

## Caatinga: barn of antioxidant and nutraceutical bio-actives

Caatinga: celeiro de antioxidantes e bioativos nutracêuticos

Caatinga: granero de antioxidantes y bioactivos nutracéuticos

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### **Abstract**

The Caatinga, of Brazilian exclusivity, is compound by adapted plants for the climate-specific conditions present in this region. The natural resources in this phytogeography domain are used for local communities as foods source, regional economics develops, and traditional medicine. Folk medicine is applied as a local tradition through the fraction of the plants, such as roots, leaves, and fruits, to treat inflammation, tumor, metabolic or degenerative diseases. Although the technological pharmaceutical industrial process shows exponential development in benefits for human health, the search for new treatments, naturals, and alternatives linked with the significant consciousness of the populations some knowledge about folk medicine and their natural compounds are not vast. In this way, the present review aimed to realize the systematic research considering 26 species endemics in Caatinga and their most significant biological potential. As a result, it was observed that have more studies with the antioxidative and immunomodulatory activities. The previous knowledge shows the relation between the accumulation of reactive oxygen species (ROS) and the development of different immunological diseases. With this review, we propose to contribute to the biotechnological potential of the Caatinga for science health.

**Keywords:** Bioprospection; Inflammation; ROS; Folk medicine; Natural compounds; Cancer.

### **Resumo**

A Caatinga, exclusivamente brasileira, é composta por plantas adaptadas às condições climáticas específicas presentes nesta região. Os recursos naturais neste domínio fitogeográfico são usados pelas comunidades locais como fonte de alimentos, desenvolvimento da economia regional e medicina tradicional. A medicina popular é aplicada como uma tradição local através da fração das plantas, como raízes, folhas e frutos, para tratar inflamações, tumores, doenças metabólicas ou degenerativas. Embora o processo tecnológico industrial farmacêutico apresente um desenvolvimento

exponencial em benefícios para a saúde humana, a busca por novos tratamentos, naturais e alternativas atreladas à significativa conscientização das populações sobre a medicina popular e seus compostos naturais não são vastas. Desta forma, a presente revisão teve como objetivo realizar a pesquisa sistemática considerando 26 espécies endêmicas da Caatinga e seu potencial biológico mais significativo. Como resultado, observou-se que há mais estudos com as atividades antioxidante e imunomoduladora. O conhecimento prévio mostra a relação entre o acúmulo de espécies reativas de oxigênio (ROS) e o desenvolvimento de diferentes doenças imunológicas. Com esta revisão, propomos contribuir com o potencial biotecnológico da Caatinga para a saúde da ciência.

**Palavras-chave:** Bioprospecção; Inflamação; EROs; Medicina popular; Compostos naturais; Câncer.

### Resumen

La Caatinga, de exclusividad brasileña, está compuesta por plantas adaptadas a las condiciones climáticas específicas presentes en esta región. Los recursos naturales en este dominio de la fitogeografía se utilizan para las comunidades locales como fuente de alimentos, se desarrolla la economía regional y la medicina tradicional. La medicina popular se aplica como tradición local a través de la fracción de las plantas, como raíces, hojas y frutos, para tratar enfermedades inflamatorias, tumorales, metabólicas o degenerativas. Si bien el proceso industrial tecnológico farmacéutico muestra un desarrollo exponencial en beneficios para la salud humana, la búsqueda de nuevos tratamientos, naturales y alternativos ligada a la importante conciencia de las poblaciones sobre algunos conocimientos sobre la medicina popular y sus compuestos naturales no es muy amplia. De esta forma, la presente revisión tuvo como objetivo realizar la investigación sistemática considerando 26 especies endémicas de Caatinga y su potencial biológico más significativo. Como resultado, se observó que se tienen más estudios con las actividades antioxidante e inmunomoduladora. Los conocimientos previos muestran la relación entre la acumulación de especies reactivas de oxígeno (ROS) y el desarrollo de diferentes enfermedades inmunológicas. Con esta revisión, nos proponemos contribuir al potencial biotecnológico de la Caatinga para la ciencia de la salud.

**Palabras clave:** Bioprospección; Inflamación; ROS; Medicina popular; Compuestos naturales; Cáncer.

## 1. Introduction

Considered an exclusively Brazilian phytogeographic domain, the Caatinga occupies 11% of the entire national territory (M. V. da Silva et al., 2020) with a predominance in the country's Northeast region, between the Maranhão to Bahia states (Figure 1). Characterized by presenting a diversification in flora, the plants of this region are physiologically adapted to the sandy soil, temperature variations (14.29 ° C to 29.7 ° C) and long periods of drought (9 to 10 months) accumulating an annual rain volume of 1026 mm<sup>3</sup>(J. M. C. da Silva & Lacher, 2020). The environmental factors on vegetation contribute to the significant variability of phytochemical agents, with bioactivities already described, capable of attenuating or blocking biological pathways involved in neurodegenerative, tumoral, and inflammatory pathophysiological processes (Costa et al., 2020; da Silva Barbosa et al., 2020a; G. C. da Silva et al., 2020; Oliveira de Veras et al., 2020; I. B. da S. Santos et al., 2020).

**Figure 1:** Phylogeographic domain of the Caatinga in Brazilian territory (Source: [Caatinga | SiBBR](#))



Source: Authors.

Plant fractions, such as roots, stems, leaves, fruits, and seeds, are traditionally applied by popular medicine as a therapeutic resource for preventing and controlling circulatory, respiratory, dermatological, metabolic, endocrine, digestive disorders, among others. (Sile et al., 2020). Currently, the growing search for traditional folk medicine stimulates the use of natural products as a resource in the intervention of the health-disease process, bringing new concepts and the need for further research regarding the safety and efficacy of use. Because of this new scenario, some authors argue that any extract or isolated phytochemical agent capable of preventing pathologies or mitigating the pathological condition is considered a nutraceutical element aimed at rebalancing the normal physiological state (López-Gutiérrez et al., 2015).

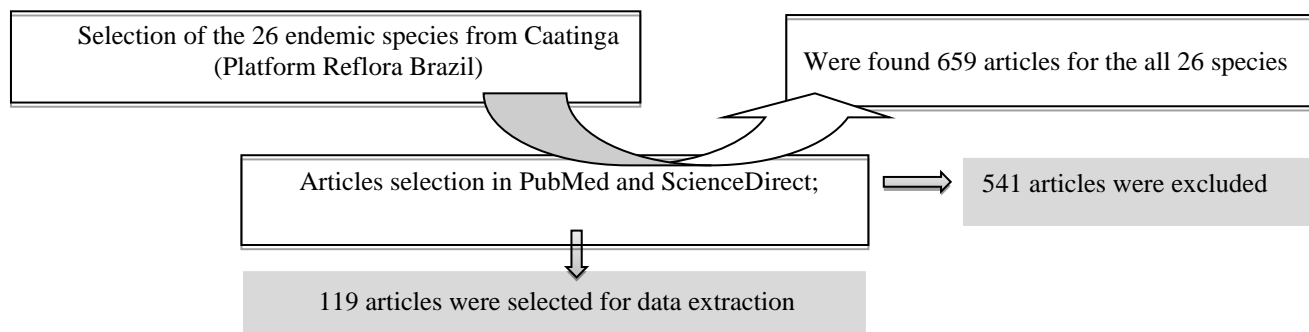
In this context, this article aims to discuss the nutraceutical potential of some plant species found in the Caatinga, addressing the main biological actions documented as antihypertensive (*Myrciaria floribunda*), hypoglycemic (*Cnidioscolus quercifolius*), regulation of the lipid profile (*Eugenia dysenterica*), cachexia inhibition (*Euphorbia tirucalli*) and anti-obesity (*Hancornia speciosa*)

## 2. Methodology

This is a narrative review, where the 26 species endemics in the Caatinga bases were selected on the “Species of Sociobiodiversity for Economics Interesting” list published by the Brazilian government in the Portaria Interministerial N° 284, March 30<sup>th</sup>, 2018. Species confirmation and inclusion of others not mentioned in the list were obtained from the website Re flora– Plants of Brazil ([www.reflora.jbrj.gov.br](http://www.reflora.jbrj.gov.br)). Articles between 2016 to 2020 were researched in Pubmed and ScienceDirect using the keywords “biological activities” and “bioactivity” associated with species' scientific names.

Articles with methodologies *in vitro*, *in vivo*, and clinical trials using extracts or molecules isolated from the vegetable fraction of the plant were selected. Review articles or studies not directed from human health were not included. The process of selection is representing in Figure 2. Based on the studies, the species with more articles were selected for being discussed. The relative rate of mainly biological activities in the total of the articles was performed in GraphPad Prism 6®.

**Figure 2:** Methodological flowchart for the selection and analysis of articles.



Source: Authors.

### 3. Results and Discussion

Considering all species researched were selected 119 articles (Table 1). The significant studies analyzed the antioxidative and immunomodulatory activity in hydroalcoholic extracts from different parts of the plants. The antioxidant assay represents 41.17% of all analyses and the immunomodulatory potential in the study in 24.16% of the articles. Both investigations correlate with diseases like neurodegenerative, anticancer properties, and metabolic dysfunction in many articles. The distribution of bioactivities present in the papers is shown in Figure 3.

**Table 1:** The all 26 species endemics in Caatinga with the respective compounds, source of extractions and biological activity.

| Species                        | Biological activity                             | Compounds  | Sources   | References  |
|--------------------------------|---|--|---|---|
| <i>Anacardium occidentale</i>  | Antihypertensive                                | Peptides   | Cashwe Nut  | (Amorim et al., 2018)   |
|                                | Anti-inflammatory, Bronchodilator               | Oleamid  | Leaves  | (Awakan et al., 2018)   |
|                                | Anticancer                                      | Polysaccharide, Cardanol Derivative, Zoapatanoloid, Agastiflavone, Anacardicin, Methyl gallate                               | Fruit residue, Chestnut, Leaves                                   | (Barros et al., 2020; Braga et al., 2021a; de Oliveira Silva Ribeiro et al., 2020; M Ashraf & Rathinasamy, 2018a; J. M. Santos et al., 2019; Sunderam et al., 2019; Taiwo et al., 2017) |
|                                | Glycemic metabolic regulation and liver markers | Fibers   | Fruit residue   | (Carvalho et al., 2018)   |
|                                | Antimicrobial activity                          | Phenolics and Flavonoids   | Bark Stalk, Leaves and Seeds                                      | (J. S. C. de Araújo et al., 2018; G. H. F. dos Santos et al., 2018; M Ashraf & Rathinasamy, 2018a; Morvin Yabesh et al., 2019; Sunderam et al., 2019)                                   |
|                                | Anti-inflammatory                               | Anacardic acid, cardol, cardanol and methylcardo, Gallic acid, Ellagic, Shikimic, phosphoric and benzoic and Polysaccharides | Chestnut, Fruit, Flowers  | (M. Q. de Souza et al., 2018a; Ferreira-Fernandes et al., 2019; Goulart da Silva et al., 2021a; A. S. Oliveira et al., 2019; Souza Filho et al., 2018a)                                 |
|                                | Gastoprotection and Motility                    | Carotenoids and anarcadic acid, Oleic acid, 11-Octadecanoic acid methyl ester, Anacardol                                     | Fruit residue and Bark Stalk                                      | (Goulart da Silva et al., 2021a; Omolaso et al., 2020a)   |
|                                | Anxiolytic, Anticonvulsant, Neuroprotective     | Anacardic Acid, Agastiflavone, Phenolics and Flavonoids  | Leaves and Chestnut   | (Gomes Júnior et al., 2018a; Junsathian et al., 2018; Luiz Gomes et al., 2018a; Velagapudi et al., 2018a)   |
| <i>Arachis hypogaea</i>        | Cell protection                                 | Phenolic compounds (catechin, epicatechin, procyanidin and proanthocyanidin and quercetin dimers)                            | Grain skin  | (Rossi et al., 2020b)   |
|                                | Antihemolytic activity                          | Luteolin   | Bark  | (M. Peng et al., 2021)  |
|                                | Anti-bacterial                                  | Stilbene (trans-arachidine-1 and trans-arachidine-3)   | Roots   | (Eungsuwan et al., 2021a)   |
|                                | Antioxidant / anti-inflammatory                 | Flavonoids, Stybenes, Anthocyanins   | Integument, roots and black peanut shells; Leaves; Sprouted grain | (Cossetin et al., 2019; Fernandes et al., 2020; Limmongkon et al., 2018, 2019; J. Peng et al., 2019b)   |
|                                | Differentiation of osteoblasts                  | Concentrated aqueous extract   | Peanut sprouts  | (Kim et al., 2019b)   |
| <i>Byrsonima verbascifolia</i> | Antioxidant, antidiarrheal,                     | Ethanol extract, ethyl acetate extract, hexane extract   | Leaves  | (de Araújo Rodrigues et al., 2019)  |

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| <i>Byrsonima crassifolia</i>     | gastroprotective                                   | Phenolic compounds   | Leaves                      | (R. O. de Souza et al., 2018)                                     |
| <i>Byrsonima cydoniifolia</i>    | Photochemical protection                           | flavonoids and stilbenes   | Fruits                      | (V. S. dos Santos et al., 2017)                                   |
| <i>Campomanesia guazumifolia</i> | Antioxidant, antimicrobial                         | Antioxidant activity<br>monoterpenes<br>terpinolene, sabinense, $\beta$ -mycrene, $\alpha$ -terpinene, $\gamma$ -terpinene and the farnesol and guaiol sesquiterpenes  | Leaves                      | (A. L. dos Santos et al., 2019)                                   |
|                                  | Anti-inflammatory, antiedematogenic                | Antimicrobial activity:<br>Bicyclogermacrene, $\beta$ -pinene and karyophylene<br>Glycosylated flavonoids and cyclohexanecarboxylic acid<br>quercetin pentose, quercetin deoxyhexoside, myricetin deoxyhexoside and quinic acid  | Leaves                      | (Catelan et al., 2018)  |
| <i>Cnidocolus quercifolius</i>   | Hypoglycemic                                       | Phenols, flavones, flavonols, xanthonenes, catechins, triterpenes and tannins.   | Leaves                      | (Lira et al., 2017)   |
|                                  | Antibacterial                                      | Lupeol-3 $\beta$ -O-cinnamate and lupeol-3 $\beta$ -O-dihydrocinnamate, bis-nor-diterpene filacantona  | Bark Stalk                  | (de Oliveira-Júnior et al., 2018)                                 |
|                                  | Antioxidant, anti-inflammatory and antinociceptive | Phenolics (vanillin, eugenol and quercetin)  | Seeds                       | (Ribeiro et al., 2021)  |
|                                  | Antioxidant  | Phenolics and Flavonoids   | Seed, oil and residual cake | (Ribeiro et al., 2017)  |
| <i>Croton argyrophyllus</i>      | Antioxidant, antimicrobial, antifungal             | Bicyclogermacrene, $\beta$ -pinene and spatulenol<br>Total phenols and flavonols   | Leaves and Stalk            | (da Silva Brito et al., 2018)                                     |
| <i>Eugenia brejoensis</i>        | Trypanocidal                                       | $\delta$ -cadineno, trans-cariofileno e $\alpha$ -Muurolol   | Leaves                      | (Oliveira de Souza et al., 2017);<br>(Bezerra Filho et al., 2020) |
|                                  | Antioxidant, anti-inflammatory                     | hydroxybenzoic acids, vanilic acid-O-hexoside, Hexoside ellagic acid / hydroxycinnamic acids and their derivatives<br><br>Catechin and Epicatechin<br>Quercetin (dihydro quercetin glycoside), Flavanones of the flavanone class (naringin. Hydrate, eriodictiol) eriodictiol-7-O-glucoside, Tellimagrandin II / pterocaryanin C, two Di-HHDP-galloyl-glucose (casuarictine / potentiline) isomers and five Di-HHDP-galloyl-glucose (casuarictine / potentiline) ellagic acid derivatives) | Pulp                        | (Soares et al., 2019); (F. F. de Araújo et al., 2021)             |
|                                  | Antinociceptive, anti-inflammatory, Antipyretic    | Guaiol, trans-karyophylene and $\beta$ -eudesmol and $\gamma$ -eudesmol  | Leave                       | (Costa et al., 2020)  |
| <i>Eugenia dysenterica</i>       | Neuroprotective, Antioxidant                       | N/D  | Leaves                      | (Thomaz et al., 2018a)  |
|                                  | Prevent hypertriglyceridemia, antioxidant          | N/D  | Leaves                      | (A. B. Lima et al., 2017)   |

