

## **Contribution to the knowledge of Polypores (Agaricomycetes) in the Amazonian Forest, with 16 new records for the state of Pará, Brazil**

**Contribuição ao conhecimento dos Políporos (Agaricomycetes) na Floresta Amazônica, com 16 novos registros para o estado do Pará, Brasil**

**Contribución al conocimiento de los Poliporos (Agaricomycetes) en la Amazonia, con 16 nuevos registros para el estado de Pará, Brasil**

Received: 10/01/2022 | Revised: 10/11/2022 | Accepted: 10/13/2022 | Published: 10/18/2022

**Douglas de Moraes Couceiro**

ORCID: <https://orcid.org/0000-0003-4189-7478>  
Universidade Federal do Oeste do Pará, Brazil  
E-mail: douglasmcouceiro@gmail.com

**Adriene Mayra Silva Soares**

ORCID: <https://orcid.org/0000-0002-1143-0706>  
Universidade Federal Rural da Amazônia, Brazil  
E-mail: adriene\_soares@yahoo.com.br

**Sheyla Regina Marques Couceiro**

ORCID: <https://orcid.org/0000-0001-8186-4203>  
Universidade Federal do Oeste do Pará, Brazil  
E-mail: sheylacouceiro@yahoo.com.br

### **Abstract**

We present an inventory of polypore fungi (Basidiomycota; Hymenochaetales and Polyporales) with the aim of knowing the diversity of an area of 8 km<sup>2</sup> of Brazilian Amazon rainforest, in the surroundings of HPP Sílvio Braga, in the west of Pará, where 91 species were collected (545 specimens), with 16 of these new records for the state of Pará, and 1 for America. These numbers tend to increase, given the projected occurrence of 118 species for the area. 87 species were considered occasional or rare, with only four considered frequent and none abundant. The richness observed in the study area was similar to other conservation units in the Amazon, for example, Caxiuanã National Forest (74 species). This observation, added to the potential increase in the number of species, the number of species with low representativeness (relative frequency) and the number of new records for the state, demonstrate the need to expand studies on polypore fungi in the region to learn about their biodiversity and the need for conservation of that area.

**Keywords:** Basidiomycota; Brazilian Amazonian; Fungi; Polyporales; Hymenochaetales.

### **Resumo**

Apresentamos um inventário de fungos políporos (Basidiomycota; Hymenochaetales e Polyporales) com o intuito de conhecer a diversidade de uma área de 8 km<sup>2</sup> de Floresta Amazônica brasileira, no entorno da UHE Sílvio Braga, no Oeste paraense, onde foram coletadas 91 espécies (545 espécimes), sendo 16 destes novos registros para o estado do Pará, e 1 para América. Estes números tendem a aumentar, visto a projeção de ocorrência de 118 espécies para a área. A maioria das espécies, 87 foram consideradas ocasionais ou raras, sendo somente quatro consideradas frequentes e nenhuma abundante. A riqueza observada na área estudada foi semelhante a outras unidades de conservação na Amazônia, como por exemplo, Floresta Nacional de Caxiuanã (74 espécies). Essa observação, somada ao potencial incremento no número de espécies, e número de espécies com baixa representatividade (frequência relativa) e número de novos registros para o estado, demonstram a necessidade de ampliação dos estudos sobre os fungos políporos na região para conhecimento de sua biodiversidade e a necessidade de conservação dessa área.

**Palavras-chave:** Basidiomycota; Amazônia brasileira; Fungi; Polyporales; Hymenochaetales.

### **Resumen**

Presentamos un inventario de hongos políporos (Basidiomycota; Hymenochaetales y Polyporales) con el objetivo de conocer la diversidad de un área de 8 km<sup>2</sup> de Selva Amazónica brasileña, en los alrededores de la UHE Sílvio Braga, en el oeste de Pará, donde se encontraron 91 especies recolectados (545 especímenes), con 16 de estos nuevos registros para el estado de Pará y 1 para América. Estos números tienden a aumentar, ya que se proyecta que 118 especies se encuentren en el área. De la mayoría de las especies, 87 fueron consideradas ocasionales o raras, con solo cuatro consideradas frecuentes y ninguna abundante. La riqueza observada en el área de estudio fue similar a otras

unidades de conservación en la Amazonía, como el Bosque Nacional Caxiuanã (74 especies). Esta observación, sumada al aumento potencial en el número de especies, y el número de especies con baja representación (frecuencia relativa) y el número de nuevos registros para el estado, demuestran la necesidad de ampliar los estudios sobre hongos poliporosos en la región para comprender su biodiversidad y la necesidad de conservación de esta área.

