum*).



Source: Authors.

Among the species with the highest repellent, larvicidal and oviposition activity from 2014 to 2022 were: *Ocimum americanum* (Field alfava), *Salvia apiana*, *S. elegans*, *S. leucanta*, *S. officinalis*, *Ocimum basilicum* (basil), *O. americanum* (Majericium - white), *Nepeta cataria* (cat weed), *Dracocephalum heterophyllum*, *Hyssopus officinalis* (holy herb), *Origanum scabrum* (Oregano), *Lavendula angustifolia* (Lavender), *Mentha piperita* (Peppermint), *Rosmarinus officinalis* (Rosemary), *Thymus serpyllum* (Dandelion herb), *Dracocephalum ruyschiana*, *D. foetidum*, *D. ruyschiana*, *D. fruticosum*, *D. peregrinum*, *Nepeta rtanjensis*, *Ocimum sanctum* (Majerica - saint), *Plectranthus amboinicus* (Spearmint), *Lavandula angustifolia* (Lavender), *Origanum vulgare* (Oregano), *Vitex negundo* (Tree re chastity), *Stachys tmorea*, *Vitex trifolia* (Pepper), *Ocimum campechianum* (Basil - large), *Pogostemon cablin* (Patchouli) and *Mentha requienii*.

Among the species mentioned, the genera *Ocimum*, *Salvia* and *Origanum* presented higher activity against *Aedes aegypti* with lethal concentrations ranging from 6.2 to 80 ppm. Many plants of the Lamiaceae family produce OEs with insecticidal activity, the presence of many substances is related to this activity, acting as a larval growth inhibitor. Among them are γ -terpinene, α -terpinene, linalool, methyl-eugenol, eugenol, β -pinene, α -pinene, 1,8-cineol, and citronellol (Table 2). Such compounds may cause toxic interference with biochemical and physiological functions, thus acting as a repellent. The processes of absorption, inhalation, and ingestion of toxins can cause an adverse effect on insects (Lima & Cardoso, 2007; Simões et al., 2004).

Table 2. Substances and activities against *Aedes aegypti* reported in the literature.

SUBSTANCE	MOLECULAR FORMULA AND MASS (g.mol ⁻¹)	SPECIES	ACTIVITY	REFERENCE
γ -terpinene	C ₁₀ H ₁₆ (136.23)	<i>Petroselinum crispum</i> <i>Pleiospermum alatum</i> <i>Mentha requienii</i> <i>Vitex rotundifolia</i> <i>Crossostephium chinense</i> <i>Lippia gracilis</i> <i>Piper hispidinervum</i> <i>Piper aduncum</i>	Larvicide and adulticide	Intirach et al. (2016); Raj et al. (2017); Huang et al. (2019); Galvão et al. (2019); Silva et al. (2019).
Linalool	C ₁₀ H ₁₈ O (154.25)	<i>Curcuma longa</i> <i>Eucalyptus globulus</i> <i>Citrus aurantium</i> <i>Dracocephalum ruyschiana</i> <i>D. foetidum</i> <i>D. moldavica</i> <i>D. fruticosum</i> <i>D. peregrinum</i> <i>Syzygium aromaticum</i> <i>Citrus sinensis</i> <i>Piper augustum</i> <i>P. corrugatum</i> <i>P. curtispicum</i> <i>P. darienense</i> <i>P. grande</i> <i>Curcuma longa</i> <i>Eucalyptus globulus</i> <i>Citrus aurantium</i> <i>Rhanterium epapposum</i> <i>Origanum onites L</i> <i>Origanum onites L</i> <i>Xylopia laevigata</i> <i>Xylopia frutescens</i> <i>Lippia pedunculosa</i> <i>Mentha piperita</i> <i>Aristolochia trilobata</i> <i>Severinia monophylla</i> <i>Arisaema fargesii</i> <i>Piper hispidinervum</i> <i>Piper aduncum</i> <i>Illicium verum</i> <i>Pimenta dioica</i> <i>Myristica fragrans</i> <i>Mentha piperita</i> <i>Allium sativum</i> <i>Ocimum sanctum</i> <i>Callistemon linearis</i> <i>Eucalyptus maculata</i>	Insecticide, larvicide, adulticide and repellent	Auysawasdi et al. (2016); ÖZEK et al. (2016); Araujo et al., (2016); Santana et al. (2016); Demirci et a. (2017); Carroll et al. (2017); Demirci et al. (2017); Nascimento et al. (2017); Ramos et al. (2017); Silva et al., (2018); Satyal et al. (2019); Huang et al. (2019); Silva et al. (2019); Huang et al. (2020); Bailão, E, F. L. C., (2022).
Methyl-eugenol	C ₁₁ H ₁₄ O ₂ (178.23)		Insecticide, larvicide and adulticide	Voris et al. (2018); Sarma et al., (2019).
Eugenol	C ₁₀ H ₁₂ O ₂ (164.20)	<i>Plectranthus barbatus</i> <i>Dracocephalum ruyschiana</i> <i>D. foetidum</i> <i>D. moldavica</i> <i>D. fruticosum</i> <i>D. peregrinum</i> <i>Syzygium aromaticum</i> <i>Citrus sinensis</i> <i>Cymbopogon citratus</i> <i>Angelica archangelica</i> <i>Ferula assafoetida</i> <i>Citrus bergamia</i> <i>Cedrus deodora</i> <i>Cymbopogon nardus</i> <i>Syzygium aromaticum</i> <i>Eucalyptus globulus</i> <i>Allium sativum</i> <i>Pelargonium graveolens</i> <i>Citrus paradisi</i> <i>Dipterocarpus jourdainii</i> <i>Lavandula angustifolia</i> <i>Cymbopogon flexuosus</i> <i>Leptospermum scoparium</i>	Larvicide, adulticide and repellent	Govindarajan et al. (2016); Özak et al. (2016); Araujo et al. (2016); Soonwera et al. (2016); Affonso et al. (2018); Muturi et al. (2017). Tyagi et al. (2017); Voris et al. (2018); Huang et al. (2019); Sarma et al. (2019); Scalvenzi et al. (2019); Pandiyan et al. (2019); Harikarnpakdee et al. (2018); Lucia et al. (2020); Oliveira et al. (2020); Almadiy et al. (2020); Santos et al. (2020).