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|                            | Regulation of the lipid profile, fasting hyperglycemia and glucose intolerance  | Polyphenols  | Pulp                                      | (Donado-Pestana et al., 2018b)                       |
|                            | Anti-inflammatory, Angiogenic   | $\alpha$ -humuleno, $\beta$ -cariofileno   | Leaves                                    | (S. M. M. Da Silva et al., 2019a)                    |
|                            | Hypotensive   | Proanthocyanidins  | Leaves                                    | (Fidelis-de-Oliveira et al., 2020)                   |
|                            | Antioxidant and chelator  | Polyphenols  | Leaves                                    | (Ávila et al., 2016a)                                |
|                            | Moderate cytotoxic against SH-SY5Y  | Flavonoids, quercetin and catechin   | Leaves                                    | (Gasca et al., 2017)                                 |
|                            | Antioxidant and anti-glycation  | Ferulic and gallic acids, myricetin, quercetin and kaempferol-pentosides   | Pulp                                      | (Justino et al., 2020a)                              |
| <i>Euphorbia tirucalli</i> | Antimicrobial   | Phenolics: myricetin, 3,3'-dimethoxy-4-O-a-rhamnopyranoside-ellagic acid, 4 O-methyl-gallicacid and ampelopsin.  | Root                                      | (M. de F. R. de Lima et al., 2021)                   |
|                            | Antioxidant, Anticancer   | Steroidal groups: Pregn-4-ene-3,20-dione, 11-Hydroxyl , 9,19-Cyclo-9.beta.-lanostane-3.beta.,25-diol, Lanosterol, Phenolics: Galic, Ferulic, Sinapic, Rutin, Quercetin | Stems                                     | (Abdel-Aty et al., 2019)                             |
|                            | Anticancer  | Euphol, and tirucallol, myristic, palmitic, linoleic, oleic and stearic acids,   | Stems                                     | (L. S. de Souza et al., 2019)                        |
|                            | Anticancer  | Tirucadalenone, Euphorol L   | Stems                                     | (Duong et al., 2019)                                 |
|                            | Nociceptive, anti inflammatory  | Tannin, Lupeol, b-Sitosterol   | Root                                      | (Palit et al., 2018)                                 |
|                            | Anticancer, immunomodulation and cachexia inhibition  | Euphorol   | Stems                                     | (Martins et al., 2020)                               |
|                            | immunomodulation  | Triterpenes  | Leaves                                    | (Ibrahim et al., 2019)                               |
|                            | Anticancer  | Euphol   | Stems                                     | (V. A. O. Silva et al., 2019)                        |
| <i>Eryngium foetidum</i>   | Antimicrobial   | Alkaloids, flavonoids, phenolics, anthraquinones, steroid  | Methanol extract from leaves              | (Kouitchou Mabeku et al., 2016)                      |
|                            | Antioxidant   | (E)-2-Dodecenal, 13-tetradecenal, dodecanal, 2,4,5-trimethylbenzaldehyde   | Essential oil from roots, stem and leaves | (Thomas et al., 2017)                                |
| <i>Hancornia speciosa</i>  | Angiogenic, antibacterial, antioxidant  | Flavones, flavonols, flavanones, and tannins   | Latex fractions                           | (D'abadia et al., 2020)                              |
|                            | Antioxidant, antimutagenic, enzymatic inhibition (acetylcholinesterase, butyrylcholinesterase, tyrosinase, hyaluronidase, lipase, $\alpha$ -amylase, and $\alpha$ -glycosidase), antiobesity, antihyperglycemic | Carotenoids and polyunsaturated fatty acids  | Ethanol extract of leaves                 | (U. P. Dos Santos et al., 2018a)                     |
|                            | Anti-inflammatory   | Rutin and chlorogenic acid   | Aqueous extract from the fruits           | (Bitencourt et al., 2019; Torres-Rêgo et al., 2016a) |
|                            | Antihypertensive  | Cyclitol bornesitol  | Leaves                                    | (Moreira et al., 2019a)                              |

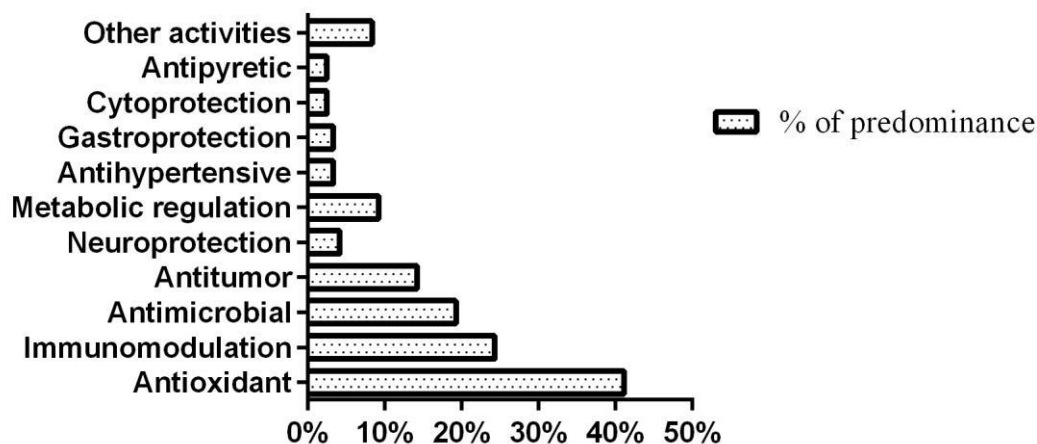
|                              |  |  |  |   |
|------------------------------|--|--|--|---|
|                              | Antioxidant, antimicrobial, cytotoxic  | Quinic acid, chlorogenic acid, catechin, rutin, isoquercitrin, kaempferol-rutinoside, and catechin-pentoside.  | ethanolic extract of leaves                                    | (U. P. Santos, Campos, Torquato, Paredes-Gamero, Carollo, Estevinho, De Picoli Souza, et al., 2016) |
|                              | Anti-inflammatory, antioxidant   | Bornesitol, quinic acid, ascorbic acid, chlorogenic acid, isochlorogenic acid, 3-feruloylquinic acid, rutin, 5-feruloylquinic acid, quercetin-3- <i>O</i> -hexoside, kaempferol-rutinoside, kaempferol-hexoside, isorhamnetin-3- <i>O</i> -rutinoside, and quercetin | Fruit juice  | (de Oliveira Yamashita et al., 2020)  |
|                              | Antioxidant, <i>In vitro</i> gastrointestinal digestion  | Protocatechuic acid, <i>p</i> -Coumaric acid, Salicylic acid, Syringic acid, <i>trans</i> -Cinnamic acid, Gentisic acid, Vanillic acid, Ferulic acid, Elagic acid, Gallic acid, Rutin, Myricetin, Quercetin, Catechin, Hesperetin, Kaempferol                        | Pulp of fruits   | (Dutra et al., 2017a)   |
| <i>Hymenaea cangaceira</i>   | Antinociceptive, antioxidant, antimicrobial  | Hydrocarbon sesquiterpenes (SH): $\alpha$ -Copaene, $\beta$ -Elemene, (E)-Caryophyllene, $\alpha$ -Guaiene, $\alpha$ -Humulene, Germacrene D   | Essential oil from leaves                                      | (Oliveira de Veras et al., 2020)  |
| <i>Hymenaea stignocarpa</i>  | Enzymatic inhibition ( $\alpha$ -amylase and $\alpha$ -glucosidase), nutritional quality, glycemic index | caffeic acid, kaempferol, quercetin-3-rutinoside and quercetin-3-rhamnoside  | N/D  | (C. P. da Silva et al., 2019)   |
| <i>Myrciaria floribunda</i>  | Antitumor, antioxidant   | Phenolics, flavonoids and tannin   | Ethyl acetate extract from the leaves                          | (de Azevedo et al., 2019; Tietbohl et al., 2017)  |
|                              | Enzymatic inhibition of acetylcholinesterase   | Sesquiterpenes: $\delta$ -Cadinene, $\gamma$ -Cadinene, $\gamma$ -Muurolene, $\alpha$ -Selinene, $\alpha$ -Muurolene (E)-Caryophyllene   | Essential oil from the fruit peel                              | (da Silva Barbosa et al., 2020b)  |
|                              | Antimicrobial, antioxidant   | Nor-lupane triterpenoids platanic acid and messagenic I acid; triterpenoids (betulinic aldehyde, ursolic acid acetate, betulinic acid, 2 $\alpha$ ,6 $\alpha$ ,30-trihydroxybetulinic acid); flavonoids (catechin, quercetin and myricitrin)                         | Methanol extract from the leaves                               | (de Azevedo et al., 2019)   |
| <i>Oocotea glomerata</i>     | Antifungal   | cinnamic acid derivatives and flavonoids   | Hydro alcoholic extracts from the leaves                       | (Anjos et al., 2020)  |
| <i>Psidium guineense</i>     | antioxidant, anti-inflammatory, antiproliferative and antimycobacterial                                  | Monoterpenes, oxygenated monoterpene, sesquiterpene, oxygenated sesquiterpene  | Essential oil from the leaves                                  | (do Nascimento et al., 2018)  |
|                              | Antinociceptive and anti-inflammatory  | Spathulenol  | Essential oil from the leaves                                  | (E. Dos Santos et al., 2020)  |
| <i>Passiflora cincinnata</i> | Atimicrobial   | N/D  | hydroalcoholic extracts of leaves, stems, bark, pulp and seeds | (Siebra et al., 2018)   |



|                                 |  |   |  |  |
|---------------------------------|--|---|--|--|
|                                 | antinociceptive and anti-inflammatory                                    | Flavonoids  | Ethanol extract of the aerial parts                          | (de Lavor et al., 2018)  |
|                                 | Antioxidant  | Phenolics: isoquercetin, caftaric acid and rutin; $\beta$ -carotene, flavonoids   | Pulp   | (de Souza Silva et al., 2020)  |
|                                 | Antioxidant  | Phenolic compounds, flavonoids  | Ethanol extracts of leaves, seeds, stem, flowers, fruit peel | (Leal et al., 2020)  |
| <i>Schinus terebinthifolius</i> | Antioxidant  | doxorubicin, polysaccharides $\alpha$ -pinene and $\alpha$ -felandrene, oleic acid, $\alpha$ -felandrene, $\delta$ -cadynene, oleic and palmitic. | Fruits and Leaves  | (Aumeeruddy-Elalfi et al., 2015; P. D. S. da Rocha et al., 2020a; dos Santos da Rocha et al., 2019b; Feriani et al., 2020a, 2021; M. D. C. L. Lima et al., 2020a; P. dos S. da Rocha et al., 2018a; Salem et al., 2018b; Todirascu-Ciornea et al., 2019) |
|                                 | Atimicrobial   | ácido gálico, galotaninos e flavonóis glicosilados, $\alpha$ -pineno e $\alpha$ -felandreno   | Fruits   | (P. dos S. da Rocha et al., 2018b; Salem et al., 2018b)  |
|                                 | Antidiabetic Activity  | gallic acid, galotanins and glycosylated flavonols  | Fruits   | (Feriani et al., 2020a)  |
|                                 | Antinociceptiva  | Flavonoids  | Fruits   | (Feriani et al., 2020a)  |
|                                 | Endodontic treatment   | Flavonoids  | N/D  | (Pinto et al., 2020a)  |
|                                 | Anti-inflammatory  | Polysaccharides and flavanones  | Fruits   | (Estevão et al., 2017)   |
|                                 | Angiogenics  | $\alpha$ -pinene and $\alpha$ -felandrene   | Leaves   | (Estevão et al., 2017)   |
|                                 | Keratitis  | $\alpha$ -pinene and $\alpha$ -felandrene   | Leaves   | (M. D. C. L. Lima et al., 2020a)   |
|                                 | Atin viral   | Phenolic compounds (resveratrol, catechin and epicatechin)  | Peel and fruits  | (M. B. S. Oliveira et al., 2020)   |
| <i>Spondias tuberosa</i>        | Regulation in lipid metabolism   | N/D   | Bark   | (de Moura Barbosa et al., 2018)  |
|                                 | Antifungal   | N/D   | Folhas   | (Cordeiro et al., 2020)  |
| <i>Syagrus Coronata</i>         | Antibacterial and antibiofilm, Antifungal, anti-inflammatory and healing | Fatty acids (octanoic acid, dodecanoic acid, decanoic acid and $\gamma$ -eudesmol)  | Seeds  | (Souza dos Santos et al., 2019)  |
| <i>Verbesina macrophylla</i>    | Hemolytic, antimicrobial, anti-inflammatory and antipyretic activity     | Sesquiterpenos e hidrocarbonetos  | Essential oil of the leaves                                  | (de Veras et al., 2021)  |
| <i>Xanthosoma sagittifolium</i> | Significant effects on the intestinal microbiota.                        | Starch, amylose and milopectin  | Fruits   | (Graf et al., 2018)  |

Source: Authors.

**Figure 3:** Distribution of biological activity study in the select articles.



Total of articles: 119

Source: Authors.

### 3.1 Chemical composition of plants observed in the Caatinga

Environmental conditions such as soil, climate, and interaction between fauna and flora determine factors for structuring the phytochemical composition of bioactive compounds (Teixeira et al., 2010). The Caatinga domain, when compared to the other biomes found in Brazil, presents as main characteristics the predominance of regions with dry soil between 9 to 10 months of drought, and irregular distribution of rainfall during 2 to 3 months, accumulating in some regions approximately 1.026 mm<sup>3</sup> (J. M. C. da Silva & Lacher, 2020). Irregularities in the region's climate favor the development of vegetation adapted to low rainfall, requiring the production of primary and secondary metabolites for the maintenance and survival of the species, changes in the environmental conditions affect directly in the metabolic production of the plant (Jia et al., 2014). The metabolites mainly the secondaries metabolites as phenolics and flavonoids are necessary for the plant survive, including activities in growth regulation, enzyme inhibition, antioxidant activity and ultraviolet light (UV) (Jia et al., 2014; Räisänen et al., 2008).

Considering what was observed in Table 1, the bioactive obtained from different fractions between roots, leaves, stems, fruits, and residues, are included among the classes of Flavanoids: Agathisflavone (*Anacardium occidentale*), Luteolin (*Arachis hypogaea*), and kaempferol (*Hancornia speciosa*); Phenolics: Stilbene (*Byrsonima cydoniifolia*) among others, and Terpenes:  $\alpha$ -Muurolol (*Eugenia brejoensis*), Zoapatanol (*Anacardium occidentale*) and Bicyclogermacrene (*Campomanesia guazumifolia* and *Croton argyrophyllus*).

The phenolic compounds and flavonoids are the major groups referred to with antioxidant activity in hydrophilic or lipophilic systems, such as cardanol, comprised among the phenolic lipids found in *Anacardium occidentale*, with anti-inflammatory action (Cossetin et al., 2019; Souza Filho et al., 2018a). Under stressful environmental conditions, as observed in the Caatinga, there is a natural increase in the concentration of phenolic compounds as a self-defense mechanism (Sharma et al., 2019). Chemically, phenolic compounds have in their molecular structure a hydroxyl radical (-OH) linked to an aromatic ring (Figure 4). The antioxidant activity of phenolic compounds comprises the structuring of the chemically active molecule; changes in the hydroxyl radical can positively or negatively compromise the antioxidant action of the compound (Regueira et al., 2017; Teixeira et al., 2010). The group of phenolics can be divided into flavonoid compounds (Ex. Anthocyanins) and non-

flavonoids (Ex. Stilbenes and Lignins). Both groups are present in the plants cataloged in this study, with specific biological activities, such as antibacterial and anti-inflammatory activity (*Arachis hypogaea*), photoprotection (*Byrsonima cydoniifolia*), hypotensive activity (*Eugenia dysenterica*), and hypoglycemic (*Cnidioscolus quercifolius*) (Eungsuwan et al., 2021a; Fidelis-de-Oliveira et al., 2020; Lira et al., 2017).

Terpenes are the second most observed class in bioactive compounds extracted from the plant fractions presented in this paper. Terpenoid groups are included within the secondary compounds of plant metabolism. Made up of sequences of isoprenes with five carbon units (C5), terpenes are chemically classified as monoterpenes (C10), sesquiterpenes (C15), and diterpenes (C20) according to the number of isoprenes associated with the final molecule (Yang et al., 2016). Terpenes comprise the most abundant secondary metabolites, with approximately more than 36,000 being recognized and acting as a crucial element in the interaction of the plant species with the environment in which it is inserted (Gershenzon & Dudareva, 2007).

Among the activities described were reported the antioxidant, antimicrobial, and antifungal potential of the compound bicyclogermacrene isolated from *Campomanesia guazumifolia* and *Croton argyrophyllus* species. The Spathulenol obtained by extracting the leaves of *Psidium guineense* demonstrates antinociceptive and anti-inflammatory activity, similar to the terpene caryophyllene from *Eugenia brejoensis* leaves.

### 3.2 Correlation of antioxidant potential and immunomodulatory capacity

Antioxidant compounds are reducing agents with the ability to regulate the concentration of reactive oxygen species (ROS) (e.g., OH<sup>·</sup>, NO<sup>·</sup>, HOO<sup>·</sup>) that are naturally produced by mitochondrial cellular respiratory metabolism or NADPH oxidase (NOX) activity, present in phagocytes and epithelial cells. Under normal physiological conditions, ROS plays an essential role in molecular signaling, cell differentiation, and activation of apoptotic mechanisms in tumor cells (Mittal et al., 2014).