**Palabras clave:** Basidiomycota; Amazonía brasileña; Fungi; Polyporales; Hymenochaetales.

## 1. Introduction

Brazilian biodiversity is valued at approximately four trillion dollars, twice the value of the country's gross domestic product (Costanza et al., 2014). The Amazon includes a considerable share of this biodiversity, which is being reduced with the devastation of protected and unprotected areas (Ritter et al., 2017; Singh et al., 2021).

Tropical forests such as the Amazon are ecosystems conducive to the development of a rich assemblage of fungi (Hawksworth & Lücking, 2017), still very little studied, mainly due to the restriction of access to many locations and the limited number of researchers working in the region. Species and their medical and biotechnological properties are disappearing even before they are discovered, causing an inestimable impact on human knowledge (Tabarelli et al., 2004). Thus, a considerable sampling effort is necessary in order to identify ecological patterns and interactions of species of fungi in the Amazon (Gibertoni, 2008; Gibertoni et al., 2016).

Of the 1,050 species of fungi listed for the Brazilian Amazon, 252 are polypores (Basidiomycota, Agaricomycetes), and of these, only 148 species are registered for the state of Pará (Maia et al., 2015). These species are presented by Gomes-Silva et al. (2009) and make up the continuously updated list of Flora do Brasil species (<http://floradobrasil.jbrj.gov.br>).

The Polypores are macroscopic fungi such as tubular/poroid hymenophore (Ryvarden, 2004; Justos et al. 2017), commonly known as shelf-fungi or as polyporoid fungi, the last one is the most common among specialists in the group. This characteristic has evolved several times within the group, being an example of morphological convergence on the hymenial surface (Hibbett & Binder, 2002), in addition to making them key elements in the dynamics and health of any type of forest ecosystem worldwide from the main function to nature which is the decomposition of dead trees standing or fallen (Boddy & Heilmann-Clausen, 2008; Lonsdale et al., 2008; Yang et al. 2021).

Besides the ecological importance, polypores demonstrate biotechnological importance, through the production of cellulose and lignin degrading enzymes, these fungi are used in the textile industry, cleaning tributaries and soil contaminated by oil, pesticide production, among others (Maciel et al., 2010; Lomascolo et al., 2011; Bekai et al. 2012; Tsigain et al. 2022).

Polypore studies in Brazilian Amazon were conducted in the states of Amapá, Amazonas, Pará, Roraima and Rondônia. In Pará, although there are studies on these fungi, all were concentrated at the Estação Científica Ferreira Penna (Caxiuanã National Forest) (1°47'32.3" S, 51°26'2.5" W) (e.g., Sotão et al., 1997, 2003, 2009; Gibertoni, 2008; Gibertoni et al., 2013, and Medeiros et al., 2013, 2015), leaving other areas equally important, little or not sampled and, consequently, there is no study of the diversity of polypores in a forest fragment area. Thus, the present study aimed to inventory the species of polypores and determine the frequency of these species in a fragment of Amazon forest in the western state of Pará, Brazil.

## 2. Methodology

### 2.1 Study area

The collections were carried out in a fragment of Amazonon forest, along the PA-370 highway, near the Silvio Braga Hydropower Plant (HPP) (2°49'11.49" S, 54°17'56.64" W), popularly known as Curuá-Una hydropower, 68 km southeast of the city center of Santarém, Pará state, northern Brazil.

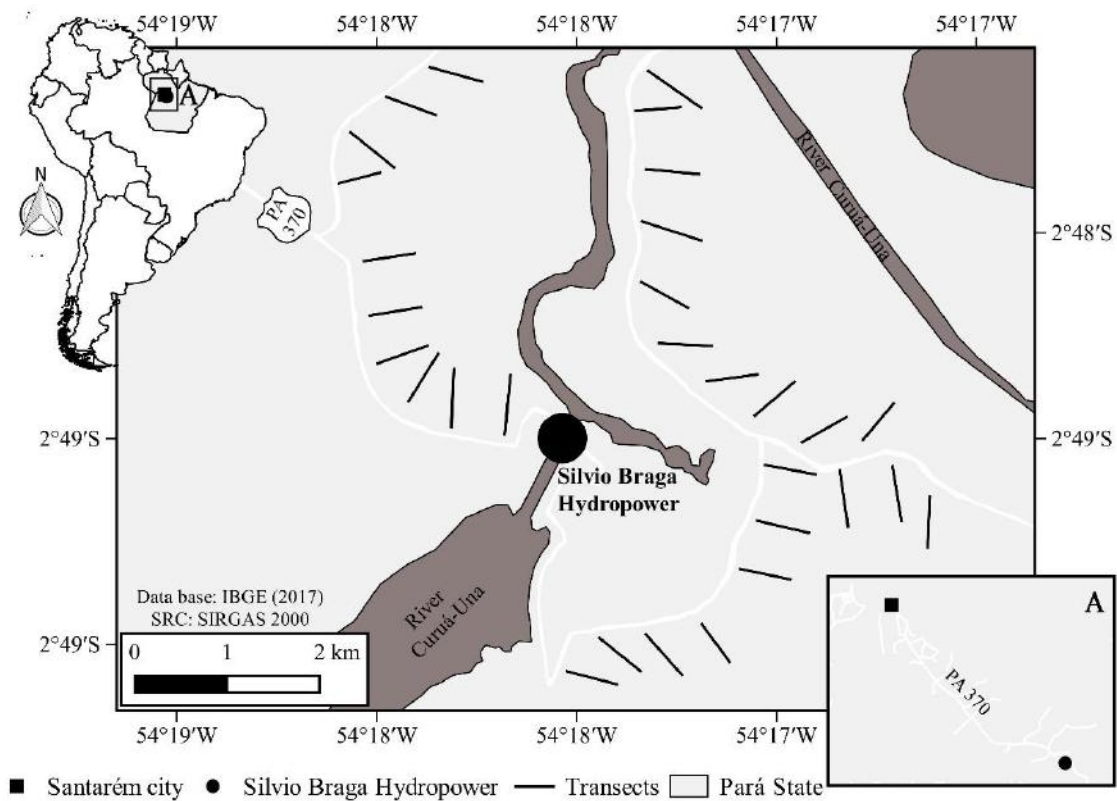
The study area has vegetation cover of dense ombrophylous forest (Veloso et al. 1991) and yellow latosol (Jati & Silva, 2017). Humid tropical climate, with an average temperature of 27 °C ( $\pm$  5 °C). The average relative humidity of the air is