		<i>Citrus sinensis</i> <i>Origanum vulgare</i> <i>Cymbopogon martinii</i> <i>Mentha piperita</i> <i>Phoebe porosa</i> <i>Rosmarinus officinalis</i> <i>Melaleuca alternifolia</i> <i>Jasminum officinale</i> <i>Cinnamomum camphora</i> <i>Cedrus deodara</i> <i>Illicium verum</i> <i>Pimenta dioica</i> <i>Myristica fragrans</i> <i>Arisaema fargesii</i> <i>Mentha piperita</i> <i>Allium sativum</i> <i>Ocimum sanctum</i> <i>Callistemon linearis</i> <i>Eucalyptus maculata</i> <i>Ocimum campechianum</i> <i>Ocotea quixos</i> <i>Piper aduncum</i> <i>Illicium verum</i> <i>Trachyspermum ammi</i> <i>Ocimum gratissimum</i>	
β -pinene	$C_{10}H_{16}$ (136.23)	<i>Lippia microphylla</i> <i>Dracocephalum heterophyllum</i> <i>Hyssopus officinalis</i> <i>Eucalyptus globulus</i> <i>Citrus aurantium</i> <i>Curcuma longa</i> <i>Piper augustum</i> <i>P.corrugatum</i> <i>P.curispicum</i> <i>P.darienense</i> <i>P.grande</i> <i>P.hispidum</i> <i>P.jacquemontianum</i> <i>P.longispicum</i> <i>P.multiplinervium</i> <i>P.reticulatum</i> <i>P.trigonum</i> <i>Pinus kesiya</i> <i>Curcuma longa</i> <i>Eucalyptus globulus</i> <i>Citrus aurantium</i> <i>Boswellia ovalifoliolata</i> <i>Daucus carota</i>	Repellent, larvicide and insecticide
1,8-cineol	$C_{10}H_{18}O$ (154.25)	<i>Lippia microphylla</i> <i>Eugenia piauhiensis</i> <i>Myrcia erythroxylon</i> <i>Psidium myrsinoides</i> <i>Siparuna camporum</i> <i>Curcuma longa</i> <i>Croton jacobinensis</i> <i>Amomum subulatum</i> <i>Eucalyptus nitens</i> <i>Lippia gracilis</i> <i>Eucalyptus globulus</i> <i>Azadirachta indica</i> <i>Salvia officinalis</i> <i>Eucalyptus globulus</i>	Repellent, larvicide and insecticide

β -caryophyllene	$C_{15}H_{24}$ (204.35)	<i>Salvia apiana</i> <i>Salvia elegans</i> <i>Salvia leucantha</i> <i>Salvia officinalis</i> <i>Nigella sativa</i> <i>Murraya exótica</i> <i>Plectranthus barbatus</i> <i>Zingiber nimmonii</i> <i>Lantana montevidensis</i> <i>Syzygium lanceolatum</i> <i>Piper augustum</i> <i>P.corrugatum</i> <i>P.curtispicum</i> <i>P.darienense</i> <i>P.grande</i> <i>P.hispidum</i> <i>P.jacquemontianum</i> <i>P.longispicrum</i> <i>P.multiplinervium</i> <i>P.reticulatum</i> <i>P.trigonum</i> <i>Cymbopogon citratus</i> <i>Syzygium aromaticum</i> <i>Syzygium aromaticum L</i> <i>Xylopia laevigata</i> <i>Xylopia frutescens</i> <i>Lippia pedunculosa</i> <i>Severinia monophylla</i> <i>Salvia officinalis</i> <i>Plectranthus amboinicus</i> , <i>Mentha requienii</i> <i>Vitex rotundifolia</i> <i>Crossostephium chinense</i>	Larvicide and repellent	Ali et al. (2014); Raj et al. (2015); Krishnamoorthy et al. (2015); Govindarajan et al. (2016); Blythe et al. (2016); Benelli et al. (2016); Santana et al. (2016); Soonwera et al. (2016); Ffonso et al. (2017); Nascimento et al. (2017); Satyal et al., (2019); Morales et al., (2019); Huang et al., (2019); Borrero-landazabal et al., (2020).
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Source: Authors.

Myrtaceae (Figure 3) is a very complex family that accounts for about 1.32% of all known Angiosperms, divided into two large subfamilies: Myrtoideae and Leptospermoideae and has 142 genera and over 5,500 species. In Brazil, it is among the most important families in most vegetation formations (Lima et al., 2015; Stadnik et al., 2016).

Figure 3. Species of the Lamiaceae family (A - Eucalyptus globulus; B - Syzygium aromaticum; D - Eucalyptus globulus; E - Psidium guajava).



Source: Authors.

Its species have great economic potential because they are used as food, as the species of *Psidium guajava* L. of guava and *Eugenia uniflora* L. of "pitanga", consumed in the form of juice, sweets, jellies and ice cream. Some species such as *Eugenia sprengelii* and *Leptospermum scoparium* are used as ornamental plants and *Eucalyptus globulus* and *Myrciaria dubia* stand out in folk medicine for the treatment of influenza, nasal congestion and sinusitis (Lorenzi et al., 2006; Lorenzi & Matos, 2002; Lorenzi & Souza, 2001, Morais et al., 2014).

OEs extracted from species of the Myrtaceae family have a lethal concentration ranging from 7.9 to 106.9 ppm. Species include *Eucalyptus globulus* (Eucalyptus), *Melaleuca leucadendron* (Seven capotes), *Syzygium aromaticum* (Clove), *Eugenia piauhiensis* (Guava), *Eucalyptus grandis* (Pink Eucalyptus), *Eucalyptus camaldulensis* (Red Eucalyptus). *Myrcia sylvatica* (Cumatê), *Eucalyptus citriodora* (Eucalyptus lemon), *Syzygium lanceolatum* (Jambo), *Melaleuca quinquenervia* (myrtle), *Eucalyptus nitens* (Ecalypt robusta), *Leptospermum scoparium* (Erica), *Melaleuca alternifolia* Call (Tree) linearis (Bottle - brush) and *Eucalyptus maculata* (Spotted Eucalyptus). Eugenol, eugenyl acetate, and β-caryophyllene (Table 2) are the major compounds identified in EO_s of the Myrtaceae family.

The family is recognized for its opposite leaves, summertime inflorescences, flowers with stamens arranged in a ring and variable fruits, the Rutaceae family (Figure 4) consists of 150-162 genera and 1500-2096 species, are widely distributed throughout the humid, tropical and temperate regions in the world; and about 29 genera 182 species are found in Brazil (Melo & Zickel, 2004; Albuquerque, 1976; Pirani et al., 2002).

Figure 4. Species of the Lamiaceae family (A - *Citrus aurantifolia*; B - *Murraya exotica*; C - *Citrus paradisi*; D - *Citrus sinensis*; E - *Pleiospermum alatum*).



Source: Authors.

An important feature of this family reported by Pirani et al. (2002) is the development of strong aromatic oil-producing glands, which contributes to the genera of this family being responsible for several aromas, thus having a great economic importance. Several secondary metabolites are commonly found, such as alkaloids, especially those derived from

anthranilic acid, coumarins, lignans, flavonoids, terpenes and limonoids with a broad-spectrum of biological activities (Albarici et al., 2010).