The maintenance of physiological ROS concentrations is performed by organic molecules such as superoxide dismutase (SOD), catalase (CAT), glutathione peroxidase (GPx), glutathione reductase (GR), glutathione S-transferase (GST), and glutathione (GSH) (Ansari et al., 2020; Fukui & Zhu, 2010; Rhee et al., 2012). The imbalance between these organic molecules and the generation of ROS, marked by an increase in the [antioxidant] / [ROS] ratio, contributes to the deleterious effects of ROS on proteins, lipids, and DNA / RNA molecules, such as the activation of cellular immune response with a pro-inflammatory profile mediated initially by T-helper 1 (Th-1) and Th-17 cells, with activation of macrophages, neutrophils and local release of cytokines and chemokines (Agita & Thaha, 2017). The phagocytic system is an essential source of peroxidases responsible for combating pathogens. Neutrophils comprise significant concentrations of myeloperoxidases (MPO), and their activation in inflammatory conditions contributes to the elevation of ROS (Goulart da Silva et al., 2021a; Souza Filho et al., 2018a; Zhang et al., 2020), increasing the imbalance in the [antioxidant] / [ROS] ratio.

The inflammatory process mediated by ROS initially involves the transcription factor-kappa B / activating protein 1 (NF- $\kappa$ B), found in connection with the molecule I $\kappa$ B $\alpha$  in the cell cytoplasm. The I $\kappa$ B $\alpha$  / NF- $\kappa$ B complex inhibits the translocation of NF- $\kappa$ B from the cytoplasm to the nucleus, thereby controlling the expression of genes encoding cytokines chemokines that act in the inflammatory process (Prasad et al., 2017). Antioxidant therapies have been a step towards the alternative treatment of inflammatory disorders that result in pathologies such as cancer, diabetes, and neurodegenerative diseases (Belcaro et al., 2018; Seyyedebrahimi et al., 2018).

The antioxidant action of phytochemicals can occur by (i) increasing SOD, CAT, GPx, GR, GST, or GSH (ii) as direct reducing agents when interacting with ROS, and (iii) inhibiting Toll-like receptor (TLR). Palitt et al. (2018) demonstrated the action of terpene groups of aqueous extracts of *Euphorbia tirucalli* (Alvelós) in the inhibition of TLR-4 in

macrophage cells treated and exposed to lipopolysaccharide (LPS). The endotoxin LPS is recognized by the TLR-4, which presents as one of the intracellular domains TRIF, an inducer of INF-1 gene transcription (Chen et al., 2018). The authors found a decrease in inflammatory markers IL-6, IL-12, TNF- $\alpha$ , and INF- $\gamma$  cytokines (Palit et al., 2018), proving the involvement of terpene antioxidants in inhibiting the inflammatory pathway. Other studies point to a positive correlation between antioxidant phytochemicals and anti-inflammatory potential in different pathologies, which are better discussed below.

### ***Arachis hypogaea***

Peanut (*Arachis hypogaea L.*) is an herbaceous plant belonging to the Fabaceae family and grown worldwide. The genus *Arachis* is native to South America and includes 80 species. More than 50% of the world's production of peanuts is used to make peanut butter and oil, their primary forms of consumption. In addition, it is also ingested in the form of flour and candy (Limmongkon et al., 2018; Treuter et al., 2017). It is a legume with an abundance of nutrients and chemical constituents, such as proteins, carbohydrates, fibers, fats, niacin, folate, thiamine, arachidic acid, flavonoids, magnesium, phosphorus, polyphenols, and bioactive components (Limmongkon et al., 2018; Menis Candela et al., 2020).

Among the selected articles, the analysis of the compounds of the roots, leaves, seed shoots, and byproducts of peanut processing, such as skin and shell, were the study targets. Among the activities identified are antioxidants and anti-inflammatory drugs in a significant way, as well as a display of anti-hemolytic activity, inhibition of prostatic, antibacterial, and anti-adipogenic hyperplasia (Cossetin et al., 2019; Eungsuwan et al., 2021b; Fernandes et al., 2020; Kim et al., 2019a; Limmongkon et al., 2018; J. Peng et al., 2019a; Rossi et al., 2020a).

The industrial manufacturing process is usually done by blanching or roasting the peanuts without the skin of the seeds, where they are destined for animal feed (resulting in about  $7.5 \times 10^5$  tons of shell each year (J. Peng et al., 2019a; Rossi et al., 2020a). However, it is known that these byproducts are sources of phenolic acids, flavonoids, stilbenes, and various procyanidin and proanthocyanidin oligomers, which configures them as bioactive and nutraceutical compounds, mainly due to their properties of elimination against reactive oxygen species (ROS) in biological systems (Bodoira et al., 2017; Larrauri et al., 2016). The general phenolic profiles vary according to the peanut cultivar, germination stage, growing season, and growing conditions storage (Rossi et al., 2020a). Studies have also reported that such biological activities can be enhanced through peanut sprouts (Kim et al., 2019a; Limmongkon et al., 2019).

It has been established that increased ROS production could lead to tissue damage and the mediation of chronic and inflammatory diseases, such as diabetes, cardiovascular diseases, neurodegenerative diseases, and osteoarthritis (Lepetsos et al., 2019; Locatelli et al., 2018). The stilbene compounds, abundantly found in peanuts, belong to a polyphenolic group and are characterized by a 1,2-diphenylethylene-based phytoalexin with trans-resveratrol acting by inhibiting pro-inflammatory mediators, such as prostaglandins, thromboxanes, and leukotrienes (Limmongkon et al., 2018).

Through the germination of the peanut seed, Kim et al. (2019) demonstrated that the aqueous extract and its phytochemicals, the yasaponin Bb, potentiate the differentiation of osteoblasts mediated by bone morphogenetic protein - 2, leading to the expression of factors necessary for bone formation.

In addition to these properties, Peng et al. (2021) found that the anthocyanins in black peanut shells have an anti-adipogenic function, inhibiting lipids' accumulation depending on the concentration used. This function is based on the partial inhibition of digestion enzymes:  $\alpha$ -glucosidase,  $\alpha$ -amylase, and lipase, by decreasing the digestion of fat and carbohydrate in fatty acids and glucose, acting as a strategy to regulate body weight.

### ***Anacardium occidentale***

Comprised in the Anacardiaceae family, *Anacardium occidentale* L. is geographically distributed throughout the Brazilian territory, with high occurrence in the Northeast region (Borges, 2021). According to data from the Brazilian Institute of Geography and Statistics- IBGE (<https://sidra.ibge.gov.br/>), the fruits, popularly known as cashew, are an important economic source for local agriculture that concentrates approximately 98% of the country's cultivars.

From the cashew processing, four byproducts sources of bioactive compounds can be obtained, i) raw nuts, ii) cashew grains, iii) apple-shaped stems (cashew apples) and the residue after juice extraction, and iv) the cashew nut shell liquid (CNSL) (Braga et al., 2021a). The identification and isolation of phytochemicals from these sources can be therapeutic alternatives for pathologies such as cancer (de Oliveira Silva Ribeiro et al., 2020), periodontitis (Souza Filho et al., 2018b), diarrheal conditions (Omolaso et al., 2020a), neurodegenerative diseases (Junsathian et al., 2018; Velagapudi et al., 2018a) and enzyme inhibitor (Amorim et al., 2018).

The anacardic acid, cardol, cardanol, and methylcardo, mainly isolated from nuts and CNSL, are among the primary compounds with the biological activities mentioned (Braga et al., 2021b; M. Q. de Souza et al., 2018a; Gomes Júnior et al., 2018a; Goulart da Silva et al., 2021b; Luiz Gomes et al., 2018a; M Ashraf & Rathinasamy, 2018a; Omolaso et al., 2020b; Taiwo et al., 2017). Compounds like Gallic acid, Ellagic acid, Shikimic acid, phosphoric acid, and benzoic acid, known for their potential antioxidants, are found preferentially in the leaves and flowers of cashew trees (A. S. Oliveira et al., 2019; J. M. Santos et al., 2019; Sunderam et al., 2019).

The imbalance between natural antioxidant molecules, such as natural antioxidants GSH, CAT, and SOD at the neuronal level, compromises neuronal functionality by accumulating ROS with activation of the local inflammatory response inducing the neurodegenerative process with consequent cell death. The accumulation of ROS is one of the factors involved in the pathophysiology of anxiety, depression, seizures, and aggravation in neurodegenerative diseases, such as Alzheimer's disease (AD) and Parkinson's (PD) (Simpson & Oliver, 2020).

In neuronal cells, anacardic acid has anxiolytic action, via GABA receptor, by increasing the concentration of GSH, SOD, and CAT that result in the inhibition of lipid peroxidation in the hippocampus and frontal cortex, without suppression of the neuromotor response observed in traditional anxiolytics (Gomes Júnior et al., 2018b; Luiz Gomes et al., 2018b). Contrary to anacardic acid, the neuroprotective action, in microglia cells, of the isolate Agastiflavone obtained from the cashew leaves inhibits the elevation of the ROS concentration. It is possible by modulation in the inflammatory process by blocking the NF- $\kappa$ B pathway, decreasing the expression of the COX-2 genes (81%), iNOS (100%), IL-1 $\beta$  (93%), IL-6 (88%), and TNF- $\alpha$  (88%) (M. Q. de Souza et al., 2018b; Velagapudi et al., 2018b). The 100% inhibition of the iNOS gene activity is confirmed by the reduction in the generation of nitric oxide (NO), an essential molecule in the oxidative stress process and produced by the action of the enzyme nitric oxide synthase expressed by the transcription of iNOS (Abdolahi et al., 2019).

IL-1  $\beta$  and TNF- $\alpha$  play a central role in the innate immune system, being inducers in the differentiation of monocytic-phagocytic system with increased dendritic cells (DC), macrophages of type M1, activation of neutrophils, and natural clonal cell expansion killer (NK) (Bent et al., 2018). In addition to the immunomodulatory action of phytochemicals in cashews, the compounds contribute to increased SIRT 1 protein in microglial cells. Proteins are weakly expressed in AD inducing oligomerization of A $\beta$  peptides, a ROS-generating phenotype and recognized by the pathophysiological neurotoxicity of the disease (Velagapudi et al., 2018b).

At the tumor level, the mechanism of anticancer action of phytochemicals is still not well understood. However, some hypotheses point to dysregulation in the signaling of proteins involved in the migration, proliferation, maturation, and colonization of tumor cells by activating apoptotic flags (Ex. Caspase-8) to reduce metastasis and tumor mass (Blanco-Vaca et al., 2018). *A. occidentale* compounds demonstrate the antitumor activity by inducing the apoptotic mechanism via caspase-3

activation (Luiz Gomes et al., 2018b) and mitotic block, with depolymerization of the microtubules in the phase of the interphase, compromising the alignment and chromosomal division (Barros et al., 2020; M Ashraf & Rathinasamy, 2018b) that results in the disruption of cell proliferation.

In HeLa cells treated with extracts of *A. occidentale*, morphological changes are observed with loss of 40 and 73% of tumor cell viability at concentrations below 0.01% (M Ashraf & Rathinasamy, 2018b). In composite tumor cells derived from Cardonol, it presented activity in IC<sub>50</sub> in 0.12 to 42 µg / mL, with more significant action in renal tumor cells (Braga et al., 2021b). According to the American National Cancer Institute, compounds with anticancer activity, in pre-clinical trials, must have an IC<sub>50</sub> <30µg / mL to be considered a promising herbal medicine in palliative cancer treatment (Aini et al., 2008).

### ***Eugenia dysenterica***

*Eugenia dysenterica* (Mart.) DC, belonging to the Myrtaceae family, is a tree found in the Atlantic Forest, Cerrado and Caatinga, popularly known as “cagaita” or “cagaiteira”(Cardoso et al., 2011). Compared to other species in the Cerrado, the use of fresh fruit is limited, with losses as it is highly perishable. It is necessary to encourage the use of the fruit in new technological processes for its use (de Sousa et al., 2018).

The fruits and leaves have various bioactive compounds, such as polyphenols, proanthocyanidins, flavonoids, quercetin, and catechins, guaranteeing different biological activities as shown in Table 1. The extracts obtained from the pulp stand out for improving glucose homeostasis with the α-glucosidase enzyme inhibition mechanism (Donado-Pestana et al., 2018a; Justino et al., 2020b), promoting slower absorption of dietary carbohydrates and minimizing glycemic pikes (Dornas et al., 2009).

Studies that evaluate leaf extracts emphasize antioxidant activity, as in work carried out by [56], which aimed to evaluate the neuroprotective potential of the hydroalcoholic extract of *E. dysenterica* leaves, *in vitro* and *in vivo*, and found that the extract could protect the brain against damage induced by oxidation.

Another study, also using the hydroalcoholic extract of the leaves, attested the *in vitro* antioxidant activity capable of reducing reactive oxygen species and the chelating action that prevented damage induced by toxic metals. In the *in vivo* test, animals exposed to chromium and pretreated with the extract showed a reduction in liver and kidney damage and lower concentrations of the metal in these organs and the plasma (Ávila et al., 2016b).

In addition to the activities presented, the essential oil obtained from the leaves of *E. dysenterica* had an anti-inflammatory effect by inhibiting the production of excess nitric oxide within the cell (S. M. M. Da Silva et al., 2019b). Nitric oxide is a fundamental mediator, but when in excess, they form free radicals, such as superoxide, causing peroxynitrite synthesis, a reactive species with great oxidative potential and related to various inflammatory diseases (Eming et al., 2017).

### ***Hancornia speciosa***

Popularly known as “magabeira”, *Hancornia speciosa* Gomes is a Brazilian native tree belonging to the Apocynaceae family. It is distributed in different Brazilian regions comprising some phytogeographic domains, including the Caatinga. Mangaba fruits can be consumed fresh or processed in jams, sweets, ice cream, juices, syrups, and others. In addition to the fruits, this tree can provide byproducts with pharmacological potential extracted from different parts such as stem and leaves (De Almeida et al., 2016).

The leaf extract composition can vary according to climatic conditions and the form of extraction, but some constituents are familiar, such as carotenoids, polyunsaturated fatty acids, phenolic compounds, and flavonoids (Table 1) which can act as bioactive compounds. Bornesitol, a cyclitol isolated from an ethanolic extract of the *H. speciosa* leaves, was used by Moreira et al. (2019) in normotensive Wistar rats to investigate the ability of this compound to reduce blood pressure.

Results demonstrated that the administration of bornesitol reduced blood arterial pressure in normotensive rats, increased the plasma level of nitrite, and decreased the angiotensin-converting enzyme activity while in the aorta, the cyclitol induced endothelium-dependent dilatation (Moreira et al., 2019b). The results described by the authors corroborate the traditional use of *H. speciosa* to reduce blood pressure, thus demonstrating the antihypertensive potential of this plant.

The ethanolic extract also demonstrated antioxidant activity, which occurred by the scavenging of free radicals, hemolysis inhibition, and lipid peroxidation inhibition in human erythrocytes; it can be explained by the presence of a high concentration of phenolic compounds found in the extract of the leaves, such as rutin, a flavonoid identified as a major compound by the authors (U. P. Dos Santos et al., 2018b; U. P. Santos, et al., 2016). The leaves of *H. speciosa* were considered safe according to microbiological quality parameters, indicating the safety of their use as a pharmacological potential (U. P. Dos Santos et al., 2018a).

A study using the frozen pulps of *H. speciosa* fruits also showed the antioxidant activity by free phenolic compounds and their bioaccessibility after exposure to simulated gastrointestinal conditions. The activity was demonstrated by the high iron reduction capacity, and rutin was the free phenolic compound that demonstrated an increase in their bioaccessible amount after exposure to simulated gastric conditions; rutin is generated from quercetin by breaking the bond with sugar when exposed to acidic conditions, which may justify its increase after simulating gastric conditions (Dutra et al., 2017b).

In addition, the aqueous extract from the fruits of *H. speciosa* presented the rutin and chlorogenic acid as the main secondary metabolites, molecules that can be in part responsible for the anti-inflammatory activity described by the authors. The aqueous extract administered at various concentrations was able to reduce the cell recruitment into the peritoneal cavity of mice and inhibited the production of cytokines; it also was capable of reducing the ear edema, indicating a possible anti-phlogistic effect suggesting their use as an alternative option for treating inflammatory disorders (Torres-Rêgo et al., 2016b).