88% and the average annual rainfall is 2,200 mm, with highest rainfall between the months of January to May (rainy season; monthly average of 231 mm) and lower rainfall from August to November (period dry; 61 mm monthly average) (Alvares et al., 2013).

## 2.2 Collecting, herborizing and identifying of polypores

Excursions to collect polypores fungi were carried out between January 29 to February 4, April 23 to 29, July 28 to August 1 and, October 30 to 31, 2018. In each excursion 30 perpendicular transects of 300 m long were covered, equidistant 250 m from each other from the starting point (Figure 1).

**Figure 1** - Location of Silvio Braga HPP (Curuá-Una) with established transects (georeferenced) for the collection of fungi. A) Distance Santarém city for Silvio Braga (HPP).



Source: Authors.

The polypore fungi specimens were photographed with the aid of a digital camera (Nikon COOLPIX L820) still on the occurring substrates (living wood, dead wood and soil) to later be removed manually with the aid of chisels, packed individually in paper bags, with date recording, collector type of substrate and collecting coordinates (Garmin GPS, GPSMAP 64S model).

In the Laboratory, the specimens were dehydrated at 35 °C ( $\pm$  5 °C) in an oven (Ethik, model 404/1D) for a period of two to three days (Fidalgo & Bononi, 1999) and mounted on exsiccates. The polypores were identified by macro and microscopic analysis.

Macroscopic analysis consisted of detailed observations of the basidiomes with the naked eye and/or with the aid of a stereoscopic microscope (Diagtech, model XLT6445T-B2), being analyzed its insertion in the substrate, size (length, width and thickness), color, consistency, characteristics of the surfaces of the pileus and pores, the tubes, context and margin of the

basidioma. The color of the basidioma was determined by comparisons with Kueppers' color chart (Kueppers, 1982). Morphological measurements were carried out with an analog caliper.

For microscopic analysis, sections of different parts of the basidiomes (pileus, context and tubes) were made with the aid of a steel blade, under a binocular optical microscope (Nova, model 107-T). The cuts were arranged between slides and coverslips immersed in different aqueous solutions: 3% potassium hydroxide (hydrating), phloxine, methylene blue (dyes) and Melzer reagent to show the reactions of the walls of the microstructure, which can be positive or negative and, which vary according to each species: hyaline (colorless tone, with visible cell wall), amyloid (blue to purple/violet tone) or dextrinoid (gold to reddish tone) (Teixeira, 1995; Ryvarden, 2004). The hyphal system (monomitic, di-trimitic), reproductive structures (basidia and basidiospores) and sterile structures (cystidia, cystidioles, setae, among others), were also analyzed.

After analysis, the characteristics were compared with the literature in this group of fungi (e.g., Reid, 1965; Ryvarden & Johansen, 1980; Furtado, 1981; Núñez, 1995; Ryvarden, 2004; Dai, 2010; Gomes-Silva et al., 2014, 2015; Ryvarden, 2015, 2016; Costa-Rezende et al., 2016, Palacio et al., 2017). The species classification has been continuously updated according to the Index Fungorum (<http://www.indexfungorum.org>). The identified fungi were deposited at the HSTM Herbarium of the Universidade Federal do Oeste do Pará (UFOPA).