Among the groups of plants with positive insecticidal activity are citrus fruits with a lethal concentration ranging from 43.7 to 538 ppm, among which are *Citrus aurantifolia* (Lemon), *C. hystrix* (combava), *Fortunella japonica* (Ximxim), *C. sinensis* (Orange), *Exotic Murraja* (Jasmin - Orange), *C. grandis* (Cimbo), *Amyris balsamifera* (Torch Wood), *C. aurantium* (Bitter Orange), *Swinglea glutinosa* (Lemon Swinglea), *C. bergamia* (Orange - Clove), *C. paradisi* (Grapefruit), *Zanthoxylum limonella* (Cat's hand), *Pleiospermum alatum* (lime orange), *Severinia monophylla* and *Zanthoxylum limoncello*. The genus Citrus is the most prominent in larvicidal activity, such property may be linked to the abundance of limonene (Table 2) present in the EO.

Pavela (2015) presented a bibliographic review from 2003 to 2014, in which the OEs of best lethal activity against mosquitoes are reported in five families: Lamiaceae, Cupressaceae, Rutaceae, Apiaceae, and Myrtaceae, confirming the results presented here.

3.2 Metabolites responsible for activity against *Aedes aegypti*

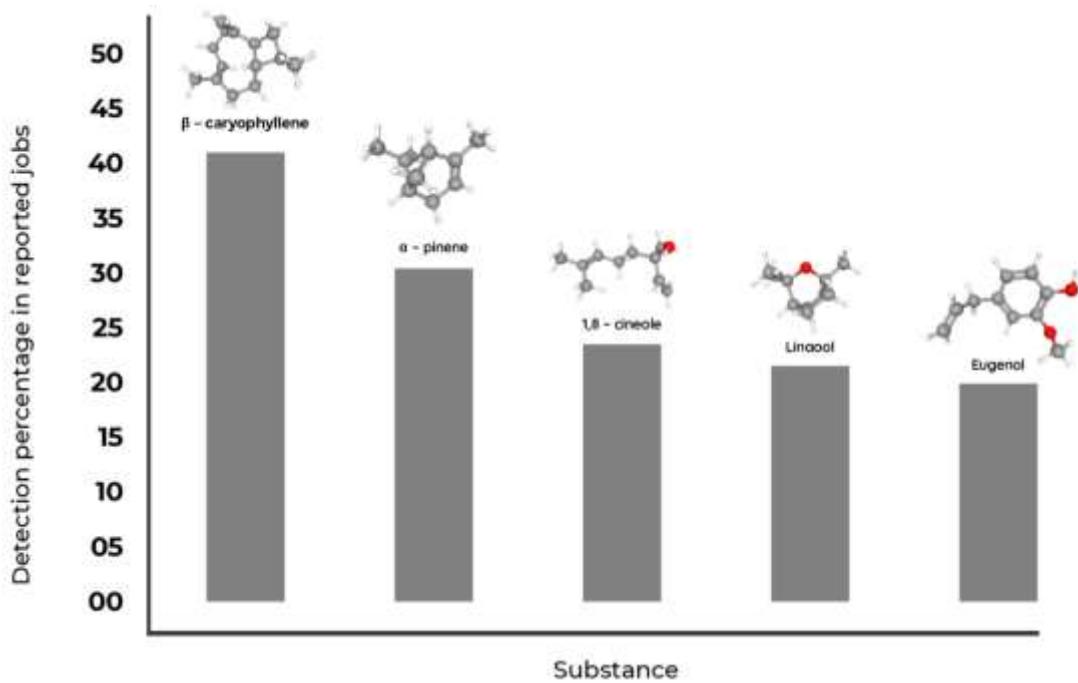
The secondary metabolites in plants are composed of hydrocarbons (terpenes and sesquiterpenes) and oxygenated compounds (alcohols, esters, ethers, aldehydes, ketones, lactones, phenols, and phenolic ethers) that have pharmaceutical properties and are responsible for the characteristic odor of plants. Such compounds present high vapor pressures under atmospheric conditions, which favors their significant release into the air (Pichersky & Gershenson, 2002; Edris, 2007).

The basic route of biosynthesis of these volatile constituents is conveniently treated in three stages. The first is the formation of the C₅ basic unit by two alternative pathways: the mevalonate pathway from acetyl-CoA located on the cytosol and is responsible for providing C₅ units for sesquiterpenes and the methylthrititol phosphate pathway. from pyruvate and glyceraldehyde-3-phosphate located on plastids, providing IPP (isopentyl diphosphate) and DMAPP (dimethylallyl diphosphate) for hemiterpene, monoterpene and diterpene biosynthesis (Dudareva et al., 2004; Dewick, 2009).

In the second phase of terpene biosynthesis, condensation reactions of prenyltransferases catalyzed IPP and DMAPP units form the precursors of monoterpenes (geranyl diphosphate - GPP), sesquiterpenes (farnesyl diphosphate - FPP) and diterpenes (geranylgeranyl diphosphate). And in the third stage of formation of volatile terpenes involves the conversion of the various prenyl diphosphates, DMAPP (C₅), GPP (C₁₀), FPP (C₁₅) and GGPP (C₂₀), to hemiterpenes (isoprene and 2-methyl-3-butene), 2-ol, monoterpenes, sesquiterpenes, and diterpenes, respectively. Terpene synthase enzymes are responsible for such reactions. Many terpene volatiles are direct products, but others are formed by transforming the starting products by oxidation, dehydrogenation, acylation, and other types of reactions (Edris, 2007; Dudareva et al., 2004).

Biosynthesis normally occurs in plant tissue epidermal cells, from which compounds can escape into the atmosphere or rhizosphere after being synthesized, so a wide variety of secondary metabolites have a number of effects such as herbivore and pathogen repulsion (Dudareva et al., 2004). These include β - caryophyllene; α - pinene; 1,8 - cineol; linalool and eugenol (Table 2) as the main compound that presents larvicidal and repellent activity against *Aedes aegypti* in the last seven years (Graph 2).

Graph 2. Percentage of detection of chemical substances present in essential oils.



Source: Authors.

The 42.32% of the published reports provide evidence that β - caryophyllene is the most common bicyclic sesquiterpene found in OEs of various plants, such as *Erechtites hieraciifolius*, *Psidium myrsinifolium* and *Plectranthus amboinicus*. Krishnamoorthy et al. (2015) reported that the significant larvicidal activity of *Murraya exotica* essential oil may be due to the presence of important chemical constituents such as β - humulene (40.62%), benzyl benzoate (23.96%) and β -caryophyllene (7.05%), presenting an LC₅₀ of 74.7 ppm against the 3rd and 4th stage larvae after 12h of exposure. The same is true for *Plectranthus amboinicus* EO, LC₅₀ values for β - caryophyllene against *Aedes aegypti* was 74.46 ppm, a better value than for *Culex quinquefasciatus* mosquito which was 146.58 ppm (Huang et al., 2019). Huang et al. (2019) reported values LC₅₀ of larvicidal activity of β - caryophyllene against species of *Aedes aegypti*, *Aedes albopictus*, *Anopheles subpictus*, *Culex pipiens pallens*, *Culex tritaeniorhynchus* and *Ochlerotatus togoi*. The activity values LC₅₀ ranged from 38.58. ppm to 97.90 ppm, and for *Aedes aegypti* β - caryophyllene presented the best activity (Hung et al., 2019).