### ***Schinus terebinthifolius***

The red mastic (*Schinus terebinthifolius* Raddi) is a native Brazilian plant with a high association with alternative, socio-cultural and botanical therapies. According to popular knowledge, mastic fractions are applied to the treatment of urinary and respiratory infections, wounds and skin ulcers, tumors, diarrhea and arthritis (P. D. S. da Rocha et al., 2020b), antidiabetic, antinociceptive, and anti-inflammatory (Feriani et al., 2020b), endodontic treatment (Pinto et al., 2020b), angiogenic and keratitis (M. D. C. L. Lima et al., 2020b). The compounds demonstrated high antioxidant activity in vivo and in vitro models, with  $\alpha$ -pinene and  $\alpha$ -phellandrene being reported with high bioactivity (Salem et al., 2018a).

The methanolic extract of the leaves of *S. terebinthifolius* obtained high concentrations of antioxidant compounds, evaluated by DPPH assay, in an animal model; results point to reducing oxidative stress and inhibiting cardiotoxicity induced by the drug doxorubicin (P. D. S. da Rocha et al., 2020b). In another study, the leaves methanolic extracts inhibited the action of  $\alpha$ -glucosidase, an enzyme responsible for the rapid breakdown of polysaccharides in the intestinal wall and consequently its rapid ingestion (dos Santos da Rocha et al., 2019a). The extract also had greater functionality with the reference drug for the treatment of diabetes, metformin, at the concentrations tested in the work; with reduced postprandial glycemia after glucose overload in diabetic mice, decreased liver weight, blood glucose, and reduced serum glycated hemoglobin, aspartate transaminase, and alanine transaminase levels. (dos Santos da Rocha et al., 2019a).

In metabolic conditions such as type 2 diabetes, the levels of natural enzymatic and non-enzymatic antioxidants (vitamin E and C) are reduced, compromising the physiological balance of ROS production antioxidants, significantly increasing biomarkers related to oxidative stress. This state is possibly inhibited by the presence of phenolic groups (Ciocoiu et al., 2009).

Hyperglycemia is a favorable condition for increasing the generation of ROS. The chronic exposure of  $\beta$  cells to a high glucose rate and low content of antioxidant enzymes results in a significant increase in  $H_2O_2$ , causing damage to  $\beta$  cells most susceptible to the deleterious effects of ROS, with the involvement of self-oxidation, oxidative phosphorylation, and glycosylation (Rehman & Akash, 2017). In patients with type 1 diabetes, insulin resistance is correlated with TNF- $\alpha$  (Farinha et al., 2018). As previously discussed, TNF- $\alpha$  transcription is regulated by translocating the NF /  $K\beta$  from the cytoplasm to the nucleus by accumulating ROS.

#### 4. Final Considerations

This work aimed to survey the biological potential of some species in the Caatinga domain without initially considering a specific biological activity. The results showed a greater propensity to search for antioxidant and anti-inflammatory activity extracts from various materials, such as leaves, fruits, stems, and roots. It is known that innumerable pathological dysfunctions present at the core of the aggravation of diseases and the imbalance in the immunological condition. Many studies point to the correlation of antioxidant actives in the control of pathologies such as diabetes, neurodegenerative diseases, and metabolic disorders, aiming to control the immune response of these diseases by reducing the concentration of ROS and the consequent reduction in the pro-inflammatory stimulus. As noted, the Caatinga biome is a source of antioxidative and functional bioactive capable of regulating biological dysfunctions, being recognized as a potential nutraceutical resource.

In this way, we highlight the importance of defending the Caatinga and encouraging new experimental research to evaluate dosages and duration of use to obtain health benefits and elucidate the knowledge of folk medicine in this uniquely Brazilian biome.

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#### References

- Abdel-Aty, A. M., Hamed, M. B., Salama, W. H., Ali, M. M., Fahmy, A. S., & Mohamed, S. A. (2019). Ficus carica, Ficus sycomorus and Euphorbia tirucalli latex extracts: Phytochemical screening, antioxidant and cytotoxic properties. *Biocatalysis and Agricultural Biotechnology*, 20(April), 101199. <https://doi.org/10.1016/j.bcab.2019.101199>
- Abdollahi, M., Jafarieh, A., Sarraf, P., Sedighyan, M., Yousefi, A., Tafakhori, A., Abdollahi, H., Salehinia, F., & Djalali, M. (2019). The Neuromodulatory Effects of  $\omega$ -3 Fatty Acids and Nano-Curcumin on the COX-2/ iNOS Network in Migraines: A Clinical Trial Study from Gene Expression to Clinical Symptoms. *Endocrine, Metabolic & Immune Disorders - Drug Targets*, 19(6), 874–884. <https://doi.org/10.2174/1871530319666190212170140>
- Agita, A., & Thaha, M. (2017). Agita A., Alsagaff M.T. Inflammation, Immunity, and Hypertension. *Acta. Med. Indones.* 2017; 49:158–165. *Acta Medica Indonesiana*, 49(2), 158–165.
- Aini, A. S. N., Merrina, A., Stanslas, J., & Sreeramana, S. (2008). Cytotoxic Potential on Breast Cancer Cells Using Selected Forest Species Found in Malaysia. *International Journal of Cancer Research*, 4(3), 103–109. <https://doi.org/10.3923/ijcr.2008.103.109>
- Amorim, M., Pereira, J. O., Silva, L. B., Ormense, R. C. S. C., Pacheco, M. T. B., & Pintado, M. (2018). Use of whey peptide fraction in coated cashew nut as functional ingredient and salt replacer. *LWT - Food Science and Technology*, 92, 204–211. <https://doi.org/10.1016/j.lwt.2017.12.075>
- Anjos, M. N. V., de Araújo-Neto, L. N., Buonafina, M. D. S., Neves, R. P., de Souza, E. R., Bezerra, I. C. F., Ferreira, M. R. A., Soares, L. A. L., Coutinho, H. D. M., Martins, N., da Silva, M. V., & Correia, M. T. dos S. (2020). *Ocotea glomerata* (Nees) mez extract and fractions: Chemical characterization, anti-Candida activity and related mechanism of action. *Antibiotics*, 9(7), 1–12. <https://doi.org/10.3390/antibiotics9070394>
- Ansari, M. Y., Ahmad, N., & Haqqi, T. M. (2020). Oxidative stress and inflammation in osteoarthritis pathogenesis: Role of polyphenols. In *Biomedicine and Pharmacotherapy* (Vol. 129). Elsevier Masson SAS. <https://doi.org/10.1016/j.biopha.2020.110452>
- Aumeeruddy-Elalfi, Z., Gurib-Fakim, A., & Mahomoodally, F. (2015). Antimicrobial, antibiotic potentiating activity and phytochemical profile of essential oils from exotic and endemic medicinal plants of Mauritius. *Industrial Crops and Products*, 71, 197–204. <https://doi.org/10.1016/j.indcrop.2015.03.058>



- Ávila, R. I. de, Mattos Alvarenga, C. B., Ávila, P. H. M. de, Moreira, R. C., Arruda, A. F., Fernandes, T. de O., Rodrigues, B. dos S., Andrade, W. M., Batista, A. C., Paula, J. R. de, & Valadares, M. C. (2016a). *Eugenia dysenterica* DC. (Myrtaceae) exerts chemopreventive effects against hexavalent chromium-induced damage in vitro and in vivo. *Pharmaceutical Biology*, 54(11), 2652–2663. <https://doi.org/10.1080/13880209.2016.1178306>
- Ávila, R. I. de, Mattos Alvarenga, C. B., Ávila, P. H. M. de, Moreira, R. C., Arruda, A. F., Fernandes, T. de O., Rodrigues, B. dos S., Andrade, W. M., Batista, A. C., Paula, J. R. de, & Valadares, M. C. (2016b). *Eugenia dysenterica* DC. (Myrtaceae) exerts chemopreventive effects against hexavalent chromium-induced damage in vitro and in vivo. *Pharmaceutical Biology*, 54(11), 2652–2663. <https://doi.org/10.1080/13880209.2016.1178306>
- Awakan, O. J., Malomo, S. O., Adejare, A. A., Igunnu, A., Atolani, O., Adebayo, A. H., & Owoyele, B. V. (2018). Anti-inflammatory and bronchodilatory constituents of leaf extracts of *Anacardium occidentale* L. in animal models. *Journal of Integrative Medicine*, 16(1), 62–70. <https://doi.org/10.1016/j.joim.2017.12.009>
- Barros, A. B., Moura, A. F., Silva, D. A., Oliveira, T. M., Barreto, F. S., Ribeiro, W. L. C., Alves, A. P. N. N., Araújo, A. J., Moraes Filho, M. O., Iles, B., Medeiros, J. V. R., & Marinho-Filho, J. D. B. (2020). Evaluation of antitumor potential of cashew gum extracted from *Anacardium occidentale* Linn. *International Journal of Biological Macromolecules*, 154(2020), 319–328. <https://doi.org/10.1016/j.ijbiomac.2020.03.096>
- Belcaro, G., Saggino, A., Cornelli, U., Luzzi, R., Dugall, M., Hosoi, M., Feragalli, B., & Cesarone, M. R. (2018). Improvement in mood, oxidative stress, fatigue, and insomnia following supplementary management with Robuvit®. *Journal of Neurosurgical Sciences*, 62(4), 423–427. <https://doi.org/10.23736/S0390-5616.18.04384-9>
- Bent, R., Moll, L., Grabbe, S., & Bros, M. (2018). Interleukin-1 beta—A friend or foe in malignancies? *International Journal of Molecular Sciences*, 19(8). <https://doi.org/10.3390/ijms19082155>
- Bezerra Filho, C. M., da Silva, L. C. N., da Silva, M. V., Løbner-Olesen, A., Struve, C., Krogfelt, K. A., Correia, M. T. dos S., & Vilela Oliva, M. L. (2020). Antimicrobial and Antivirulence Action of *Eugenia brejoensis* Essential Oil in vitro and in vivo Invertebrate Models. *Frontiers in Microbiology*, 11(March), 1–11. <https://doi.org/10.3389/fmicb.2020.00424>
- Bitencourt, M. A. O., Torres-Rêgo, M., de Souza Lima, M. C. J., Furtado, A. A., de Azevedo, E. P., do Egito, E. S. T., da Silva-Júnior, A. A., Zucolotto, S. M., & Fernandes-Pedrosa, M. de F. (2019). Protective effect of aqueous extract, fractions and phenolic compounds of *Hancornia speciosa* fruits on the inflammatory damage in the lungs of mice induced by *Tityus serrulatus* envenomation. *Toxicon*, 164, 1–9. <https://doi.org/10.1016/j.toxicon.2019.03.018>
- Blanco-Vaca, F., Cedó, L., & Julve, J. (2018). Phytosterols in Cancer: From Molecular Mechanisms to Preventive and Therapeutic Potentials. *Current Medicinal Chemistry*, 26(37), 6735–6749. <https://doi.org/10.2174/0929867325666180607093111>
- Bodoira, R., Rossi, Y., Montenegro, M., Maestri, D., & Velez, A. (2017). Extraction of antioxidant polyphenolic compounds from peanut skin using water-ethanol at high pressure and temperature conditions. *Journal of Supercritical Fluids*, 128, 57–65. <https://doi.org/10.1016/j.supflu.2017.05.011>
- Borges, J. (2021). Cashew tree (*Anacardium occidentale*): Possible applications in dermatology. *Clinics in Dermatology*, xxx, 2020–2022. <https://doi.org/10.1016/j.clinidermatol.2020.11.014>
- Braga, F. C., Ojeda, M., Perdomo, R. T., de Albuquerque, S., Rafique, J., de Lima, D. P., & Beatriz, A. (2021a). Synthesis of cardanol-based 1,2,3-triazoles as potential green agents against neoplastic cells. *Sustainable Chemistry and Pharmacy*, 20(February), 2–7. <https://doi.org/10.1016/j.scp.2021.100408>
- Braga, F. C., Ojeda, M., Perdomo, R. T., de Albuquerque, S., Rafique, J., de Lima, D. P., & Beatriz, A. (2021b). Synthesis of cardanol-based 1,2,3-triazoles as potential green agents against neoplastic cells. *Sustainable Chemistry and Pharmacy*, 20(February), 2–7. <https://doi.org/10.1016/j.scp.2021.100408>
- Cardoso, L. de M., Martino, H. S. D., Moreira, A. V. B., Ribeiro, S. M. R., & Pinheiro-Sant’Ana, H. M. (2011). Cagaita (*Eugenia dysenterica* DC.) of the Cerrado of Minas Gerais, Brazil: Physical and chemical characterization, carotenoids and vitamins. *Food Research International*, 44(7), 2151–2154. <https://doi.org/10.1016/j.foodres.2011.03.005>
- Carvalho, D. V., Santos, F. A., de Lima, R. P., Viana, A. F. S. C., Fonseca, S. G. C., Nunes, P. I. G., de Melo, T. S., Gallão, M. I., & de Brito, E. S. (2018). Influence of low molecular weight compounds associated to cashew (*Anacardium occidentale* L.) fiber on lipid metabolism, glycemia and insulinemia of normal mice. *Bioactive Carbohydrates and Dietary Fibre*, 13(July), 1–6. <https://doi.org/10.1016/j.bcdf.2017.12.001>
- Catelan, T. B. S., Santos Radai, J. A., Leitão, M. M., Branquinho, L. S., Vasconcelos, P. C. de P., Heredia-Vieira, S. C., Kassuya, C. A. L., & Cardoso, C. A. L. (2018). Evaluation of the toxicity and anti-inflammatory activities of the infusion of leaves of *Campomanesia guazumifolia* (Cambess.) O. Berg. *Journal of Ethnopharmacology*, 226, 132–142. <https://doi.org/10.1016/j.jep.2018.08.015>
- Chen, C. Y., Kao, C. L., & Liu, C. M. (2018). The cancer prevention, anti-inflammatory and anti-oxidation of bioactive phytochemicals targeting the TLR4 signaling pathway. *International Journal of Molecular Sciences*, 19(9). <https://doi.org/10.3390/ijms19092729>
- Ciocoiu, M., Mirón, A., Mares, L., Tutunaru, D., Pohaci, C., Groza, M., & Badescu, M. (2009). The effects of *Sambucus nigra* polyphenols on oxidative stress and metabolic disorders in experimental diabetes mellitus. *Journal of Physiology and Biochemistry*, 65(3), 297–304. <https://doi.org/10.1007/BF03180582>
- Cordeiro, B. M. P. C., Carvalho Junior, A. R., Santos, J. R. A., Araújo, A. D., Silva, A. G., Correia, M. T. S., Silva, M. V., Napoleão, T. H., Silva, L. C. N., Santos, N. D. L., & Paiva, P. M. G. (2020). Anticryptococcal activity of hexane extract from *Spondias tuberosa* Arruda and associated cellular events. *Journal de Mycologie Medicale*, 30(2). <https://doi.org/10.1016/j.mycmed.2020.100965>
- Cossetin, J. F., da Silva Brum, E., Casoti, R., Camponogara, C., Dornelles, R. C., Maziero, M., Tatiane de David Antoniazzi, C., Guex, C. G., Ramos, A. P., Pintos, F. G., Engelmann, A. M., Melazzo de Andrade, C., Manfron, M. P., Oliveira, S. M., de Freitas Bauermann, L., Sagrillo, M. R., Machado, A. K., Soares Santos, A. R., & Trevisan, G. (2019). Peanut leaf extract has antioxidant and anti-inflammatory activity but no acute toxic effects. *Regulatory Toxicology and Pharmacology*, 107, 104407. <https://doi.org/10.1016/j.yrtph.2019.104407>
- Costa, W. K., Oliveira, J. R. S. de, Oliveira, A. M. de, Santos, I. B. da S., Cunha, R. X. da, Freitas, A. F. S. de, Silva, J. W. L. M. da, Silva, V. B. G., Aguiar, J. C. R. de O. F. de, Silva, A. G. da, Navarro, D. M. do A. F., Lima, V. L. de M., & Silva, M. V. da. (2020). Essential oil from *Eugenia stipitata* McVaugh leaves

has antinociceptive, anti-inflammatory and antipyretic activities without showing toxicity in mice. *Industrial Crops and Products*, 144(August 2019), 112059. <https://doi.org/10.1016/j.indcrop.2019.112059>

da Rocha, P. D. S., Paula, V. M. B., Olinto, S. C. F., Dos Santos, E. L., Souza, K. de P., & Estevinho, L. M. (2020a). Diversity, chemical constituents and biological activities of endophytic fungi isolated from *Schinus terebinthifolius* raddi. *Microorganisms*, 8(6), 1–13. <https://doi.org/10.3390/microorganisms8060859>

da Rocha, P. D. S., Paula, V. M. B., Olinto, S. C. F., Dos Santos, E. L., Souza, K. de P., & Estevinho, L. M. (2020b). Diversity, chemical constituents and biological activities of endophytic fungi isolated from *Schinus terebinthifolius* raddi. *Microorganisms*, 8(6), 1–13. <https://doi.org/10.3390/microorganisms8060859>

da Silva Barbosa, D. C., Holanda, V. N., de Assis, C. R. D., de Oliveira Farias de Aguiar, J. C. R., do Nascimento, P. H., da Silva, W. V., do Amaral Ferraz Navarro, D. M., Silva, M. V. da, de Menezes Lima, V. L., & dos Santos Correia, M. T. (2020a). Chemical composition and acetylcholinesterase inhibitory potential, in silico, of *Myrciaria floribunda* (H. West ex Willd.) O. Berg fruit peel essential oil. *Industrial Crops and Products*, 151(November 2019), 112372. <https://doi.org/10.1016/j.indcrop.2020.112372>

da Silva Barbosa, D. C., Holanda, V. N., de Assis, C. R. D., de Oliveira Farias de Aguiar, J. C. R., do Nascimento, P. H., da Silva, W. V., do Amaral Ferraz Navarro, D. M., Silva, M. V. da, de Menezes Lima, V. L., & dos Santos Correia, M. T. (2020b). Chemical composition and acetylcholinesterase inhibitory potential, in silico, of *Myrciaria floribunda* (H. West ex Willd.) O. Berg fruit peel essential oil. *Industrial Crops and Products*, 151, 112372. <https://doi.org/10.1016/j.indcrop.2020.112372>

da Silva Brito, S. S., Silva, F., Malheiro, R., Baptista, P., & Pereira, J. A. (2018). *Croton argyrophyllus* Kunth and *Croton heliotropiifolius* Kunth: Phytochemical characterization and bioactive properties. *Industrial Crops and Products*, 113, 308–315. <https://doi.org/10.1016/j.indcrop.2018.01.044>

da Silva, G. C., de Veras, B. O., de Assis, C. R. D., Navarro, D. M. do A. F., Diniz, D. L. V., Brayner dos Santos, F. A., de Aguiar, J. C. R. de O. F., da Silva, M. V., & dos Santos Correia, M. T. (2020). Chemical composition, antimicrobial activity and synergistic effects with conventional antibiotics under clinical isolates by essential oil of *Hymenaea rubriflora* Ducke (FABACEAE). *Natural Product Research*, 0(0), 1–5. <https://doi.org/10.1080/14786419.2020.1729150>

da Silva, J. M. C., & Lacher, T. E. (2020). Caatinga—South America. In *Encyclopedia of the World's Biomes*. Elsevier Inc. <https://doi.org/10.1016/b978-0-12-409548-9.11984-0>