### 2.3 Data analysis

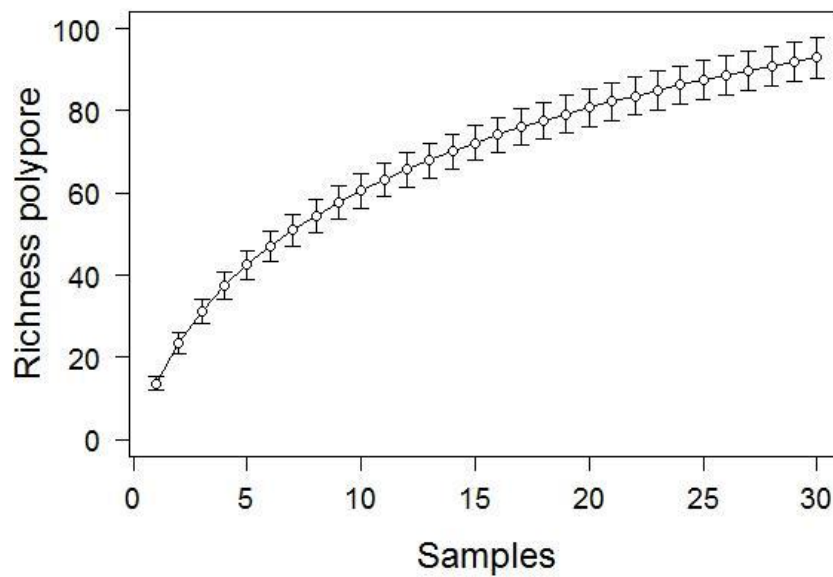
The total and transect diversity was estimated using the Shannon-Wiener index, for reducing the effects of rare species in the sample, considering the relative abundances of individuals per species (Magurran, 2004), the Vegan package (Oksanen et al., 2015) in R 3.5.2 (R Core Team, 2012) was used.

The expected species richness was estimated by Jackknife 1 (Efron & Tibshirani, 1993, Santos, 2004), using the BiodiversityR package (Kindt & Coe 2005) and the *Mao Tau* resampling method (Colwell et al., 2004) to perform the species accumulation curve in the Vegan package (Oksanen et al., 2015). The relative frequency of species ( $F$ ) was calculated using the function:  $F = n / N \times 100$ , where ( $n$ ) is the number of specimens of a species and ( $N$ ) is the total number of specimens found (Urcelay & Robledo, 2004), considering the following classes frequency:  $0.5 \geq F \leq 1.5\%$  = rare;  $1.6 \geq F \leq 5\%$  = occasional;  $5.1 \geq F \leq 9.9\%$  = frequent;  $F \geq 10\%$  = abundant (Soares et al., 2014; Medeiros et al., 2015).

## 3. Results

A total of 545 polypore specimens were collected. In total 8 families, 45 genera and 91 species were represented (Table 1), of which 16 species are new records for the state of Pará, one of them registered for the first time in America (Table 2). The number of records for the state, considering the study area, tends to increase, since the Jackknife 1 richness estimator predicts the occurrence of 118 species in the study area (24% more than was collected) (Figure 2). Shannon's diversity was 3.30, this diversity is more expressive in the transect 2 with a value reaching 3.12 and lower in the transect 16 with a value of 1.10, revealing the distribution of frequent species.

**Figure 2** - Accumulation curve of species of polypore fungi in relation to the number of samples based on the *Mao Tao* method with a 95% confidence interval.



Source: Authors.

Table 1 lists the families and genera with the largest number of representatives, with Polyporaceae being the most representative with 43 species followed by Hymenochaetaceae with 23 species. The most representative genus was *Amauroderma* Murril (Polyporaceae), with 11 species.

The relative frequency of each species indicates 16 species (41%) as occasional and 71 species (40%) as rare. Of these, 30 species (31%) were represented by a single individual. Four species (19%) were classified as frequent and none was considered abundant (Table 1). The four most frequent species were: *Cerrena hydnoides* (Sw.) Zmitr., with 34 specimens, *Trametes elegans* (Spreng.) Fr. with 31 specimens, *Rigidoporus lineatus* (Pers.) Ryvarden with 30 specimens and *Hymenochaete damicornis* (Link) Lév. with 28 specimens. The four species together represent only 23% of the collected specimens.

**Table 1** - Number of individuals (N) species classification according to frequency (FC) and type of substrate (SU) observed (LW = living wood, DW=dead wood, SO = soil) referring to polypore from a stretch of Amazon forest, near Silvio Braga HPP (Curuá-Una), Wester of Pará state, Brazil.