Hung et al. (2019) mentioned the presence of α - pinene in *Erechtites hieraciifolius* with 14.5% abundance and in *Erechtites valerianifolius* with 30.2% abundance, 10. 6 and 12.5 ppm for the death of 50% of the larvae, such values suggest that α - pinene has a more significant effect against *Aedes aegypti*. Ali et al. (2015) performed larvicidal activity against *A. egypti* and *A. quadrimaculatus* in *Salvia* species that did not show significant activity against larvae due to the low abundance of α - pinene. In *Angelica dahurica* and *A. pubescens* species presented by Tabanca et al. (2014) the abundance of α - pinene was 46.3 and 37.6%, but it was 1-dodecanol and 1-tridecanol that showed larvicidal activity. *Pinus kesiya* EO demonstrated 21.8% α - pinene with the lethal death concentration of 50% of the 57 ppm larvae (Govindarajan et al., 2016). Studies published from 2014 to 2019 related good larvicidal activity, ranging from 2.3 to 86 ppm for oils with α - pinene in their composition.

The constituent of several plants OEs used in folk medicine, 1,8 - cineol acts on the respiratory, cardiovascular and bone systems. *Lippia microphylla* has a LC₅₀ of 75.6 ppm against *A. aegypti*, which can be applied to the presence of 1,8 - cineol in its chemical composition (Simões et al., 2015). The values of 43.8 and 52.8 ppm are presented by Costa et al. (2017) and Costa et al. (2018) with 1,8 - cineol as the major compound in OEs. Linalool, even present in many published studies,

presents low abundance and low activity compared to *A. aegypti*, Silva et al. (2016) found a high linalol abundance of around 51.8% in *M. piperita* EO more than low potential with the LC₅₀ of 367.6 ppm. For eugenol, it is observed that the articles with the best relative abundance are *Syzygium* species of the Myrtaceae family, the relative abundance found is around 50% and an average lethal concentration activity of 60 ppm (Pandiyan et al., 2019). An important factor to be considered is the presence of synergistic interactions between the components, thus increasing bioavailability or reducing doses for activity evaluation.

4. Conclusion

The essential oils of plants of the Lamiaceae, Myrtaceae, and Rutaceae families have shown great potential for repellent and larvicidal activity against various arthropod species, especially against *A. aegypti*. Most of the metabolites with the best repellent/larvicidal activity were β - caryophyllene, α - pinene, 1,8 - cineol, linalol, and eugenol. These products when mixed presented excellent repellent potential, obtaining lower values than those found for DEET.

Carrying out this type of review is extremely important, as it makes it possible to identify what is being researched and discussed on a particular topic. After carrying out this review, a variety of compounds extracted from plants can be seen that have shown potential activity against the *Aedes aegypti* mosquito, becoming promising in this regard, as they are ecologically effective and generally have low toxicity. Understanding how and which compounds and plant families have shown such significant activities contributes to the production of drugs that prevent the transmission and spread of dengue in low-income countries.

References

- Affonso, R. S., Lima, J. A., Lessa, B. M., Caetano, J. V., Obara, M. T., Nóbrega, A. B., Nepovimova, E., Musilek, K., Kuca, K., Slana, G. B. C. A., & França, T. C. (2018). Quantification through TLC-densitometric analysis, repellency and anticholinesterase activity of the homemade extract of Indian cloves. *Biomedical Chromatography*, 32 (2), 1-8. <https://doi.org/10.1002/bmc.4096>
- Albarici, T. R., Vieira, P. C., Fernandes, J. B., & Silva, M. F. D. G. F. (2010). Coumarin and alkaloids of *Rauia resinosa* (Rutaceae); Cumarinas e alcaloides de *Rauia resinosa* (Rutaceae). *Química Nova (Online)*, 33 (10), 2130-2134. <https://doi.org/10.1590/S0100-40422010001000024>
- Albuquerque, B. W. (1976). Revisão taxonômica das Rutaceae do Estado do Amazonas. *Acta amazônica*, 6, 5-67. <https://doi.org/10.1590/1809-43921976063s005>
- Ali, A., Tabanca, N., Demirci, B., Blythe, E. K., Ali, Z., Baser, K. H. C., Khan, & Ikhlas A. (2014) Chemical Composition and Biological Activity of Four *Salvia* Essential Oils and Individual Compounds against Two Species of Mosquitoes. *Journal Agriculture Food Chemistre*, 63 (2), 447-456. <https://doi.org/10.1021/jf504976f>
- Ali, A., Tabanca, N., Ozek, G., Ozek, T., Aytac, Z., Bernier, U. R., Agramonte, N. M., Husnu, B. L. K & Khan, I. A. (2015). Essential oils of *Echinophora lamondiana* (Apiales: Umbelliferae): A relationship between chemical profile and biting deterrence and larvicidal activity against mosquitoes (Diptera: Culicidae). *Journal of medical entomology*, 52 (1), 93-100. <https://doi.org/10.1093/jme/tju014>
- Almadiy, A. A. (2020). Chemical composition, insecticidal and biochemical effects of two plant oils and their major fractions against *Aedes aegypti*, the common vector of dengue fever. *Heliyon*, 6 (9), 1-9. <https://doi.org/10.1016/j.heliyon.2020.e04915>
- Almeida, A. O. (2005). *Dissertação (Mestrado) – Dissertação de Mestrado em Gerenciamento e Tecnologias Ambientais no Processo Produtivo*. UFBA.
- Araujo, A. F. D. O., Ribeiro-Paes, J. T., Deus, J. T. D., Cavalcanti, S. C. D. H., Nunes, R. D. S., Alves, P. B., & Macoris, M. D. L. D. G. (2016). Larvicidal activity of *Syzygium aromaticum* (L.) Merr and *Citrus sinensis* (L.) Osbeck essential oils and their antagonistic effects with temephos in resistant populations of *Aedes aegypti*. *Memórias do Instituto Oswaldo Cruz*, 111, 443-449. <https://doi.org/10.1590/0074-02760160075>
- Auyswasdi, N., Chuntranuluck, S., Phasomkusolsil, S., & Keeratinijakal, V. (2016). Improving the effectiveness of three essential oils against *Aedes aegypti* (Linn.) and *Anopheles dirus* (Peyton and Harrison). *Parasitology research*, 115 (1), 99-106. <https://doi.org/10.1007/s00436-015-4725-3>
- Bailão, E. F. L. C., Pereira, D. G., Romano, C. A., Santana Paz, A. T., Silva, T. M., Paula, J. R., Gomes, C. M., & Borges, L. L. (2022). Larvicidal effect of the *Citrus limettioides* peel essential oil on *Aedes aegypti*. *South African Journal of Botany*, 144, 257-260. <https://doi.org/10.1016/j.sajb.2021.09.013>
- Bemelli, G., Rajeswary, M., & Govindarajan, M. (2018). Towards green oviposition deterrents? Effectiveness of *Syzygium lanceolatum* (Myrtaceae) essential oil against six mosquito vectors and impact on four aquatic biological control agents. *Environ Sci Pollut Res.*, 25 (11), 10218-10227. <https://doi.org/10.1007/s11356-016-8146-3>
- Benelli, G., Rajeswary, M., & Govindarajan, M. (2016). Rumo aos impedimentos de oviposição verde? Efetividade do óleo essencial de *Syzygium lanceolatum* (Myrtaceae) contra seis mosquitos vetores e impacto em quatro agentes de controle biológico aquático. *Environmental Science and Pollution Research*, 1, 1-8. <https://doi.org/10.1007/s11356-016-8146-3>