Da Silva, S. M. M., Costa, C. R. R., Gelfuso, G. M., Guerra, E. N. S., De Medeiros Nóbrega, Y. K., Gomes, S. M., Pic-Taylor, A., Fonseca-Bazzo, Y. M., Silveira, D., & De Oliveira Magalhães, P. (2019a). Wound healing effect of essential oil extracted from *eugenia dysenterica* DC (Myrtaceae) leaves. *Molecules*, 24(1). <https://doi.org/10.3390/molecules24010002>

Da Silva, S. M. M., Costa, C. R. R., Gelfuso, G. M., Guerra, E. N. S., De Medeiros Nóbrega, Y. K., Gomes, S. M., Pic-Taylor, A., Fonseca-Bazzo, Y. M., Silveira, D., & De Oliveira Magalhães, P. (2019b). Wound healing effect of essential oil extracted from *eugenia dysenterica* DC (Myrtaceae) leaves. *Molecules*, 24(1). <https://doi.org/10.3390/molecules24010002>

D'abadia, P. L., Bailão, E. F. L. C., Lino Júnior, R. S., Oliveira, M. G., Silva, V. B., Oliveira, L. A. R., Conceição, E. C., Melo-Reis, P. R., Luiborges, L., Gonçalves, P. J., & Almeida, L. M. (2020). *Hancornia speciosa* serum fraction latex stimulates the angiogenesis and extracellular matrix remodeling processes. *Anais Da Academia Brasileira de Ciencias*, 92(2), 1–17. <https://doi.org/10.1590/0001-3765202020190107>

De Almeida, L. M., Nogueira, C. A., Borges, P. P., Do Prado, A. D. L., & Gonçalves, P. J. (2016). State Of The Art Of Scientific Literature On *Hancornia Speciosa*: Trends And Gaps. *Revista Brasileira de Fruticultura*, 38(4). <https://doi.org/10.1590/0100-29452016869>

de Araújo, F. F., de Paulo Farias, D., Neri-Numa, I. A., Dias-Audibert, F. L., Delafiori, J., de Souza, F. G., Catharino, R. R., do Sacramento, C. K., & Pastore, G. M. (2021). Influence of high-intensity ultrasound on color, chemical composition and antioxidant properties of araçá-boi pulp. *Food Chemistry*, 338, 127747. <https://doi.org/10.1016/j.foodchem.2020.127747>

de Araújo, J. S. C., de Castilho, A. R. F., Lira, A. B., Pereira, A. V., de Azevêdo, T. K. B., de Brito Costa, E. M. de M., Pereira, M. do S. V., Pessoa, H. F. L., & Pereira, J. V. (2018). Antibacterial activity against cariogenic bacteria and cytotoxic and genotoxic potential of *Anacardium occidentale* L. and *Anadenanthera macrocarpa* (Benth.) Brenan extracts. *Archives of Oral Biology*, 85(May 2017), 113–119. <https://doi.org/10.1016/j.archoralbio.2017.10.008>

de Araújo Rodrigues, P., de Moraes, S. M., Aguiar, L. A., Vila-Nova, N. S., & Benjamin, S. R. (2019). Effect of *Byrsonima sericea* DC. leaf extracts on mice gastrointestinal tract. *Toxicology Reports*, 6, 1182–1187. <https://doi.org/10.1016/j.toxrep.2019.10.018>

de Azevedo, M. M. L., Cascaes, M. M., Guilhon, G. M. S. P., Andrade, E. H. A., Zoghbi, M. das G. B., da Silva, J. K. R., Santos, L. S., & da Silva, S. H. M. (2019). Lupane triterpenoids, antioxidant potential and antimicrobial activity of *Myrciaria floribunda* (H. West ex Willd.) O. Berg. *Natural Product Research*, 33(4), 506–515. <https://doi.org/10.1080/14786419.2017.1402311>

de Lavor, É. M., Leal, A. E. B. P., Fernandes, A. W. C., Ribeiro, F. P. R. de A., Barbosa, J. de M., Gama e Silva, M., Teles, R. B. de A., Oliveira, L. F. da S., Silva, J. C., Rolim, L. A., de Menezes, I. R. A., & Almeida, J. R. G. da S. (2018). Ethanolic extract of the aerial parts of *Passiflora cincinnata* Mast. (Passifloraceae) reduces nociceptive and inflammatory events in mice. *Phytomedicine*, 47, 58–68. <https://doi.org/10.1016/j.phymed.2018.04.052>

de Lima, M. de F. R., Cavalcante, L. A., Costa, E. C. T. de A., de Veras, B. O., da Silva, M. V., Cavalcanti, L. N., & Araújo, R. M. (2021). Bioactivity flavonoids from roots of *Euphorbia tirucalli* L. *Phytochemistry Letters*, 41(January), 186–192. <https://doi.org/10.1016/j.phytol.2020.10.017>

de Moura Barbosa, H., Amaral, D., do Nascimento, J. N., Machado, D. C., de Sousa Araújo, T. A., de Albuquerque, U. P., Guedes da Silva Almeida, J. R., Rolim, L. A., Lopes, N. P., Gomes, D. A., & Lira, E. C. (2018). *Spondias tuberosa* inner bark extract exert antidiabetic effects in streptozotocin-induced diabetic rats. *Journal of Ethnopharmacology*, 227, 248–257. <https://doi.org/10.1016/j.jep.2018.08.038>

de Oliveira Silva Ribeiro, F., de França Dourado, F., Silva, M. F. S., Brito, L. M., Pessoa, C., de Lima, L. R. M., de Paula, R. C. M., de Souza de Almeida Leite, J. R., de Araújo, A. R., & da Silva, D. A. (2020). Anti-proliferative profile of *Anacardium occidentale* polysaccharide and characterization by AFM. *International Journal of Biological Macromolecules*, 156, 981–987. <https://doi.org/10.1016/j.ijbiomac.2020.03.145>

- de Oliveira Yamashita, F., Torres-Rêgo, M., dos Santos Gomes, J. A., Félix-Silva, J., Ramos Passos, J. G., de Santis Ferreira, L., da Silva-Júnior, A. A., Zucolotto, S. M., & Fernandes-Pedrosa, M. de F. (2020). Mangaba (*Hancornia speciosa* Gomes) fruit juice decreases acute pulmonary edema induced by *Tityus serrulatus* venom: Potential application for auxiliary treatment of scorpion stings. *Toxicon*, 179, 42–52. <https://doi.org/10.1016/j.toxicon.2020.02.025>
- de Oliveira-Júnior, R. G., Alves Ferraz, C. A., Pontes, M. C., Cavalcante, N. B., da Cruz Araújo, E. C., de Oliveira, A. P., Picot, L., Rolim, L. A., & da Silva Almeida, J. R. G. (2018). Antibacterial activity of terpenoids isolated from *Cnidoscolus quercifolius* Pohl (Euphorbiaceae), a Brazilian medicinal plant from Caatinga biome. *European Journal of Integrative Medicine*, 24, 30–34. <https://doi.org/10.1016/j.eujim.2018.10.011>
- de Sousa, E. R. B., Camilo, Y. M. V., & Vera, R. (2018). Cagaita— *Eugenia dysenterica*. *Exotic Fruits*, 77–83. <https://doi.org/10.1016/b978-0-12-803138-4.00011-3>
- de Souza, L. S., Puziol, L. C., Tosta, C. L., Bittencourt, M. L. F., Ardisson, J. S., Kitagawa, R. R., Filgueiras, P. R., & Kuster, R. M. (2019). Analytical methods to access the chemical composition of an *Euphorbia tirucalli* anticancer latex from traditional Brazilian medicine. *Journal of Ethnopharmacology*, 237(January), 255–265. <https://doi.org/10.1016/j.jep.2019.03.041>
- de Souza, M. Q., Teotônio, I. M. S. N., de Almeida, F. C., Heyn, G. S., Alves, P. S., Romeiro, L. A. S., Pratesi, R., de Medeiros Nóbrega, Y. K., & Pratesi, C. B. (2018a). Molecular evaluation of anti-inflammatory activity of phenolic lipid extracted from cashew nut shell liquid (CNSL). *BMC Complementary and Alternative Medicine*, 18(1), 1–11. <https://doi.org/10.1186/s12906-018-2247-0>
- de Souza, M. Q., Teotônio, I. M. S. N., de Almeida, F. C., Heyn, G. S., Alves, P. S., Romeiro, L. A. S., Pratesi, R., de Medeiros Nóbrega, Y. K., & Pratesi, C. B. (2018b). Molecular evaluation of anti-inflammatory activity of phenolic lipid extracted from cashew nut shell liquid (CNSL). *BMC Complementary and Alternative Medicine*, 18(1), 1–11. <https://doi.org/10.1186/s12906-018-2247-0>
- de Souza, R. O., de Assis Dias Alves, G., Aguilera, A. L. S., Rogez, H., & Fonseca, M. J. V. (2018). Photochemoprotective effect of a fraction of a partially purified extract of *Byrsonima crassifolia* leaves against UVB-induced oxidative stress in fibroblasts and hairless mice. *Journal of Photochemistry and Photobiology B: Biology*, 178, 53–60. <https://doi.org/10.1016/j.jphotobiol.2017.10.033>
- de Souza Silva, G., da Silva Campelo Borges, G., Pinho da Costa Castro, C. D., de Tarso Aidar, S., Telles Biasoto Marques, A., Tonetto de Freitas, S., Poloni Rybka, A. C., & Cardarelli, H. R. (2020). Physicochemical quality, bioactive compounds and in vitro antioxidant activity of a new variety of passion fruit cv. BRS Sertão Forte (*Passiflora cincinnata* Mast.) from Brazilian Semiarid region. *Scientia Horticulturae*, 272, 109595. <https://doi.org/10.1016/j.scienta.2020.109595>
- de Veras, B. O., de Oliveira, J. R. S., de Menezes Lima, V. L., do Amaral Ferraz Navarro, D. M., de Oliveira Farias de Aguiar, J. C. R., de Medeiros Moura, G. M., da Silva, J. W., de Assis, C. R. D., Gorchach-Lira, K., de Assis, P. A. C., de Souza Barbosa, J. I., de Melo, M. R. C. S., de Oliveira, M. B. M., da Silva, M. V., & de Souza Lopes, A. C. (2021). The essential oil of the leaves of *Verbesina macrophylla* (Cass.) S.F.Blake has antimicrobial, anti-inflammatory and antipyretic activities and is toxicologically safe. *Journal of Ethnopharmacology*, 265, 113248. <https://doi.org/10.1016/j.jep.2020.113248>
- do Nascimento, K. F., Moreira, F. M. F., Alencar Santos, J., Kassuya, C. A. L., Croda, J. H. R., Cardoso, C. A. L., Vieira, M. do C., Góis Ruiz, A. L. T., Ann Foglio, M., de Carvalho, J. E., & Formagio, A. S. N. (2018). Antioxidant, anti-inflammatory, antiproliferative and antimycobacterial activities of the essential oil of *Psidium guineense* Sw. and spathulenol. *Journal of Ethnopharmacology*, 210, 351–358. <https://doi.org/10.1016/j.jep.2017.08.030>
- Donado-Pestana, C. M., dos Santos-Donado, P. R., Daza, L. D., Belchior, T., Festuccia, W. T., & Genovese, M. I. (2018a). Cagaita fruit (*Eugenia dysenterica* DC.) and obesity: Role of polyphenols on already established obesity. *Food Research International*, 103, 40–47. <https://doi.org/10.1016/j.foodres.2017.10.011>
- Donado-Pestana, C. M., dos Santos-Donado, P. R., Daza, L. D., Belchior, T., Festuccia, W. T., & Genovese, M. I. (2018b). Cagaita fruit (*Eugenia dysenterica* DC.) and obesity: Role of polyphenols on already established obesity. *Food Research International*, 103, 40–47. <https://doi.org/10.1016/j.foodres.2017.10.011>
- Dornas, W. C., De Oliveira, T. T., Dores, R. G. R., Fabres, M. H. A., & Nagem, T. J. (2009). Antidiabetic effects of the medicinal plants. *Brazilian Journal of Pharmacognosy*, 19(2 A), 488–500. <https://doi.org/10.1590/S0102-695X2009000300024>
- dos Santos, A. L., Polidoro, A. dos S., Cardoso, C. A. L., Batistote, M., do Carmo Vieira, M., Jacques, R. A., & Caramão, E. B. (2019). GC×GC/qMS analyses of *Campomanesia guazumifolia* (Cambess.) O. Berg essential oils and their antioxidant and antimicrobial activity. *Natural Product Research*, 33(4), 593–597. <https://doi.org/10.1080/14786419.2017.1399383>
- dos Santos da Rocha, P., de Araújo Boleti, A. P., do Carmo Vieira, M., Carollo, C. A., da Silva, D. B., Estevinho, L. M., dos Santos, E. L., & de Picoli Souza, K. (2019a). Microbiological quality, chemical profile as well as antioxidant and antidiabetic activities of *Schinus terebinthifolius* Raddi. *Comparative Biochemistry and Physiology Part - C: Toxicology and Pharmacology*, 220(February), 36–46. <https://doi.org/10.1016/j.cbpc.2019.02.007>
- dos Santos da Rocha, P., de Araújo Boleti, A. P., do Carmo Vieira, M., Carollo, C. A., da Silva, D. B., Estevinho, L. M., dos Santos, E. L., & de Picoli Souza, K. (2019b). Microbiological quality, chemical profile as well as antioxidant and antidiabetic activities of *Schinus terebinthifolius* Raddi. *Comparative Biochemistry and Physiology Part - C: Toxicology and Pharmacology*, 220, 36–46. <https://doi.org/10.1016/j.cbpc.2019.02.007>
- Dos Santos, E., Radai, J. A. S., do Nascimento, K. F., Formagio, A. S. N., de Matos Balsalobre, N., Ziff, E. B., Castelon Konkiewitz, E., & Kassuya, C. A. L. (2020). Contribution of spathulenol to the anti-nociceptive effects of *Psidium guineense*. *Nutritional Neuroscience*. <https://doi.org/10.1080/1028415X.2020.1815330>
- dos Santos, G. H. F., Amaral, A., & da Silva, E. B. (2018). Antibacterial activity of irradiated extracts of *Anacardium occidentale* L. on multiresistant strains of *Staphylococcus aureus*. *Applied Radiation and Isotopes*, 140, 327–332. <https://doi.org/10.1016/j.apradiso.2018.07.035>
- Dos Santos, U. P., Tolentino, G. S., Morais, J. S., De Picoli Souza, K., Estevinho, L. M., & Dos Santos, E. L. (2018a). Physicochemical characterization, microbiological quality and safety, and pharmacological potential of *Hancornia speciosa* gomes. *Oxidative Medicine and Cellular Longevity*, 2018. <https://doi.org/10.1155/2018/2976985>
- Dos Santos, U. P., Tolentino, G. S., Morais, J. S., De Picoli Souza, K., Estevinho, L. M., & Dos Santos, E. L. (2018b). Physicochemical characterization, microbiological quality and safety, and pharmacological potential of *Hancornia speciosa* gomes. *Oxidative Medicine and Cellular Longevity*, 2018. <https://doi.org/10.1155/2018/2976985>