Order/Family/Species	N	FC	SU	Voucher Number
<b>Hymenochaetales Oberw.</b>				
<b>Hymenochaetaceae Donk</b>				
<i>Coltricia barbata</i> Ryvarden & A.de Meijer	1	Rare	SO	HSTM 12674
<i>Coltricia</i> cf. <i>focicola</i> (Berk. & Curt.) Murrill	2	Rare	SO	HSTM 12679-12680
<i>Coltricia cinnamomea</i> (Jacq.) Murrill	4	Rare	SO	HSTM 12675-12678
<i>Coltricia globispora</i> Gomes-Silva, Ryvarden & Gibertoni	1	Rare	SO	HSTM 12681
<i>Fomitiporia maxonii</i> Murrill	1	Rare	DW	HSTM 12685
<i>Fomitiporia punctata</i> (P. Karst.) Murrill	8	Rare	DW	HSTM 12757-12764
<i>Fomitiporia robusta</i> (P. Karst.) Fiasson & Niemelä	3	Rare	DW	HSTM 12815-13043
<i>Fulvifomes imbricatus</i> L.W. Zhou	1	Rare	DW	HSTM 13133
<i>Fuscoporia callimorpha</i> (Lév.) Groposo, Log.-Leite & Góes-Neto	4	Rare	DW	HSTM 12765-12768
<i>Fuscoporia chrysea</i> (Lév.) Baltazar & Gibertoni	1	Rare	DW	HSTM 12608
<i>Fuscoporia rhabarbarina</i> (Berk.) Groposo, Log.-Leite & Góes-Neto	4	Rare	DW, LW	HSTM 12769-12772
<i>Fuscoporia undulata</i> (Murrill) Bondartseva & S. Herrera	1	Rare	DW	HSTM 12817
<i>Fuscoporia wahlbergii</i> (Fr.) T. Wagner & M. Fisch.	1	Rare	DW	HSTM 12818
<i>Hymenochaete damicornis</i> (Link) Lév.	28	Frequent	SO	HSTM 12773-12800
<i>Hymenochaete luteobadia</i> (Fr.) Höhn. & Litsch.	11	Occasional	DW	HSTM 12801-12811
<i>Hymenochaete rubiginosa</i> (Dicks.) Lév.	1	Rare	DW	HSTM 12812
<i>Inonotus tabacinus</i> (Mont.) G. Cunn.	3	Rare	DW	HSTM 12682-12684
<i>Phellinus fastuosus</i> (Lév.) S. Ahmad	2	Rare	DW, LW	HSTM 12813-12814
<i>Phellinus gilvus</i> (Schwein.) Pat.	20	Occasional	DW	HSTM 12617-12628
<i>Phellinus</i> sp.	1	Rare	DW	HSTM 12629
<i>Phylloporia chrysites</i> (Berk.) Ryvarden	11	Occasional	LW	HSTM 12819-12829
<i>Phylloporia spathulata</i> (Hook.) Ryvarden	3	Rare	SO	HSTM 12830-12832
<b>Polyporales Gäum.</b>				
<b>Cerrena Miettinen, Justo &amp; Hibbett</b>				
<i>Cerrena hydnoides</i> (Sw.) Zmitr.	34	Frequent	DW	HSTM 12914-12947
<b>Fomitopsidaceae Jülich</b>				
<i>Fomitella supina</i> (Sw.) Murrill	1	Rare	DW	HSTM 12913
<i>Fomitopsis roseoalba</i> A.M.S. Soares, Ryvarden & Gibertoni	6	Rare	DW, LW	HSTM 12599-12604
<i>Ranadivia modesta</i> (Kunze ex Fr.) Zmitr.	18	Occasional	DW	HSTM 13107-13124
<b>Meripilaceae Jülich</b>				
<i>Rigidoporus biokoensis</i> (Bres. ex Lloyd) Ryvarden	16	Occasional	DW	HSTM 12833-12848
<i>Rigidoporus lineatus</i> (Pers.) Ryvarden	30	Frequente	DW	HSTM 12849-12878
<b>Meruliaceae Jülich</b>				
<i>Flavodon flavus</i> (Klotzsch) Ryvarden	14	Occasional	DW	HSTM 10790-12368
<i>Mycorrhaphium adustulum</i> (Banker) Ryvarden	1	Rare	DW	HSTM 12888
<i>Podoscypha nitidula</i> (Berk.) Pat.	2	Rare	DW	HSTM 12882-12883
<i>Podoscypha parvula</i> (Lloyd) D.A. Reid	2	Rare	DW	HSTM 12630-12639
<i>Stereopsis hiscens</i> (Berk. & Ravenel) D.A. Reid	3	Rare	SO	HSTM 12884-12886