Benelli, G., Rajeswary, M., Vijayan, P., Senthilmurugan, S., Alharbi, N.S., Kadaikunnan, S., Khaled, J. M., & Govindarajan, M. (2018). Óleo essencial de Boswellia ovalifoliolata (Burseraceae) como larvicida ecologicamente correto? Toxicidade contra seis mosquitos vetores de importância para a saúde pública, peixes mosquitos não visados, nadadores de costas e insetos aquáticos. Ambiente. *Environmental Science and Pollution Research*, 25 (11), 10264-10271. <https://doi.org/10.1007/s11356-017-8820-0>

Blythe, E. K., Tabanca, N., Demirci, B., Tsikolia, M., Bloomquist, J. R., & Bernier, U. R. (2016). *Lantana montevidensis* Essential Oil: Chemical Composition and Mosquito Repellent Activity against *Aedes aegypti*. *Nat Prod Commun.* 11 (11), 1713-1716.

Borrero-Landazabal, M. A., Duque, J. E., & Mendez-Sanchez, S. C. (2020). Model to design insecticides against *Aedes aegypti* using in silico and in vivo analysis of different pharmacological targets. *Comparative Biochemistry and Physiology Part C: Toxicology & Pharmacology*, 229, 1-9. <https://doi.org/10.1016/j.cbpc.2019.108664>

Carréa, J. C., Ilku-Borges, F., Rodrigues, S. T., Rodrigues, A. D. C. (2015). *Espécies de Lamiaceae (Labiatae) mais utilizadas na medicina popular do estado do Pará*. 66º Congresso Nacional de Botânica.

Carroll, J. F., Demirci, B., Kramer, M., Bernier, U. R., Agramonte, N. M., Baser, K. H. C., & Tabanca, N. (2017). Repellency of the *Origanum onites* L. essential oil and constituents to the lone star tick and yellow fever mosquito. *Natural product research*, 31 (18), 2192-2197. <https://doi.org/10.1080/14786419.2017.1280485>

Costa, A. A., Gonzalez, P. V., Harburguer, L. V., & Masuh, H. M. (2018). Effects of temephos, permethrin, and Eucalyptus nitens essential oil on survival and swimming behavior of *Aedes aegypti* and *Anopheles pseudopunctipennis* (Diptera: Culicidae) larvae. *Journal of medical entomology*, 55 (5), 1098-1104. <https://doi.org/10.1093/jme/tjy086>

Costa, A. A., Naspi, C. V., Lucia, A., & Masuh, H. M. (2017). Repellent and larvicidal activity of the essential oil from *Eucalyptus nitens* against *Aedes aegypti* and *Aedes albopictus* (Diptera: Culicidae). *Journal of medical Entomology*, 54(3), 670-676. <https://doi.org/10.1093/jme/tjw222>

Demirci, B., Yusufoglu, H. S., Tabanca, N., Temel, H. E., Bernier, U. R., Agramonte, N. M., Alqasoumi, S. I., Al-Rehaily, A. J., Baser, K. H. C., & Demirci, F. (2017). *Rhanterium epapposum* Oliv. essential oil: Chemical composition and antimicrobial, insect-repellent and anticholinesterase activities. *Saudi Pharmaceutical Journal*, 25(5), 703-708. <https://doi.org/10.1016/j.jsps.2016.10.009>

Dethier, V. G. (1956). *Repellents*. Annu. Rev. Entomol.

Dewick, P. M. (2009). *Medicinal Natural Products: A Biosynthetic Approach*. Wiley.

Dias, C. N., Alves, L. P. L., Rodrigues, K. A. F., Brito, M. C. A., Rosa, C. S., Amaral, F. M. M., Monteiro, O. S., Andrade, E. H. A., Maia, J. G. S., & Moraes, D. F. C. (2015). Chemical Composition and Larvicidal Activity of Essential Oils Extracted from Brazilian Legal Amazon Plants against *Aedes aegypti* L. (Diptera: Culicidae). *Evidence-Based Complementary and Alternative Medicine*. 2005, 1-8. <http://dx.doi.org/10.1155/2015/490765>

Drewes, S. E., Mudau, K. E., Van Vuuren, S. F., & Viljoen, A. M. (2006). Antimicrobial monomeric and dimeric diterpenes from the leaves of *Helichrysum tenax* var *tenax*. *Phytochemistry*, 67 (7), 716-722. <https://doi.org/10.1016/j.phytochem.2005.12.015>

Dudareva, N., Pichersky, E., & Gershenzon, J. (2004). Biochemistry of plant volatiles. *Plant physiology*, 135 (4), 1893-1902. <https://doi.org/10.1104/pp.104.049981>

Edris, A. E. (2007). Pharmaceutical and therapeutic potentials of essential oils and their individual volatile constituents: a review. *Phytotherapy Research*, 21(4), 308-323. <https://doi.org/10.1002/ptr.2072>

Figueiredo, A. C., Barroso, J. G., & Pedro, L. G. (2007) *Potencialidades e aplicações das Plantas aromáticas e Medicinais*. ULisboa.

Flora do Brasil, Rio de Janeiro (2020). <http://floradobrasil.jbrj.gov.br>

Galvão, J. G., Cerpe, P., Santos, D. A., Gonsalves, J. K., Santos, A. J., Nunes, R. K., Lira, A. A., Alves, P. B., Corte, R. L., Branco, A. F., Silva, G. F., Cavalcanti, S. C., & Nunes, R. S. (2019). *Lippia gracilis* essential oil in β-cyclodextrin inclusion complexes: an environmentally safe formulation to control *Aedes aegypti* larvae. *Pest management science*, 75 (2), 452-459. <https://doi.org/10.1002/ps.5138>

Giulietti, A. M., Harley, R. M., DE Queiroz, L. P., Wanderley, M. DAS G. L., Den Berg, C. V. (2005). Biodiversidade e conservação das plantas no Brasil. *Megadiversidade*, 1 (1), 53-61.