- Duong, T. H., Beniddir, M. A., Genta-Jouve, G., Nguyen, H. H., Nguyen, D. P., Nguyen, T. A. T., Mac, D. H., Boustie, J., Nguyen, K. P. P., Chavasiri, W., & Le Pogam, P. (2019). Further terpenoids from *Euphorbia tirucalli*. *Fitoterapia*, *135*(April), 44–51. <https://doi.org/10.1016/j.fitote.2019.04.001>
- Dutra, R. L. T., Dantas, A. M., Marques, D. de A., Batista, J. D. F., Meireles, B. R. L. de A., de Magalhães Cordeiro, Â. M. T., Magnani, M., & Borges, G. da S. C. (2017a). Bioaccessibility and antioxidant activity of phenolic compounds in frozen pulps of Brazilian exotic fruits exposed to simulated gastrointestinal conditions. *Food Research International*, *100*, 650–657. <https://doi.org/10.1016/j.foodres.2017.07.047>
- Dutra, R. L. T., Dantas, A. M., Marques, D. de A., Batista, J. D. F., Meireles, B. R. L. de A., de Magalhães Cordeiro, Â. M. T., Magnani, M., & Borges, G. da S. C. (2017b). Bioaccessibility and antioxidant activity of phenolic compounds in frozen pulps of Brazilian exotic fruits exposed to simulated gastrointestinal conditions. *Food Research International*, *100*, 650–657. <https://doi.org/10.1016/j.foodres.2017.07.047>
- Eming, S. A., Wynn, T. A., & Martin, P. (2017). Inflammation and metabolism in tissue repair and regeneration. *Science*, *356*(6342), 1026–1030. <https://doi.org/10.1126/science.aam7928>
- Estevão, L. R. M., Simões, R. S., Cassini-Vieira, P., Canesso, M. C. C., Barcelos, L. da S., Rachid, M. A., da Câmara, C. A. G., & Evêncio-Neto, J. (2017). *Schinus terebinthifolius raddi* (aroeira) leaves oil attenuates inflammatory responses in cutaneous wound healing in mice. *Acta Cirúrgica Brasileira*, *32*(9), 726–735. <https://doi.org/10.1590/s0102-865020170090000005>
- Eungsuwan, N., Chayjarung, P., Pankam, J., Pilaisangsuree, V., Wongshaya, P., Kongbangkerd, A., Sriphannam, C., & Limmongkon, A. (2021a). Production and antimicrobial activity of trans-resveratrol, trans-arachidin-1 and trans-arachidin-3 from elicited peanut hairy root cultures in shake flasks compared with bioreactors. *Journal of Biotechnology*, *326*, 28–36. <https://doi.org/10.1016/j.jbiotec.2020.12.006>
- Eungsuwan, N., Chayjarung, P., Pankam, J., Pilaisangsuree, V., Wongshaya, P., Kongbangkerd, A., Sriphannam, C., & Limmongkon, A. (2021b). Production and antimicrobial activity of trans-resveratrol, trans-arachidin-1 and trans-arachidin-3 from elicited peanut hairy root cultures in shake flasks compared with bioreactors. *Journal of Biotechnology*, *326*, 28–36. <https://doi.org/10.1016/j.jbiotec.2020.12.006>
- Farinha, J. B., Ramis, T. R., Vieira, A. F., Macedo, R. C. O., Rodrigues-Krause, J., Boeno, F. P., Schroeder, H. T., Müller, C. H., Boff, W., Krause, M., De Bittencourt, P. I. H., & Reischak-Oliveira, A. (2018). Glycemic, inflammatory and oxidative stress responses to different high-intensity training protocols in type 1 diabetes: A randomized clinical trial. *Journal of Diabetes and Its Complications*, *32*(12), 1124–1132. <https://doi.org/10.1016/j.jdiacomp.2018.09.008>
- Feriani, A., Tir, M., Arafah, M., Gómez-Caravaca, A. M., Contreras, M. del M., Nahdi, S., Taamalli, A., Allagui, M. S., Alwasel, S., Segura-Carretero, A., Harrath, A. H., & Tlili, N. (2021). *Schinus terebinthifolius* fruits intake ameliorates metabolic disorders, inflammation, oxidative stress, and related vascular dysfunction, in atherogenic diet-induced obese rats. Insight of their chemical characterization using HPLC-ESI-QTOF-MS/MS. *Journal of Ethnopharmacology*, *269*. <https://doi.org/10.1016/j.jep.2020.113701>
- Feriani, A., Tir, M., Hamed, M., Sila, A., Nahdi, S., Alwasel, S., Harrath, A. H., & Tlili, N. (2020a). Multidirectional insights on polysaccharides from *Schinus terebinthifolius* and *Schinus molle* fruits: Physicochemical and functional profiles, in vitro antioxidant, anti-genotoxicity, antidiabetic, and antihemolytic capacities, and in vivo anti-inflammatory and anti-nociceptive properties. *International Journal of Biological Macromolecules*, *165*(Pt B), 2576–2587. <https://doi.org/10.1016/j.ijbiomac.2020.10.123>
- Feriani, A., Tir, M., Hamed, M., Sila, A., Nahdi, S., Alwasel, S., Harrath, A. H., & Tlili, N. (2020b). Multidirectional insights on polysaccharides from *Schinus terebinthifolius* and *Schinus molle* fruits: Physicochemical and functional profiles, in vitro antioxidant, anti-genotoxicity, antidiabetic, and antihemolytic capacities, and in vivo anti-inflammatory and anti-nociceptive properties. *International Journal of Biological Macromolecules*, *165*(Pt B), 2576–2587. <https://doi.org/10.1016/j.ijbiomac.2020.10.123>
- Fernandes, A. C. F., Vieira, N. C., Santana, Á. L. de, Gandra, R. L. de P., Rubia, C., Castro-Gamboa, I., Macedo, J. A., & Macedo, G. A. (2020). Peanut skin polyphenols inhibit toxicity induced by advanced glycation end-products in RAW264.7 macrophages. *Food and Chemical Toxicology*, *145*, 111619. <https://doi.org/10.1016/j.fct.2020.111619>
- Ferreira-Fernandes, H., Barros, M. A. L., Souza Filho, M. D., Medeiros, J. V. R., Vasconcelos, D. F. P., Silva, D. A., Leódidio, A. C. M., Silva, F. R. P., França, L. F. C., Di Lenardo, D., Yoshioka, F. K. N., Rey, J. A., Burbano, R. R., & Pinto, G. R. (2019). Topical application of cashew gum or chlorhexidine gel reduces overexpression of proinflammatory genes in experimental periodontitis. *International Journal of Biological Macromolecules*, *128*, 934–940. <https://doi.org/10.1016/j.ijbiomac.2019.02.002>
- Fidelis-de-Oliveira, P., Aparecida-Castro, S., Silva, D. B., Morais, I. B. de M., Miranda, V. H. M. de, de Gobbi, J. I., Canabrava, H. A. N., & Bispo-da-Silva, L. B. (2020). Hypotensive effect of *Eugenia dysenterica* leaf extract is primarily related to its vascular action: The possible underlying mechanisms. *Journal of Ethnopharmacology*, *251*. <https://doi.org/10.1016/j.jep.2019.112520>
- Fukui, M., & Zhu, B. T. (2010). Mitochondrial superoxide dismutase SOD2, but not cytosolic SOD1, plays a critical role in protection against glutamate-induced oxidative stress and cell death in HT22 neuronal cells. *Free Radical Biology and Medicine*, *48*(6), 821–830. <https://doi.org/10.1016/j.freeradbiomed.2009.12.024>
- Gasca, C. A., Castillo, W. O., Takahashi, C. S., Fagg, C. W., Magalhães, P. O., Fonseca-Bazzo, Y. M., & Silveira, D. (2017). Assessment of anticholinesterase activity and cytotoxicity of cagaita (*Eugenia dysenterica*) leaves. *Food and Chemical Toxicology*, *109*(Pt 2), 996–1002. <https://doi.org/10.1016/j.fct.2017.02.032>
- Gershenzon, J., & Dudareva, N. (2007). The function of terpene natural products in the natural world. *Nature Chemical Biology*, *3*(7), 408–414. <https://doi.org/10.1038/nchembio.2007.5>
- Gomes Júnior, A. L., Tchekalarova, J. D., Machado, K. da C., Moura, A. K. S., Paz, M. F. C. J., da Mata, A. M. O. F., Nogueira, T. R., Islam, M. T., Rios, M. A. de S., Graças Lopes Citó, A. M. das, Uddin, S. J., Shilpi, J. A., Das, A. K., Lopes, L. da S., & Melo-Cavalcante, A. A. de C. (2018a). Anxiolytic effect of anacardic acids from cashew (*Anacardium occidentale*) nut shell in mice. *IUBMB Life*, *December 2017*, 1–12. <https://doi.org/10.1002/iub.1738>

- Gomes Júnior, A. L., Tchekalarova, J. D., Machado, K. da C., Moura, A. K. S., Paz, M. F. C. J., da Mata, A. M. O. F., Nogueira, T. R., Islam, M. T., Rios, M. A. de S., Graças Lopes Citó, A. M. das, Uddin, S. J., Shilpi, J. A., Das, A. K., Lopes, L. da S., & Melo-Cavalcante, A. A. de C. (2018b). Anxiolytic effect of anacardic acids from cashew (*Anacardium occidentale*) nut shell in mice. *IUBMB Life, December 2017*, 1–12. <https://doi.org/10.1002/iub.1738>
- Goulart da Silva, G., de Oliveira Braga, L. E., Souza de Oliveira, E. C., Valério Tinti, S., de Carvalho, J. E., Goldoni Lazarini, J., Rosalen, P. L., Dionísio, A. P., & Tasca Gois Ruiz, A. L. (2021a). Cashew apple byproduct: Gastroprotective effects of standardized extract. *Journal of Ethnopharmacology*, 269(September 2020). <https://doi.org/10.1016/j.jep.2020.113744>
- Goulart da Silva, G., de Oliveira Braga, L. E., Souza de Oliveira, E. C., Valério Tinti, S., de Carvalho, J. E., Goldoni Lazarini, J., Rosalen, P. L., Dionísio, A. P., & Tasca Gois Ruiz, A. L. (2021b). Cashew apple byproduct: Gastroprotective effects of standardized extract. *Journal of Ethnopharmacology*, 269(September 2020). <https://doi.org/10.1016/j.jep.2020.113744>
- Graf, B. L., Zhang, L., Corradini, M. G., Kuhn, P., Newman, S. S., Salbaum, J. M., & Raskin, I. (2018). Physicochemical differences between malanga (*Xanthosoma sagittifolium*) and potato (*Solanum tuberosum*) tubers are associated with differential effects on the gut microbiome. *Journal of Functional Foods*, 45, 268–276. <https://doi.org/10.1016/j.jff.2018.04.032>
- Ibrahim, E. H., Kilany, M., Mostafa, O. M. S., Shaker, K. H., Alshehri, M., Alsaad, K. M., Alshehri, A., Khan, K. A., Qasim, M., Kotb, N., Alahmari, A. S., Ghramh, H. A., & Dajem, S. M. (2019). TH1/TH2 chemokines/cytokines profile in rats treated with tetanus toxoid and *Euphorbia tirucalli*. *Saudi Journal of Biological Sciences*, 26(7), 1716–1723. <https://doi.org/10.1016/j.sjbs.2018.08.005>
- Jia, X., Wang, W., Chen, Z., He, Y., & Liu, J. (2014). Concentrations of secondary metabolites in tissues and root exudates of wheat seedlings changed under elevated atmospheric CO<sub>2</sub> and cadmium-contaminated soils. *Environmental and Experimental Botany*, 107, 134–143. <https://doi.org/10.1016/j.envexpbot.2014.06.005>
- Junsathian, P., Yordtong, K., Corpuz, H. M., Katayama, S., Nakamura, S., & Rawdkuen, S. (2018). Biological and neuroprotective activity of Thai edible plant extracts. *Industrial Crops and Products*, 124(April), 548–554. <https://doi.org/10.1016/j.indcrop.2018.08.008>
- Justino, A. B., de Moura, F. R. B., Franco, R. R., & Espindola, F. S. (2020a).  $\alpha$ -Glucosidase and non-enzymatic glycation inhibitory potential of *Eugenia dysenterica* fruit pulp extracts. *Food Bioscience*, 35, 100573. <https://doi.org/10.1016/j.fbio.2020.100573>
- Justino, A. B., de Moura, F. R. B., Franco, R. R., & Espindola, F. S. (2020b).  $\alpha$ -Glucosidase and non-enzymatic glycation inhibitory potential of *Eugenia dysenterica* fruit pulp extracts. *Food Bioscience*, 35, 100573. <https://doi.org/10.1016/j.fbio.2020.100573>
- Kim, S. H., Yuk, H. J., Ryu, H. W., Oh, S. R., Song, D. Y., Lee, K. S., Park, K. I., Choi, S. W., & Seo, W. D. (2019a). Biofunctional soyasaponin Bb in peanut (*Arachis hypogaea* L.) sprouts enhances bone morphogenetic protein-2-dependent osteogenic differentiation via activation of runt-related transcription factor 2 in C2C12 cells. *Phytotherapy Research*, 33(5). <https://doi.org/10.1002/ptr.6341>
- Kim, S. H., Yuk, H. J., Ryu, H. W., Oh, S. R., Song, D. Y., Lee, K. S., Park, K. I., Choi, S. W., & Seo, W. D. (2019b). Biofunctional soyasaponin Bb in peanut (*Arachis hypogaea* L.) sprouts enhances bone morphogenetic protein-2-dependent osteogenic differentiation via activation of runt-related transcription factor 2 in C2C12 cells. *Phytotherapy Research*, 33(5), 1490–1500. <https://doi.org/10.1002/ptr.6341>
- Kouitcheu Mabeku, L. B., Eyoum Bille, B., & Nguépi, E. (2016). In Vitro and In Vivo Anti- Helicobacter Activities of *Eryngium foetidum* (Apiaceae), *Bidens pilosa* (Asteraceae), and *Galinsoga ciliata* (Asteraceae) against *Helicobacter pylori*. *BioMed Research International*, 2016. <https://doi.org/10.1155/2016/2171032>
- Larrauri, M., Zunino, M. P., Zygodlo, J. A., Grosso, N. R., & Nepote, V. (2016). Chemical characterization and antioxidant properties of fractions separated from extract of peanut skin derived from different industrial processes. *Industrial Crops and Products*, 94, 964–971. <https://doi.org/10.1016/j.indcrop.2016.09.066>
- Leal, A. E. B. P., de Oliveira, A. P., Santos, R. F. dos, Soares, J. M. D., Lavor, E. M. de, Pontes, M. C., Lima, J. T. de, Santos, A. D. da C., Tomaz, J. C., Oliveira, G. G. de, Neto, F. C., Lopes, N. P., Rolim, L. A., & Almeida, J. R. G. da S. (2020). Determination of phenolic compounds, in vitro antioxidant activity and characterization of secondary metabolites in different parts of *Passiflora cincinnata* by HPLC-DAD-MS/MS analysis. *Natural Product Research*, 34(7), 995–1001. <https://doi.org/10.1080/14786419.2018.1548445>
- Lepetsos, P., Papavassiliou, K. A., & Papavassiliou, A. G. (2019). Redox and NF- $\kappa$ B signaling in osteoarthritis. In *Free Radical Biology and Medicine* (Vol. 132, pp. 90–100). Elsevier Inc. <https://doi.org/10.1016/j.freeradbiomed.2018.09.025>
- Lima, A. B., Delwing-de Lima, D., Vieira, M. R., Poletto, M. Z., Delwing-Dal Magro, D., Barauna, S. C., Alberton, M. D., Pereira, E. M., Pereira, N. R., Salamaia, E. M., & Siebert, D. A. (2017). Hypolipemiant and antioxidant effects of *Eugenia brasiliensis* in an animal model of coconut oil-induced hypertriglyceridemia. *Biomedicine and Pharmacotherapy*, 96, 642–649. <https://doi.org/10.1016/j.biopha.2017.10.047>
- Lima, M. D. C. L., De Araújo, J. I. F., Gonçalves Mota, C., Magalhães, F. E. A., Campos, A. R., Da Silva, P. T., Rodrigues, T. H. S., Matos, M. G. C., De Sousa, K. C., De Sousa, M. B., Saker-Sampaio, S., Pereira, A. L., Teixeira, E. H., & Dos Santos, H. S. (2020a). Antinociceptive Effect of the Essential Oil of *Schinus terebinthifolius* (female) Leaves on Adult Zebrafish (*Danio rerio*). *Zebrafish*, 17(2), 112–119. <https://doi.org/10.1089/zeb.2019.1809>
- Lima, M. D. C. L., De Araújo, J. I. F., Gonçalves Mota, C., Magalhães, F. E. A., Campos, A. R., Da Silva, P. T., Rodrigues, T. H. S., Matos, M. G. C., De Sousa, K. C., De Sousa, M. B., Saker-Sampaio, S., Pereira, A. L., Teixeira, E. H., & Dos Santos, H. S. (2020b). Antinociceptive Effect of the Essential Oil of *Schinus terebinthifolius* (female) Leaves on Adult Zebrafish (*Danio rerio*). *Zebrafish*, 17(2), 112–119. <https://doi.org/10.1089/zeb.2019.1809>
- Limmongkon, A., Nopprang, P., Chaikandee, P., Somboon, T., Wongshaya, P., & Pilaisangsree, V. (2018). LC-MS/MS profiles and interrelationships between the anti-inflammatory activity, total phenolic content and antioxidant potential of Kalasin 2 cultivar peanut sprout crude extract. *Food Chemistry*, 239, 569–578. <https://doi.org/10.1016/j.foodchem.2017.06.162>
- Limmongkon, A., Pankam, J., Somboon, T., Wongshaya, P., & Nopprang, P. (2019). Evaluation of the DNA damage protective activity of the germinated peanut (*Arachis hypogaea*) in relation to antioxidant and anti-inflammatory activity. *LWT*, 101. <https://doi.org/10.1016/j.lwt.2018.11.009>