<i>Stereopsis radicans</i> (Berk.) Reid	1	Rare	SO	HSTM 12887
<b>Phanerochaetaceae Jülich</b>				
<i>Inflatostereum glabrum</i> (Lév.) D.A. Reid	1	Rare	DW	HSTM 12640
<i>Mycorrhaphium adustum</i> (Banker) Ryvarden	1	Rare	DW	HSTM 12888
<b>Panaceae Miettinen, Justo &amp; Hibbett</b>				
<i>Cymatoderma caperatum</i> (Berk. & Mont.) D.A. Reid	3	Rare	DW	HSTM 12879-12881
<i>Panus neostrigosus</i> Drechsler-Santos & Wartchow	1	Rare	DW	HSTM 13021
<b>Polyporaceae Corda</b>				
<i>Amauroderma aurantiacum</i> (Berk.) Torrend	1	Rare	SO	HSTM 12689
<i>Amauroderma calcigenum</i> (Berk.) Torrend	6	Rare	SO	HSTM 12690-12695
<i>Amauroderma elegantissimum</i> Ryvarden & Iturriaga	1	Rare	SO	HSTM 12696
<i>Amauroderma laccatostiptatum</i> Gomes-Silva, Ryvarden & Gibertoni	1	Rare	SO	HSTM 12605
<i>Amauroderma omphalodes</i> (Berk.) Torrend	8	Rare	SO	HSTM 12697-12704
<i>Amauroderma partitum</i> (Berk.) Wakef.	7	Rare	SO	HSTM 12705-12711
<i>Amauroderma praetervisum</i> (Pat.) Torrend	16	Occasional	DW, SO	HSTM 12712-12727
<i>Amauroderma rude</i> (Berk.) Torrend	3	Rare	SO	HSTM 12728-12730
<i>Amauroderma schomburgkii</i> (Mont. & Berk.) Torrend	12	Occasional	SO	HSTM 12731-12740
<i>Amauroderma sprucei</i> (Pat.) Torrend	9	Occasional	SO	HSTM 12741-12749
<i>Amauroderma subsessile</i> Gomes-Silva, Ryvarden & Gibertoni	1	Rare	DW	HSTM 12606
<i>Atroporus diabolicus</i> (Berk.) Ryvarden	12	Occasional	DW	HSTM 12641-12652
<i>Atroporus rufoatratus</i> (Berk.) Palacio, Reck & Robledo	8	Rare	DW	HSTM 12653-12659
<i>Bresadolia uda</i> (Jungh.) Audet	4	Rare	DW, SO	HSTM 13062-13065
<i>Cerioporus cavernulosus</i> (Berk.) Zmitr.	3	Rare	DW	HSTM 12903-12905
<i>Cerioporus flavus</i> (Sw.) Zmitr.	4	Rare	DW	HSTM 13016-13019
<i>Cerioporus mollis</i> (Sommerf.) Zmitr. & Kovalenko	1	Rare	DW	HSTM 12902
<i>Cerioporus varius</i> (Pers.) Zmitr. & Kovalenko	3	Rare	DW	HSTM 13066-13068
<i>Earliella scabrosa</i> (Pers.) Gilb. & Ryvarden	2	Rare	DW	HSTM 12909-12910
<i>Favolus brasiliensis</i> (Fr.) Fr.	3	Rare	DW	HSTM 12660-12662
<i>Favolus grammocephalus</i> (Berk.) Imazeki	2	Rare	DW	HSTM 13046-13047
<i>Fomes fasciatus</i> (Sw.) Cooke	2	Rare	DW	HSTM 12911-12912
<i>Fomes fomentarius</i> (L.) Fr.	1	Rare	DW	HSTM 12663
<i>Funalia caperata</i> (Berk.) Zmitr. & Malysheva	14	Occasional	DW	HSTM 12889-12901
<i>Ganoderma amazonense</i> Weir	1	Rare	SO	HSTM 12750
<i>Ganoderma australe</i> (Fr.) Pat.	2	Rare	DW	HSTM 12751-12752
<i>Ganoderma resinaceum</i> Boud.	5	Rare	DW	HSTM 12668-12756
<i>Haddowia longipes</i> (Lév.) Steyaert	2	Rare	SO	HSTM 12669-12670
<i>Lentinus crinitus</i> (L.) Fr.	10	Occasional	DW	HSTM 12686-12971
<i>Lentinus velutinus</i> Fr.	7	Rare	DW	HSTM 12972-12978
<i>Lenzites betulina</i> (L.) Fr.	19	Occasional	DW	HSTM 12979-12997
<i>Lopharia cinerascens</i> (Schwein.) G.Cunn.	1	Rare	DW	HSTM 12998
<i>Megasporoporia setulosa</i> (Henn.) Rajchenb.	3	Rare	DW	HSTM 12906-12908
<i>Microporellus dealbatus</i> (Berk. & M.A. Curtis) Murrill	8	Rare	SO	HSTM 12999-13006
<i>Microporellus iguazuensis</i> Rajchenb.	2	Rare	SO	HSTM 13007-13008
<i>Microporellus obovatus</i> (Jungh.) Ryvarden	7	Rare	DW	HSTM 13009-13015
<i>Neodictyopus atlanticae</i> (Mont.) Palacio, Robledo & Drechsler-Santos	2	Rare	DW	HSTM 12664-12665