Govindarajan, M., & Benelli, G. (2016). α-Humulene and β-elemene from *Syzygium zeylanicum* (Myrtaceae) essential oil: highly effective and eco-friendly larvicides against *Anopheles subpictus*, *Aedes albopictus*, and *Culex tritaeniorhynchus* (Diptera: Culicidae). *Parasitology research*, 115 (7), 2771-2778. <https://doi.org/10.1007/s00436-016-5025-2>

Govindarajan, M., Kadaikunnan, S., Alharbi, N. S., & Benelli, G. (2016). Acute toxicity and repellent activity of the *Origanum scabrum* Boiss. & Heldr. (Lamiaceae) essential oil against four mosquito vectors of public health importance and its biosafety on non-target aquatic organisms. *Environmental Science and Pollution Research*, 23 (22), 23228-23238. <https://doi.org/10.1007/s11356-016-7568-2>

Govindarajan, M., Rajeswary, M., Arivoli, S., & Tennyson, S., (2016). Benelli, B. Larvicidal and repellent potential of *Zingiber nimmonii* (J. Graham) Dalzell (Zingiberaceae) essential oil: an eco-friendly tool against malaria, dengue, and lymphatic filariasis mosquito vectors?. *Parasitol Res.* 115 (5), 1807-1816. <https://doi.org/10.1007/s00436-016-4920-x>

Govindarajan, M., Rajeswary, M., Hoti, S. L., Bhattacharyya, A., & Benelli, G. (2016). Eugenol, α-pinene and β-caryophyllene from *Plectranthus barbatus* essential oil as eco-friendly larvicides against malaria, dengue and *Japanese encephalitis* mosquito vectors. *Parasitology research*, 115 (2), 807-815. <https://doi.org/10.1007/s00436-015-4809-0>

Govindarajan, M., Rajeswary, M., Senthilmurugan, S., Vijayan, P., Alharbi, N. S., Kadaikunnan, S., Khaled, M., & Benelli, G. (2018). Larvicidal activity of the essential oil from *Amomum subulatum* Roxb.(Zingiberaceae) against *Anopheles subpictus*, *Aedes albopictus* and *Culex tritaeniorhynchus* (Diptera:

Culicidae), and non-target impact on four mosquito natural enemies. *Physiological and Molecular Plant Pathology*, 101, 219-224. <https://doi.org/10.1016/j.pmp.2017.01.003>

Harikarnpakdee, S., & Chuchote, C. (2018). Oviposition Deterrent Efficacy and Characteristics of a Botanical Natural Product, *Ocimum gratissimum* (L.) Oil-Alginate Beads, against *Aedes aegypti* (L.). *The Scientific World Journal*, 2018, 1-9. <https://doi.org/10.1155/2018/3127214>

Huang, H. T., Lin, C. C., Kuo, T. C., Chen, S. J., & Huang, R. N. (2019). Phytochemical composition and larvicidal activity of essential oils from herbal plants. *Planta*, 250 (1), 59-68. <https://doi.org/10.1007/s00425-019-03147-w>

Huang, H. T., Lin, C. C., Kuo, T. C., Chen, S. J., & Huang, R. N. (2019). Phytochemical composition and larvicidal activity of essential oils from herbal plants. *Planta*, 250 (1), 59-68. <https://doi.org/10.1007/s00425-019-03147-w>

Huang, Y., Lin, M., Jia, M., Hu, J., & Zhu, L. (2020). Chemical composition and larvicidal activity against Aedes mosquitoes of essential oils from Arisaema fargesii. *Pest management science*, 76 (2), 534-542. <https://doi.org/10.1002/ps.5542>

Hung, N. H., Satyal, P., Hieu, H. V., Chuong, N. T. H., Dai, D. N., Huong, L. T., Thai, T. A., & Setzer, W. N. (2019). Mosquito larvicidal activity of the essential oils of Erechites species growing wild in Vietnam. *Insects*, 10 (2), 47. <https://doi.org/10.3390/insets10020047>

Intirach, J., Junkum, A., Lumjuan, N., Chaithong, U., Jitpakdi, A., Riyong, D., Wannasan, U., Champakaw, D., Muangmoon, R., Chansang, U., & Pitasawat, B. (2016). Antimosquito property of Petroselinum crispum (Umbelliferae) against the pyrethroid resistant and susceptible strains of *Aedes aegypti* (Diptera: Culicidae). *Environmental Science and Pollution Research*, 23 (23), 23994-24008. <https://doi.org/10.1007/s11356-016-7651-8>

Isman, M. B. (2006). Botanical insecticides, deterrents, and repellents in modern agriculture and an increasingly regulated world. *Annu. Rev. Entomol.*, 51, 45-66. <https://doi.org/10.1146/annurev.ento.51.110104.151146>

Krishnamoorthy, S., Chandrasekaran, M., RAJ, G. A., Jayaraman, M., & Venkatesal, V. (2015). Identification of chemical constituents and larvicidal activity of essential oil from *Murraya exotica* L. (Rutaceae) against *Aedes aegypti*, *Anopheles stephensi* and *Culex quinquefasciatus* (Diptera: Culicidae). *Parasitol Res.* 114 (5), 1838-1845. <https://doi.org/10.1007/s00436-015-4370-x>.

Latyki, B. L. (2017). *Trabalho de Conclusão de Curso (TCC)*. UFTPAs.

Lima, D. F., Caddah, M. K., & Goldenberg, R. (2015). A família Myrtaceae na Ilha do Mel, Paranaguá, Estado do Paraná, Brasil. *Hoehnea*, 42 (3), 497-519. <https://doi.org/10.1590/2236-8906-68/2014>

Lima, R. K., & Cardoso, M. G. (2007). Família Lamiaceae: Importantes Óleos Essenciais com Ação Biológica e Antioxidante. *Revista Fitos*, 3 (3), 14-24.

Lorenzi, H., & Matos, F. J. A. (2002). *Plantas Medicinais no Brasil*. Instituto Plantarum.

Lorenzi, H., & Souza, H. M. (2001). *Plantas Ornamentais no Brasil*. Instituto Plantarum.

Lorenzi, H., Bacher, L., Lacerda, M., & Sartori, S. (2006). *Frutas Brasileiras e Exóticas Cultivadas*. Instituto Plantarum.

Lucia, A., Toloza, A. C., Fanucce, M., Fernández-Peña, L., Ortega, F., Rubio, R. G., Coviella, C., & Guzmán, E. (2020). Nanoemulsions based on thymol-eugenol mixtures: characterization, stability and larvicidal activity against *Aedes aegypti*. *Bulletin of Insectology*, 73 (1), 153-160.

Melo, M. D. F. F., & Zickel, C. S. (2004). Os gêneros *Zanthoxylum* L. e *Esenbeckia* Kunth (Rutaceae) no Estado de Pernambuco, Brasil. *Acta Botanica Brasiliensis*, 18, 73-90. <https://doi.org/10.1590/S0102-33062004000100007>

Morais, L. A. S., & Marinho-Prado, J. S. (2016). *Defensivos agrícolas naturais: Uso e perspectivas*. Embrapa.