- Lira, S. M., Canabrava, N. V., Benjamin, S. R., Silva, J. Y. G., Viana, D. A., Lima, C. L. S., Paredes, P. F. M., Marques, M. M. M., Pereira, E. O., Queiroz, E. A. M., & Guedes, M. I. F. (2017). Evaluation of the toxicity and hypoglycemic effect of the aqueous extracts of *cnidoscolus quercifolius* pohl. *Brazilian Journal of Medical and Biological Research*, 50(10). <https://doi.org/10.1590/1414-431x20176361>
- Locatelli, M., Avato, P., Herman, F., Westfall, S., Brathwaite, J., Pasinetti, G. M., & Peters, J. J. (2018). *Suppression of Presymptomatic Oxidative Stress and Inflammation in Neurodegeneration by Grape-Derived Polyphenols*. <https://doi.org/10.3389/fphar.2018.00867>
- López-Gutiérrez, N., Romero-González, R., Plaza-Bolaños, P., Martínez Vidal, J. L., & Garrido Frenich, A. (2015). Identification and quantification of phytochemicals in nutraceutical products from green tea by UHPLC-Orbitrap-MS. *Food Chemistry*, 173, 607–618. <https://doi.org/10.1016/j.foodchem.2014.10.092>
- Luiz Gomes, A., Dimitrova Tchekalarova, J., Atanasova, M., da Conceição Machado, K., de Sousa Rios, M. A., Paz Jardim, M. F., Găman, M. A., Găman, A. M., Yele, S., Shill, M. C., Khan, I. N., Islam, M. A., Ali, E. S., Mishra, S. K., Islam, M. T., Mubarak, M. S., da Silva Lopes, L., & de Carvalho Melo-Cavalcante, A. A. (2018a). Anticonvulsant effect of anacardic acid in murine models: Putative role of GABAergic and antioxidant mechanisms. *Biomedicine and Pharmacotherapy*, 106(July), 1686–1695. <https://doi.org/10.1016/j.biopha.2018.07.121>
- Luiz Gomes, A., Dimitrova Tchekalarova, J., Atanasova, M., da Conceição Machado, K., de Sousa Rios, M. A., Paz Jardim, M. F., Găman, M. A., Găman, A. M., Yele, S., Shill, M. C., Khan, I. N., Islam, M. A., Ali, E. S., Mishra, S. K., Islam, M. T., Mubarak, M. S., da Silva Lopes, L., & de Carvalho Melo-Cavalcante, A. A. (2018b). Anticonvulsant effect of anacardic acid in murine models: Putative role of GABAergic and antioxidant mechanisms. *Biomedicine and Pharmacotherapy*, 106(July), 1686–1695. <https://doi.org/10.1016/j.biopha.2018.07.121>
- M Ashraf, S., & Rathinasamy, K. (2018a). Antibacterial and anticancer activity of the purified cashew nut shell liquid: implications in cancer chemotherapy and wound healing. *Natural Product Research*, 32(23), 2856–2860. <https://doi.org/10.1080/14786419.2017.1380022>
- M Ashraf, S., & Rathinasamy, K. (2018b). Antibacterial and anticancer activity of the purified cashew nut shell liquid: implications in cancer chemotherapy and wound healing. *Natural Product Research*, 32(23), 2856–2860. <https://doi.org/10.1080/14786419.2017.1380022>
- Martins, C. G., Appel, M. H., Coutinho, D. S. S., Soares, I. P., Fischer, S., de Oliveira, B. C., Fachi, M. M., Pontarolo, R., Bonatto, S. J. R., Fernandes, L. C., Iagher, F., & de Souza, L. M. (2020). Consumption of latex from *Euphorbia tirucalli* L. promotes a reduction of tumor growth and cachexia, and immunomodulation in Walker 256 tumor-bearing rats. *Journal of Ethnopharmacology*, 255(February), 112722. <https://doi.org/10.1016/j.jep.2020.112722>
- Menis Candela, F., Giordano, W. F., Quiroga, P. L., Escobar, F. M., Mañas, F., Roma, D. A., Larrauri, M., Comini, L. R., Soria, E. A., & Sabini, M. C. (2020). Evaluation of cellular safety and the chemical composition of the peanut (*Arachis hypogaea* L.) ethanolic extracts. *Heliyon*, 6(10), e05119. <https://doi.org/10.1016/j.heliyon.2020.e05119>
- Mittal, M., Siddiqui, M. R., Tran, K., Reddy, S. P., & Malik, A. B. (2014). Reactive oxygen species in inflammation and tissue injury. *Antioxidants and Redox Signaling*, 20(7), 1126–1167. <https://doi.org/10.1089/ars.2012.5149>
- Moreira, L. N., Silva, G. C., Câmara, D. V., Pádua, R. M., Lemos, V. S., Braga, F. C., & Cortes, S. F. (2019a). The Cyclitol L(-)-Bornesitol as an Active Marker for the Cardiovascular Activity of the Brazilian Medicinal Plant *Hancornia speciosa*. In *Biol. Pharm. Bull* (Vol. 42, Issue 12).
- Moreira, L. N., Silva, G. C., Câmara, D. V., Pádua, R. M., Lemos, V. S., Braga, F. C., & Cortes, S. F. (2019b). The Cyclitol L(-)-Bornesitol as an Active Marker for the Cardiovascular Activity of the Brazilian Medicinal Plant *Hancornia speciosa*. In *Biol. Pharm. Bull* (Vol. 42, Issue 12).
- Morvin Yabesh, J. E., Vijayakumar, S., Arulmozhi, P., & Rajalakshmi, S. (2019). Screening the antimicrobial potential of twelve medicinal plants against venereal diseases causing pathogens. *Acta Ecologica Sinica*, 39(5), 356–361. <https://doi.org/10.1016/j.chnaes.2018.09.011>
- Oliveira, A. S., Nascimento, J. R., Trovão, L. O., Alves, P. C. S., Maciel, M. C. G., Silva, L. D. M., Marques, A. A., Santos, A. P. S. A., Silva, L. A., Nascimento, F. R. F., & Guerra, R. N. M. (2019). The anti-inflammatory activity of *Anacardium occidentale* L. increases the lifespan of diabetic mice with lethal sepsis. *Journal of Ethnopharmacology*, 236(July 2018), 345–353. <https://doi.org/10.1016/j.jep.2019.03.014>
- Oliveira de Souza, L. I., Bezerra-Silva, P. C., do Amaral Ferraz Navarro, D. M., da Silva, A. G., dos Santos Correia, M. T., da Silva, M. V., & de Figueiredo, R. C. B. Q. (2017). The chemical composition and trypanocidal activity of volatile oils from Brazilian Caatinga plants. *Biomedicine and Pharmacotherapy*, 96, 1055–1064. <https://doi.org/10.1016/j.biopha.2017.11.121>
- Oliveira de Veras, B., Melo de Oliveira, M. B., Granja da Silva Oliveira, F., Queiroz dos Santos, Y., Saturnino de Oliveira, J. R., Lúcia de Menezes Lima, V., Guedes da Silva Almeida, J. R., Maria do Amaral Ferraz Navarro, D., Ribeiro de Oliveira Farias de Aguiar, J. C., Aguiar, J. dos S., Gorlach-Lira, K., Dias de Assis, C. R., Vanusa da Silva, M., & Catarina de Souza Lopes, A. (2020). Chemical composition and evaluation of the antinociceptive, antioxidant and antimicrobial effects of essential oil from *Hymenaea cangaceira* (Pinto, Mansano & Azevedo) native to Brazil: A natural medicine. *Journal of Ethnopharmacology*, 247(September 2019), 112265. <https://doi.org/10.1016/j.jep.2019.112265>
- Oliveira, M. B. S., Valentim, I. B., Rocha, T. S., Santos, J. C., Pires, K. S. N., Tanabe, E. L. L., Borbely, K. S. C., Borbely, A. U., & Goulart, M. O. F. (2020). *Schinus terebinthifolius* Raddi extracts: From sunscreen activity toward protection of the placenta to Zika virus infection, new uses for a well-known medicinal plant. *Industrial Crops and Products*, 152. <https://doi.org/10.1016/j.indcrop.2020.112503>
- Omolaso, B. O., Oluwole, F. S., Odukanmi, O. A., Adesanwo, J. K., Ishola, A. A., & Adewole, K. E. (2020a). Evaluation of the gastrointestinal anti-motility effect of *Anacardium occidentale* stem bark extract: A mechanistic study of antidiarrheal activity. *Journal of Pharmaceutical Analysis*. <https://doi.org/10.1016/j.jpah.2020.06.009>
- Omolaso, B. O., Oluwole, F. S., Odukanmi, O. A., Adesanwo, J. K., Ishola, A. A., & Adewole, K. E. (2020b). Evaluation of the gastrointestinal anti-motility effect of *Anacardium occidentale* stem bark extract: A mechanistic study of antidiarrheal activity. *Journal of Pharmaceutical Analysis*. <https://doi.org/10.1016/j.jpah.2020.06.009>
- Palit, P., Mukherjee, D., Mahanta, P., Shadab, M., Ali, N., Roychoudhury, S., Asad, M., & Mandal, S. C. (2018). Attenuation of nociceptive pain and inflammatory disorders by total steroid and terpenoid fraction of *Euphorbia tirucalli* Linn root in experimental in vitro and in vivo model. *Inflammopharmacology*, 26(1), 235–250. <https://doi.org/10.1007/s10787-017-0403-7>

- Peng, J., Jia, Y., Du, X., Wang, Y., Yang, Z., & Li, K. (2019a). Study of physicochemical stability of anthocyanin extracts from black peanut skin and their digestion enzyme and adipogenesis inhibitory activities. *LWT*, *107*. <https://doi.org/10.1016/j.lwt.2019.03.016>
- Peng, J., Jia, Y., Du, X., Wang, Y., Yang, Z., & Li, K. (2019b). Study of physicochemical stability of anthocyanin extracts from black peanut skin and their digestion enzyme and adipogenesis inhibitory activities. *LWT*, *107*, 107–116. <https://doi.org/10.1016/j.lwt.2019.03.016>
- Peng, M., Chen, Z., Deng, Q., Zhu, S., & Wang, G. (2021). The roles of luteolin in peanut shell extract - Mediated protection of erythrocytes against hypoxanthine-xanthine oxidase-induced toxicity. *Food Bioscience*, *39*, 100826. <https://doi.org/10.1016/j.fbio.2020.100826>
- Pinto, I. C., Seibert, J. B., Pinto, L. S., Santos, V. R., de Sousa, R. F., Sousa, L. R. D., Amparo, T. R., dos Santos, V. M. R., do Nascimento, A. M., de Souza, G. H. B., Vasconcellos, W. A., Vieira, P. M. A., & Andrade, Â. L. (2020a). Preparation of glass-ionomer cement containing ethanolic Brazilian pepper extract (*Schinus terebinthifolius* Raddi) fruits: chemical and biological assays. *Scientific Reports*, *10*(1). <https://doi.org/10.1038/s41598-020-79257-3>
- Pinto, I. C., Seibert, J. B., Pinto, L. S., Santos, V. R., de Sousa, R. F., Sousa, L. R. D., Amparo, T. R., dos Santos, V. M. R., do Nascimento, A. M., de Souza, G. H. B., Vasconcellos, W. A., Vieira, P. M. A., & Andrade, Â. L. (2020b). Preparation of glass-ionomer cement containing ethanolic Brazilian pepper extract (*Schinus terebinthifolius* Raddi) fruits: chemical and biological assays. *Scientific Reports*, *10*(1). <https://doi.org/10.1038/s41598-020-79257-3>
- Prasad, S., Gupta, S. C., & Tyagi, A. K. (2017). Reactive oxygen species (ROS) and cancer: Role of antioxidative nutraceuticals. *Cancer Letters*, *387*, 95–105. <https://doi.org/10.1016/j.canlet.2016.03.042>
- Räisänen, T., Ryyppö, A., & Kellomäki, S. (2008). Effects of elevated CO<sub>2</sub> and temperature on monoterpene emission of Scots pine (*Pinus sylvestris* L.). *Atmospheric Environment*, *42*(18), 4160–4171. <https://doi.org/10.1016/j.atmosenv.2008.01.023>
- Regueira, M. S., Tintino, S. R., da Silva, A. R. P., Costa, M. do S., Boligon, A. A., Matias, E. F. F., de Queiroz Balbino, V., Menezes, I. R. A., & Melo Coutinho, H. D. (2017). Seasonal variation of Brazilian red propolis: Antibacterial activity, synergistic effect and phytochemical screening. *Food and Chemical Toxicology*, *107*, 572–580. <https://doi.org/10.1016/j.fct.2017.03.052>
- Rehman, K., & Akash, M. S. H. (2017). Mechanism of Generation of Oxidative Stress and Pathophysiology of Type 2 Diabetes Mellitus: How Are They Interlinked? *Journal of Cellular Biochemistry*, *118*(11), 3577–3585. <https://doi.org/10.1002/jcb.26097>
- Rhee, S. G., Woo, H. A., Kil, I. S., & Bae, S. H. (2012). Peroxiredoxin functions as a peroxidase and a regulator and sensor of local peroxides. In *Journal of Biological Chemistry* (287(7)), pp. 4403–4410. Elsevier. <https://doi.org/10.1074/jbc.R111.283432>
- Ribeiro, P. P. C., Damasceno, K. S. F. da S. C., de Veras, B. O., de Oliveira, J. R. S., Lima, V. L. de M., de Assis, C. R. D., da Silva, M. V., de Sousa Júnior, F. C., de Assis, C. F., Padilha, C. E. de A., & Stamford, T. C. M. (2021). Chemical and biological activities of faveleira (*Cnidioscolus quercifolius* Pohl) seed oil for potential health applications. *Food Chemistry*, *337*, 127771. <https://doi.org/10.1016/j.foodchem.2020.127771>
- Ribeiro, P. P. C., E Silva, D. M. D. L., De Assis, C. F., Correia, R. T. P., & Damasceno, K. S. F. D. S. C. (2017). Bioactive properties of faveleira (*Cnidioscolus quercifolius*) seeds, oil and press cake obtained during oilseed processing. *PLoS ONE*, *12*(8). <https://doi.org/10.1371/journal.pone.0183935>
- Rocha, P. dos S. da, Campos, J. F., Nunes-Souza, V., Vieira, M. do C., Boleti, A. P. de A., Rabelo, L. A., dos Santos, E. L., & de Picoli Souza, K. (2018a). Antioxidant and Protective Effects of *Schinus terebinthifolius* Raddi Against Doxorubicin-Induced Toxicity. *Applied Biochemistry and Biotechnology*, *184*(3), 869–884. <https://doi.org/10.1007/s12010-017-2589-y>
- Rocha, P. dos S. da, Campos, J. F., Nunes-Souza, V., Vieira, M. do C., Boleti, A. P. de A., Rabelo, L. A., dos Santos, E. L., & de Picoli Souza, K. (2018b). Antioxidant and Protective Effects of *Schinus terebinthifolius* Raddi Against Doxorubicin-Induced Toxicity. *Applied Biochemistry and Biotechnology*, *184*(3), 869–884. <https://doi.org/10.1007/s12010-017-2589-y>
- Rossi, Y. E., Bohl, L. P., Vanden Braber, N. L., Ballatore, M. B., Escobar, F. M., Bodoira, R., Maestri, D. M., Porporatto, C., Cavaglieri, L. R., & Montenegro, M. A. (2020a). Polyphenols of peanut (*Arachis hypogaea* L.) skin as bioprotectors of normal cells. Studies of cytotoxicity, cytoprotection and interaction with ROS. *Journal of Functional Foods*, *67*(February), 103862. <https://doi.org/10.1016/j.jff.2020.103862>
- Rossi, Y. E., Bohl, L. P., Vanden Braber, N. L., Ballatore, M. B., Escobar, F. M., Bodoira, R., Maestri, D. M., Porporatto, C., Cavaglieri, L. R., & Montenegro, M. A. (2020b). Polyphenols of peanut (*Arachis hypogaea* L.) skin as bioprotectors of normal cells. Studies of cytotoxicity, cytoprotection and interaction with ROS. *Journal of Functional Foods*, *67*, 103862. <https://doi.org/10.1016/j.jff.2020.103862>
- Salem, M. Z. M., El-Hefny, M., Ali, H. M., Elansary, H. O., Nasser, R. A., El-Settawy, A. A. A., El Shanhorey, N., Ashmawy, N. A., & Salem, A. Z. M. (2018a). Antibacterial activity of extracted bioactive molecules of *Schinus terebinthifolius* ripened fruits against some pathogenic bacteria. *Microbial Pathogenesis*, *120*, 119–127. <https://doi.org/10.1016/j.micpath.2018.04.040>
- Salem, M. Z. M., El-Hefny, M., Ali, H. M., Elansary, H. O., Nasser, R. A., El-Settawy, A. A. A., El Shanhorey, N., Ashmawy, N. A., & Salem, A. Z. M. (2018b). Antibacterial activity of extracted bioactive molecules of *Schinus terebinthifolius* ripened fruits against some pathogenic bacteria. *Microbial Pathogenesis*, *120*, 119–127. <https://doi.org/10.1016/j.micpath.2018.04.040>
- Santos, V. S. dos, Nascimento, T. V., Felipe, J. L., Boaretto, A. G., Damasceno-Junior, G. A., Silva, D. B., Toffoli-Kadri, M. C., & Carollo, C. A. (2017). Nutraceutical potential of *Byrsonima cydoniifolia* fruits based on chemical composition, anti-inflammatory, and antihyperalgesic activities. *Food Chemistry*, *237*, 240–246. <https://doi.org/10.1016/j.foodchem.2017.05.082>
- Santos, I. B. da S., Santos dos Santos, B., Oliveira, J. R. S. de, Costa, W. K., Zagmignan, A., da Silva, L. C. N., Ferreira, M. R. A., Lermen, V. L., Lermen, M. S. B. de S., da Silva, A. G., Ximenes, R. M., Soares, L. A. L., Paiva, P. M. G., Lima, V. L. de M., Correia, M. T. dos S., & da Silva, M. V. (2020). Antioxidant Action and In Vivo Anti-Inflammatory and Antinociceptive Activities of *Myrciaria floribunda* Fruit Peels: Possible Involvement of Opioidergic System. *Advances in Pharmacological and Pharmaceutical Sciences*, *2020*, 1–11. <https://doi.org/10.1155/2020/1258707>
- Santos, J. M., Cury, N. M., Yunes, J. A., López, J. A., & Hernández-Macedo, M. L. (2019). Effect of *Anacardium occidentale* leaf extract on human acute lymphoblastic leukaemia cell lines. *Natural Product Research*, *33*(11), 1633–1636. <https://doi.org/10.1080/14786419.2018.1425841>