<i>Neodictyopus gugliottae</i> Palacio, Grassi & Robledo	1	Rare	DW	HSTM 12666
<i>Neofavolus alveolaris</i> (DC.) Sotome & T. Hatt.	2	Rare	DW	HSTM 13044-13045
<i>Nigrofomes melanoporus</i> (Mont.) Murrill	1	Rare	DW	HSTM 13020
<i>Perenniporia medulla-panis</i> (Jacq.) Donk	14	Occasional	DW, LW	HSTM 13022-13035
<i>Perenniporia ochroleuca</i> (Berk.) Ryvarden	4	Rare	DW	HSTM 13036-13039
<i>Perenniporia stipitata</i> Ryvarden	4	Rare	DW	HSTM 13040-13042
<i>Polyporus guianensis</i> Mont.	11	Occasional	DW	HSTM 13048-13058
<i>Polyporus leprieurii</i> Mont.	3	Rare	DW	HSTM 13059-13051
<i>Porogramme albocincta</i> (Cooke & Masee) Gibertoni	3	Rare	LW	HSTM 12671-12673
<i>Pycnoporus sanguineus</i> (L.) Murrill	7	Rare	DW	HSTM 13069-13075
<i>Ranadivia modesta</i> (Kunze ex Fr.) Zmitr.	18	Occasional	DW	HSTM 13107-13124
<i>Trametes elegans</i> (Spreng.) Fr.	31	Frequent	DW	HSTM 13076-13106
<i>Trametes leonina</i> (Klotzsch) Imazeki	1	Rare	DW	HSTM 12667
<i>Trametes pubescens</i> (Schumach.) Pilát	1	Rare	DW	HSTM 13125
<i>Trametes variegata</i> (Berk.) Zmitr., Wasser & Ezhov	17	Occasional	DW	HSTM 12948-12964
<i>Trametes versicolor</i> (L.) Lloyd	4	Rare	DW	HSTM 13126-13129
<i>Trichaptum byssogenum</i> (Jungh.) Ryvarden	2	Rare	DW	HSTM 13130-13131
<i>Trichaptum perrottetii</i> (Lév.) Ryvarden	1	Rare	DW	HSTM 13132

Source: Authors.

The substrates most used by the polypores collected were dead trees, associated with 68 species (70%), the species observed in the soil totaled 23 (24%) and six species (6%) occur in live trees. Five species, which together make up 5% of the samples, were found to colonizing two types of substrates (Table 1).

The 16 new occurrences of polypores for the state of Pará are shown in Table 2 and Figure 3A-O. One of these records is also new for America (Figure 4). They are three species of Hymenochaetales and 13 of Polyporales. Of these, 13 species occur in the domain of the Brazilian Amazon.



**Table 2** - New records of polypore fungi for Pará state and their distribution in other Brazilian states and outside America.  
**States:** AC= Acre, AM = Amazonas, AP = Amapá, BA = Bahia, CE = Ceará, GO = Goiás, MA = Maranhão, MS = Mato Grosso do Sul, MT = Mato Grosso, PE = Pernambuco, PI = Piauí, PR = Paraná, RJ = Rio de Janeiro, RN = Rio Grande do Norte, RO = Rondônia, RR = Roraima, RS= Rio Grande do Sul, SC = Santa Catarina, SE = Sergipe, SP = São Paulo, TO = Tocantins.

Species	Distribution	Fig.
<i>Amauroderma aurantiacum</i>	GO, RO, SP	3A
<i>Cerioporus mollis</i>	AC, PR, RS, SC, SP	3B
<i>Cymatoderma caperatum</i>	AM, BA, PR, RO, RS, SC, SP	3C
<i>Flavodon flavus</i>	AM, BA, CE, MA, PE, PI, TO	3D
<i>Fomes fomentarius</i>	AM, MG, PE, RJ, SC	3E
<i>Fomitiporia maxonii</i>	AM, PE, RO, RS, SC, SP	3F
<i>Fuscoporia callimorpha</i>	AP, BA, MT, PE, PR, RJ, RO, SC, SE	3G
<i>Fuscoporia chrysea</i>	PE, RO, SC	3H
<i>Inflatostereum glabrum</i>	AM, RR	3I
<i>Lenzites betulina</i>	AP, PR, RS, SC, SP	3J
<i>Neodictyopus atlanticae</i>	PR, SC	3K
<i>Neodictyopus gugiottae</i>	SC	3L
<i>Neofavolus alveolares</i>	RN, RR, RS	3M
<i>Perenniporia ochroleuca</i>	AM, MS, PE, RS, SC, SP	3N
<i>Stereopsis hiscens</i>	AM, PE, PR, RJ, RS, SP	3O
<i>Fulvifomes imbricatus</i>	Thailand	4A-C

Source: Authors.