Morais, L., Conceição, G., & Nascimento, J. (2014). Família Myrtaceae: Análise morfológica e distribuição geográfica de uma coleção botânica. *Agrarian Academy*, 1 (01), 318-346. https://doi.org/10.18677/Agrarian_Academy_2014_018

Morales, R. M. C., Otero, A. L. C., Sanchez, S. C. M., Silva, M. A. N., Stashenko, E. E., & Duque, J. E. (2019). Mitochondrial affection, DNA damage and AChE inhibition induced by *Salvia officinalis* essential oil on *Aedes aegypti* larvae. *Comparative Biochemistry and Physiology Part C: Toxicology & Pharmacology*, 221, 29-37. <https://doi.org/10.1016/j.cbpc.2019.03.006>

Muturi, E. J., Doll, K., Ramirez, J. L., & Rooney, A. P. (2019). Bioactivity of Wild Carrot (*Daucus carota*, Apiaceae) Essential Oil Against Mosquito Larvae. *Journal of Medical Entomology*, 56, 784–789. <https://doi.org/10.1093/jme/tjy226>

Muturi, E. J., Ramirez, J. L., Doll, K. M., & Bowman, M. J. (2017). Combined toxicity of three essential oils against *Aedes aegypti* (Diptera: Culicidae) larvae. *Journal of Medical Entomology*, 54 (6), 1684-1691. <https://doi.org/10.1093/jme/tjx168>

Nascimento, A. M. D., Maia, T. D. S., Soares, T. E. S., Menezes, L. R. A., Scher, R., Costa, E. V., Cavalcanti, S. C. H., & La Corte, R. (2017). Repellency and larvicidal activity of essential oils from *Xylopia laevigata*, *Xylopia frutescens*, *Lippia pedunculosa*, and their individual compounds against *Aedes aegypti* Linnaeus. *Neotropical entomology*, 46 (2), 223-230. <https://doi.org/10.1007/s13744-016-0457-z>

Nerio, L. S., Olivero-Verbel, J., & Stashenko, E. (2010). Repellent activity of essential oils: a review. *Bioresource technology*, 101(1), 372-378. <https://doi.org/10.1016/j.biortech.2009.07.048>

Oliveira, A. D. N., Oliveira, D. T., Angelica, R. S., Andrade, E. H. D. A., da Silva, J. K. D. R., Rocha Filho, G. N. D., Coral, N., Pires, L. H. O., Luque, R., & Nascimento, L. A. S. (2020). Efficient esterification of eugenol using a microwave-activated waste kaolin. *Reaction Kinetics, Mechanisms and Catalysis*, 130, 633-653. <https://doi.org/10.1007/s11144-020-01797-6>

Ootani, M. A. (2010). Dissertação de Mestrado. UFT.

- Özek, G., Tabanca, N., Radwan, M. M., Shatar, S., Altantsetseg, A., Baatar, D., Baser, K. H. C., Becnel, J. J., & Özak, T. (2016). Preparative capillary GC for characterization of five *Dracocephalum* essential oils from Mongolia, and their mosquito larvicidal activity. *Natural product communications*, 11 (10), 1541-1544.
- Pandiyar, G. N., Mathew, N., & Munusamy, S. (2019). Larvicidal activity of selected essential oil in synergized combinations against *Aedes aegypti*. *Ecotoxicology and environmental safety*, 174, 549-556. <https://doi.org/10.1016/j.ecoenv.2019.03.019>
- Pavela, R. (2015). Essential oils for the development of eco-friendly mosquito larvicides: a review. *Industrial crops and products*, 76, 174-187. <https://doi.org/10.1016/j.indcrop.2015.06.050>
- Pichersky, E., & Gershenson, J. (2002). The formation and function of plant volatiles: perfumes for pollinator attraction and defense. *Current opinion in plant biology*, 5 (3), 237-243. [https://doi.org/10.1016/S1369-5266\(02\)00251-0](https://doi.org/10.1016/S1369-5266(02)00251-0)
- Pinto, C. C. C., de Menezes, J. E. S., Siqueira, S. M. C., Melo, D. S., Feitosa, C. R., & Santos, H. S. (2016). Chemical Composition and larvicidal activity against *Aedes aegypti* of essential oils from *Croton jacobinensis* Baill. *Boletín Latinoamericano y del Caribe de Plantas Medicinales y Aromáticas*, 15 (2), 122-127.
- Pirani, J. R., Wanderley, M. G. L., Shepherd, G. J., Giulietti, A. M., Melhem, T. S., Bittrich, V., & Kameyama, C. (2002). *Flora Fanerogâmica do Estado de São Paulo*. Instituto de Botânica.
- Raj, G. A., Chandrasekaran, M., Krishnamoorthy, S., Jayaraman, M., & Venkatesalu, V. (2015). Phytochemical profile and larvicidal properties of seed essential oil from *Nigella sativa* L. (Ranunculaceae), against *Aedes aegypti*, *Anopheles stephensi*, and *Culex quinquefasciatus* (Diptera: Culicidae). Parasitol Res, 114 (9), 3385-3391. <https://doi.org/10.1007/s00436-015-4563-3>
- Raj, G. A., Chandrasekaran, M., Venkatesalu, V., & Jegan, S. (2017). Phytochemical composition and larvicidal activity of essential oil from the leaves of *pleiospermum alatum* (wall. Ex wt. & arn) swingle against *Aedes aegypti*, *Anopheles stephensi* and *Culex quinquefasciatus* (Diptera: culicidae). 1 (N), 1-4.
- Ramos, R. S., Rodrigues, A. B. L., Farias, A. L. F., Simões, R. C., Pinheiro, M. T., Ferreira, R. M. D. A., Barbosa, L. M. C., Souto, R. N. P., Fernandes, J. B., Santos, L. S. & Almeida, S. S. M. D. S. (2017). Chemical composition and in vitro antioxidant, cytotoxic, antimicrobial, and larvicidal activities of the essential oil of *Mentha piperita* L.(Lamiaceae). *The Scientific World Journal*, 2017, 1-8. <https://doi.org/10.1155/2017/4927214>
- Santana, A. I., Vila, R., Cañigueral, S., & Gupta, M. P. (2016). Chemical composition and biological activity of essential oils from different species of *Piper* from Panama. *Planta medica*, 82 (11/12), 986-991. <https://doi.org/10.1055/s-0042-108060>
- Santos, A. J., Pina, L. T., Galvão, J. G., Trindade, G. G., Nunes, R. K., Santos, J. S., Santos, C. P., Gonsalves, J. K. M. C. Lira, A. A. M., Cavalcanti, S. C. H. Santos, R. L. C. Sarmento, V. H. V. & Nunes, R. S. (2020). Clay/PVP nanocomposites enriched with *Syzygium aromaticum* essential oil as a safe formulation against *Aedes aegypti* larvae. *Applied Clay Science*, 185, 1-7. <https://doi.org/10.1016/j.clay.2019.105394>
- Sarma, R., Adhikari, K., Mahanta, S., & Khanikor, B. (2019). Combinations of plant essential oil based terpene compounds as larvicidal and adulticidal agent against *Aedes aegypti* (Diptera: Culicidae). *Scientific reports*, 9 (1), 1-12. <https://doi.org/10.1038/s41598-019-45908-3>
- Satyal, P., Hieu, H. V., Chuong, N. T. H., Hung, N. H., Sinh, L. H., Tai, T. A., Hien, V. T., & Setzer, W. N. (2019). Chemical composition, *Aedes* mosquito larvicidal activity, and repellent activity against *Triatoma rubrofasciata* of *Severinia monophylla* leaf essential oil. *Parasitology research*, 118 (3), 733-742. <https://doi.org/10.1007/s00436-019-06212-1>
- Scalvenzi, L., Radice, M., Toma, L., Severini, F., Boccolini, D., Bella, A., Guerrini, A., Tacchini, M., Sacchetti, G., Chiurato, M., Romi, R., & Di Luca, M. (2019). Larvicidal activity of *Ocimum campechanum*, *Ocotea quixos* and *Piper aduncum* essential oils against *Aedes aegypti*. *Parasite*, 26 (23), 1-8. <https://doi.org/10.1051/parasite/2019024>
- Silva, I. M., Martins, G. F., Melo, C. R., Santana, A. S., Faro, R. R., Blank, A. F., Alves, P. B., Picanço, M. C., Cristaldo, P. F., Araújo, A. P. A., & Bacci, L. (2018). Alternative control of *Aedes aegypti* resistant to pyrethroids: lethal and sublethal effects of monoterpene bioinsecticides. *Pest management science*, 74 (4), 1001-1012. <https://doi.org/10.1002/ps.4801>
- Silva, L. S., Mar, J. M., Azevedo, S. G., Rabelo, M. S., Bezerra, J. A., Campelo, P. H., Machado, M. B., Trovati, G., Santos, A. L., Fonseca-Filho, H. D., Souza, T. P., & Sanches, E. A. (2019). Encapsulation of *Piper aduncum* and *Piper hispidinervum* essential oils in gelatin nanoparticles: a possible sustainable control tool of *Aedes aegypti*, *Tetranychus urticae* and *Cerataphis lataniae*. *Journal of the Science of Food and Agriculture*, 99 (2), 685-695. <https://doi.org/10.1002/jsfa.9233>
- Silva, M. F. R., Bezerra-Silva, P. C., Lira, C. S., Lima Albuquerque, B. N., Neto, A. C. A., Pontual, E. V., Maciel, J. R., Paiva, P. M. G. & Navarro, D. M. D. A. F. (2016). Composition and biological activities of the essential oil of *Piper corcovadensis* (Miq.) C. DC (Piperaceae). *Experimental parasitology*, 165, 64-70. <https://doi.org/10.1016/j.exppara.2016.03.017>
- Simões, C. M. O., Schenkel, E. P., Gosmann, G., Mello, J. C. P., Mentz, L. A., & Petrovick, P. R. (2004). *Farmacognosia: da planta ao medicamento*. UFSC.
- Simões, E. R. B., Santos, E. A., Abreu, M. C., Nascimento Silva, J., Nunes, N. M. F., Costa, M. P., Pessoa, O. D. L., Pessoa, C., & Ferreira, P. M. P. (2015). Biomedical properties and potentiality of *Lippia microphylla* Cham. and its essential oils. *Journal of intercultural ethnopharmacology*, 4 (3), 256. <https://doi.org/10.5455/jice.20150610104841>
- Soonwera, M., & Phasomkusolsil, S. (2016). Effect of *Cymbopogon citratus* (lemongrass) and *Syzygium aromaticum* (clove) oils on the morphology and mortality of *Aedes aegypti* and *Anopheles dirus* larvae. *Parasitology research*, 115 (4), 1691-1703. <https://doi.org/10.1007/s00436-016-4910-z>
- Stadnik, A., Oliveira, M. I. U. D., & Roque, N. (2016). Levantamento florístico de Myrtaceae no município de Jacobina, Chapada Diamantina, estado da Bahia, Brasil. *Hoehnea*, 43 (1), 87-97. <https://doi.org/10.1590/2236-8906-46/2015>
- Stappen, I., Wanner, J., Tabanca, N., Wedge, D. E., Ali, A., Kaul, V. K., Lal, B., Jaitak, V., Gochev, V. K., Schmidt, E., & Jirovetz, L. (2015). Chemical composition and biological activity of essential oils of *Dracocephalum heterophyllum* and *Hyssopus officinalis* from Western Himalaya. *Natural product communications*, 10 (1), 133-138.