- Santos, U. P., Campos, J. F., Torquato, H. F. V., Paredes-Gamero, E. J., Carollo, C. A., Estevinho, L. M., de Picoli Souza, K., & dos Santos, E. L. (2016). Antioxidant, Antimicrobial and Cytotoxic Properties as Well as the Phenolic Content of the Extract from *Hancornia speciosa* Gomes. *PLOS ONE*, *11*(12), e0167531. <https://doi.org/10.1371/journal.pone.0167531>
- Santos, U. P., Campos, J. F., Torquato, H. F. V., Paredes-Gamero, E. J., Carollo, C. A., Estevinho, L. M., De Picoli Souza, K., & Dos Santos, E. L. (2016). Antioxidant, antimicrobial and cytotoxic properties as well as the phenolic content of the extract from *Hancornia speciosa* gomes. *PLoS ONE*, *11*(12). <https://doi.org/10.1371/journal.pone.0167531>
- Seyyedbrahimi, S. S., Khodabandehloo, H., Nasli Esfahani, E., & Meshkani, R. (2018). The effects of resveratrol on markers of oxidative stress in patients with type 2 diabetes: a randomized, double-blind, placebo-controlled clinical trial. *Acta Diabetologica*, *55*(4), 341–353. <https://doi.org/10.1007/s00592-017-1098-3>
- Sharma, A., Shahzad, B., Rehman, A., Bhardwaj, R., Landi, M., & Zheng, B. (2019). Response of phenylpropanoid pathway and the role of polyphenols in plants under abiotic stress. In *Molecules* (24(13), 2452). MDPI AG. <https://doi.org/10.3390/molecules24132452>
- Siebra, A. L. A., Oliveira, L. R., Martins, A. O. B. P. B., Siebra, D. C., Albuquerque, R. S., Lemos, I. C. S., Delmondes, G. A., Tintino, S. R., Figueredo, F. G., da Costa, J. G. M., Coutinho, H. D. M., Menezes, I. R. A., Felipe, C. F. B., & Kerntopf, M. R. (2018). Potentiation of antibiotic activity by *Passiflora cincinnata* Mast. front of strains *Staphylococcus aureus* and *Escherichia coli*. *Saudi Journal of Biological Sciences*, *25*(1), 37–43. <https://doi.org/10.1016/j.sjbs.2016.01.019>
- Sile, I., Romane, E., Reinsone, S., Maurina, B., Tirezite, D., & Dambrova, M. (2020). Data on medicinal plants in the records of Latvian folk medicine from the 19th century. *Data in Brief*, *28*, 105024. <https://doi.org/10.1016/j.dib.2019.105024>
- Silva, C. P. da, Soares-Freitas, R. A. M., Sampaio, G. R., Santos, M. C. B., do Nascimento, T. P., Cameron, L. C., Ferreira, M. S. L., & Arêas, J. A. G. (2019). Identification and action of phenolic compounds of *Jatobá-do-cerrado* (*Hymenaea stagnocarpa* Mart.) on  $\alpha$ -amylase and  $\alpha$ -glucosidase activities and flour effect on glycemic response and nutritional quality of breads. *Food Research International*, *116*, 1076–1083. <https://doi.org/10.1016/j.foodres.2018.09.050>
- Silva, M. V. da, Pandorfi, H., Lopes, P. M. O., Silva, J. L. B. da, de Almeida, G. L. P., Silva, D. A. de O., Santos, A. dos, Rodrigues, J. A. de M., Batista, P. H. D., & Jardim, A. M. da R. F. (2020). Pilot monitoring of caatinga spatial-temporal dynamics through the action of agriculture and livestock in the Brazilian semiarid. *Remote Sensing Applications: Society and Environment*, *19*, 100353. <https://doi.org/10.1016/j.rsase.2020.100353>
- Silva, V. A. O., Rosa, M. N., Miranda-Gonçalves, V., Costa, A. M., Tansini, A., Evangelista, A. F., Martinho, O., Carloni, A. C., Jones, C., Lima, J. P., Pianowski, L. F., & Reis, R. M. (2019). Euphol, a tetracyclic triterpene, from *Euphorbia tirucalli* induces autophagy and sensitizes temozolomide cytotoxicity on glioblastoma cells. *Investigational New Drugs*, *37*(2), 223–237. <https://doi.org/10.1007/s10637-018-0620-y>
- Simpson, D. S. A., & Oliver, P. L. (2020). Ros generation in microglia: Understanding oxidative stress and inflammation in neurodegenerative disease. *Antioxidants*, *9*(8), 1–27. <https://doi.org/10.3390/antiox9080743>
- Soares, J. C., Rosalen, P. L., Lazarini, J. G., Massarioli, A. P., da Silva, C. F., Nani, B. D., Franchin, M., & de Alencar, S. M. (2019). Comprehensive characterization of bioactive phenols from new Brazilian superfruits by LC-ESI-QTOF-MS, and their ROS and RNS scavenging effects and anti-inflammatory activity. *Food Chemistry*, *281*, 178–188. <https://doi.org/10.1016/j.foodchem.2018.12.106>
- Souza dos Santos, B., Bezerra Filho, C. M., Alves do Nascimento Junior, J. A., Brust, F. R., Bezerra-Silva, P. C., Lino da Rocha, S. K., Krogfelt, K. A., Maria do Amaral Ferraz Navarro, D., Tereza dos Santos Correia, M., Napoleão, T. H., Nascimento da Silva, L. C., Macedo, A. J., Vanusa da Silva, M., & Guedes Paiva, P. M. (2019). Anti-staphylococcal activity of *Syagrus coronata* essential oil: Biofilm eradication and in vivo action on *Galleria mellonella* infection model. *Microbial Pathogenesis*, *131*, 150–157. <https://doi.org/10.1016/j.micpath.2019.04.009>
- Souza Filho, M. D., Medeiros, J. V. R., Vasconcelos, D. F. P., Silva, D. A., Leódoio, A. C. M., Fernandes, H. F., Silva, F. R. P., França, L. F. C., Lenardo, D., & Pinto, G. R. (2018a). Orabase formulation with cashew gum polysaccharide decreases inflammatory and bone loss hallmarks in experimental periodontitis. *International Journal of Biological Macromolecules*, *107*(Part A), 1093–1101. <https://doi.org/10.1016/j.ijbiomac.2017.09.087>
- Souza Filho, M. D., Medeiros, J. V. R., Vasconcelos, D. F. P., Silva, D. A., Leódoio, A. C. M., Fernandes, H. F., Silva, F. R. P., França, L. F. C., Lenardo, D., & Pinto, G. R. (2018b). Orabase formulation with cashew gum polysaccharide decreases inflammatory and bone loss hallmarks in experimental periodontitis. *International Journal of Biological Macromolecules*, *107*(Part A), 1093–1101. <https://doi.org/10.1016/j.ijbiomac.2017.09.087>
- Sunderam, V., Thiyagarajan, D., Lawrence, A. V., Mohammed, S. S. S., & Selvaraj, A. (2019). In-vitro antimicrobial and anticancer properties of green synthesized gold nanoparticles using *Anacardium occidentale* leaves extract. *Saudi Journal of Biological Sciences*, *26*(3), 455–459. <https://doi.org/10.1016/j.sjbs.2018.12.001>
- Taiwo, B. J., Fatokun, A. A., Olubiyi, O. O., Bamigboye-Taiwo, O. T., van Heerden, F. R., & Wright, C. W. (2017). Identification of compounds with cytotoxic activity from the leaf of the Nigerian medicinal plant, *Anacardium occidentale* L. (Anacardiaceae). *Bioorganic and Medicinal Chemistry*, *25*(8), 2327–2335. <https://doi.org/10.1016/j.bmc.2017.02.040>
- Teixeira, É. W., Message, D., Negri, G., Salatino, A., & Stringheta, P. C. (2010). Seasonal Variation, Chemical Composition and Antioxidant Activity of Brazilian Propolis Samples. *Evidence-Based Complementary and Alternative Medicine*, *7*(3), 307–315. <https://doi.org/10.1093/ecam/nem177>
- Thomas, P., Essien, E., Ntuk, S., & Choudhary, M. (2017). *Eryngium foetidum* L. Essential Oils: Chemical Composition and Antioxidant Capacity. *Medicines*, *4*(2), 24. <https://doi.org/10.3390/medicines4020024>
- Thomaz, D. V., Peixoto, L. F., de Oliveira, T. S., Fajemiroye, J. O., da Silva Neri, H. F., Xavier, C. H., Costa, E. A., Alcantara dos Santos, F. C., de Souza Gil, E., & Ghedini, P. C. (2018a). Antioxidant and neuroprotective properties of *Eugenia dysenterica* leaves. *Oxidative Medicine and Cellular Longevity*, *2018*. <https://doi.org/10.1155/2018/8601028>



Thomaz, D. V., Peixoto, L. F., de Oliveira, T. S., Fajemiroye, J. O., da Silva Neri, H. F., Xavier, C. H., Costa, E. A., Alcantara dos Santos, F. C., de Souza Gil, E., & Ghedini, P. C. (2018b). Antioxidant and neuroprotective properties of *Eugenia dysenterica* leaves. *Oxidative Medicine and Cellular Longevity*, 2018. <https://doi.org/10.1155/2018/8601028>

Tietbohl, L. A. C., Oliveira, A. P., Esteves, R. S., Albuquerque, R. D. D. G., Folly, D., Machado, F. P., Corrêa, A. L., Santos, M. G., Ruiz, A. L. G., & Rocha, L. (2017). Antiproliferative activity in tumor cell lines, antioxidant capacity and total phenolic, flavonoid and tannin contents of *Myrciaria floribunda*. *Anais Da Academia Brasileira de Ciencias*, 89(2), 1111–1120. <https://doi.org/10.1590/0001-3765201720160461>

Todirascu-Ciornea, E., El-Nashar, H. A. S., Mostafa, N. M., Eldahshan, O. A., Boiangiu, R. S., Dumitru, G., Hritcu, L., & Singab, A. N. B. (2019). *Schinus terebinthifolius* Essential Oil Attenuates Scopolamine-Induced Memory Deficits via Cholinergic Modulation and Antioxidant Properties in a Zebrafish Model. *Evidence-Based Complementary and Alternative Medicine*, 2019. <https://doi.org/10.1155/2019/5256781>

Torres-Rêgo, M., Furtado, A. A., Bitencourt, M. A. O., Lima, M. C. J. de S., Andrade, R. C. L. C. de, Azevedo, E. P. de, Soares, T. da C., Tomaz, J. C., Lopes, N. P., da Silva-Júnior, A. A., Zucolotto, S. M., & Fernandes-Pedrosa, M. de F. (2016a). Anti-inflammatory activity of aqueous extract and bioactive compounds identified from the fruits of *Hancornia speciosa* Gomes (Apocynaceae). *BMC Complementary and Alternative Medicine*, 16(1), 1–10. <https://doi.org/10.1186/s12906-016-1259-x>

Torres-Rêgo, M., Furtado, A. A., Bitencourt, M. A. O., Lima, M. C. J. de S., Andrade, R. C. L. C. de, Azevedo, E. P. de, Soares, T. da C., Tomaz, J. C., Lopes, N. P., da Silva-Júnior, A. A., Zucolotto, S. M., & Fernandes-Pedrosa, M. de F. (2016b). Anti-inflammatory activity of aqueous extract and bioactive compounds identified from the fruits of *Hancornia speciosa* Gomes (Apocynaceae). *BMC Complementary and Alternative Medicine*, 16(1), 1–10. <https://doi.org/10.1186/s12906-016-1259-x>

Treuter, E., Fan, R., Huang, Z., Jakobsson, T., & Venteclef, N. (2017). Transcriptional repression in macrophages—basic mechanisms and alterations in metabolic inflammatory diseases. *FEBS Letters*, 591(19), 2959–2977. <https://doi.org/10.1002/1873-3468.12850>

Velagapudi, R., Ajileye, O. O., Okorji, U., Jain, P., Aderogba, M. A., & Olajide, O. A. (2018a). Agathisflavone isolated from *Anacardium occidentale* suppresses SIRT1-mediated neuroinflammation in BV2 microglia and neurotoxicity in APPSwe-transfected SH-SY5Y cells. *Phytotherapy Research*, 32(10), 1957–1966. <https://doi.org/10.1002/ptr.6122>

Velagapudi, R., Ajileye, O. O., Okorji, U., Jain, P., Aderogba, M. A., & Olajide, O. A. (2018b). Agathisflavone isolated from *Anacardium occidentale* suppresses SIRT1-mediated neuroinflammation in BV2 microglia and neurotoxicity in APPSwe-transfected SH-SY5Y cells. *Phytotherapy Research*, 32(10), 1957–1966. <https://doi.org/10.1002/ptr.6122>

Yang, L., Yang, C., Li, C., Zhao, Q., Liu, L., Fang, X., & Chen, X. Y. (2016). Recent advances in biosynthesis of bioactive compounds in traditional Chinese medicinal plants. In *Science Bulletin* (Vol. 61, Issue 1, pp. 3–17). Science in China Press. <https://doi.org/10.1007/s11434-015-0929-2>

Zhang, T., Jiang, J., Liu, J., Xu, L., Duan, S., Sun, L., Zhao, W., & Qian, F. (2020). MK2 Is Required for Neutrophil-Derived ROS Production and Inflammatory Bowel Disease. *Frontiers in Medicine*, 7. <https://doi.org/10.3389/fmed.2020.00207>