Tabanca, N., Gao, Z., Demirci, B., Techen, N., Wedge, D. E., Ali, A., Sampson, B. J.; Werle, C.; Bernier, U. R.; Khan, I. A.; & Baser, K. H. C. (2014). Molecular and phytochemical investigation of *Angelica dahurica* and *Angelica pubescens* essential oils and their biological activity against *Aedes aegypti*, *Stephanitis pyrioides*, and *Colletotrichum* species. *Journal of agricultural and food chemistry*, 62 (35), 8848-8857. <https://doi.org/10.1021/jf5024752>

Trindade, E. L., Garcia, F., Ferreira, R., & Pasa, M. C. (2016). Lamiaceae-levantamento de dados das plantas medicinais recorrentes no estado de Mato Grosso presentes no herbário UFMT campus de Cuiabá-MT. *Biodiversidade*, 15 (2), 183-190.

Tyagi, V., Patel, R., Hazarika, H., Dey, P., Goswami, D., & Chattopadhyay, P. (2017) Chemical composition and bioefficacy for larvicidal and pupicidal activity of essential oils against two mosquito species. *International Journal of Mosquito Research*, 4, 112-118.

Vivekanandhan, P., Usha-Raja-Nanthini, A., Valli, G., & Subramanian Shivakumar, M. (2020). Comparative efficacy of *Eucalyptus globulus* (Labill) hydrodistilled essential oil and temephos as mosquito larvicide. *Natural product research*, 34 (18), 2626-2629. <https://doi.org/10.1080/14786419.2018.1547290>

Voris, D. G. R., Dias, L. S., Lima, J. A., Lima, K. S. C., Lima, J. B. P., & Lima, A. L. S. (2018). Avaliação das atividades larvicida, adulticida e anticolinesterase de óleos essenciais de *Illicium verum* Hook. f., *Pimenta dioica* (L.) Merr., e *Myristica fragrans* Houtt. contra vetores do vírus Zika. Ambiente. *Environmental Science and Pollution Research*, 25 (23), 22541-22551. <https://doi.org/10.1007/s11356-018-2362-y>

Yang, S., Bai, M., Yang, J., Yuan, Y., Zhang, Y., Qin, J., Kuang, Y., & Sampietro, D. A. (2020). Chemical composition and larvicidal activity of essential oils from *Peganum harmala*, *Nepeta cataria* and *Phellodendron amurense* against *Aedes aegypti* (Diptera: Culicidae). *Saudi Pharmaceutical Journal*, 28 (5), 560-564. <https://doi.org/10.1016/j.jsps.2020.03.007>