**Figure 3:** Basidiomes of the polypores registered for the first time for the state of Pará. A) *Amauroderma aurantiacum*, B) *Cerioporus mollis*, C) *Cymatoderma caperatum*, D) *Flavodon flavus*, E) *Fomes fomentarius*, F) *Fomitiporia maxonii*, G) *Fuscoporia callimorpha*, H) *F. chrysea*, I) *Inflatostereum glabrum*, J) *Lenzites betulina*, K) *Neodictyopus atlanticae*, L) *N. gugiottae*, M) *Neofavolus alveolares*, N) *Perenniporia ochroleuca*, O) *Stereopsis hiscens*. **Scale bars:** 1 cm.



Source: Authors.

**Figure 1:** *Fulvifomes imbricatus*, first record for America. A) Imbricate basidiomes in the forest, B) upper surface, C) pore surface poroid. **Scale bars:** A = 5 cm; B, C = 1 mm.



Source: Authors.

#### 4. Discussion

Fungal conservation will always be a challenge, given the enormous mycodiversity associated with its undoubted importance for the functioning of ecosystems (Willis, 2018). Although the sampling efforts and the information produced by this study contribute to the knowledge about the diversity, distribution and ecology of the polypores in Amazon, it is still of great importance and need to expand the studies and intensify the collecting and of the taxonomic and systematic studies.

The intensive training of human resources specialized in mycology should be considered to cover the areas insufficiently studied (Maia et al., 2015). Like this study, which registers 51% of the species known in the state of Pará considering the latest surveys such as Flora do Brasil (<http://floradobrasil.jbrj.gov.br>), Maia et al. (2015), Medeiros et al. (2015). This perspective demonstrates the importance of inventories and incentives for research, especially in areas that have never been explored, such as the one in study.

Shannon index points to a high diversity when compared to other studies in tropical forests, such as Lindblad (2001), Hattori (2005) and Adarsh et al. (2019). In another biome, as in the Atlantic Forest, the diversity is lower according to Komone et al. (2018) in the only study on alpha diversity in Brazil so far.

The studied area shows a great potential for the occurrence of polypores fungi, given the registration of 91 species, 16 of which are new records for Pará state. This Number may be even greater considering the value of 118 estimated. It is worth mentioning that Sotão et al. (2009), Gibertoni et al. (2013) and Medeiros et al. (2015) performed a greater sampling effort in Caxiuanã National Forest, in the same Brazilian state, revealing 87, 96 and 74 species of polypores, respectively. These observations are also important due to the human threat that exists in the study area, such as increasing deforestation in favor of agriculture (Domingues et al., 2014; Villela & Bueno, 2017; Schwarzmüller & Kastner, 2022), electrical transmission network passages that depart from HPP Silvio Braga and opening of roads.

The families Polyporaceae and Hymenochaetaceae are the most diverse among the polypores (Wijayawardene et al., 2020; 2022) and in this study the richest, which corroborates with referenced studies. This result was also corroborated by Gibertoni (2008), Sotão et al. (2009), Gibertoni et al. (2013) and Medeiros et al. (2015) for Caxiuanã National Forest and Xavier et al. (2018), for Serra do Návio, both in Pará and by Soares et al. (2014) for Amapá National Forest, Amapá state.

In tropical areas, polypores are considered to be a diverse group, however, most species can be considered rare or occasional (e.g., Nuñez, 1996; Lindblad, 2001; Gibertoni et al., 2008; Yamashita et al., 2009; Soares et al., 2014; Medeiros et al., 2015). The less abundant or rare species in this study 71 (40%), can be threatened with the risk of disappearing due to the advancement of human activities over natural ecosystems, which highlights the need to conserve the areas where they occur (Raphael et al., 2007; Molina et al., 2011), denoting the growing importance fungal monitoring, particularly from countries with high biodiversity and disproportionate data gaps (Stephenson et al., 2022).

It is convenient to understand the way of life of polypores, as it significantly impacts mycodiversity and its relationship with ecosystems. Although the majority of polypores, mainly from the Polyporaceae and Hymenochaetaceae families, are saprobes and degrade dead trees (Deacon, 2006; Dawson & Jönsson, 2020), some species can be parasitic on live trees, and may even switch to saprobes when they kill the sapwood of the live tree (Agrios, 2005). The species found in the soil, can be associated with the roots of the trees and form ectomycorrhizae (Tedersoo et al., 2007).

Eleven species of *Amauroderma* found in the study area occur in the Amazon domain (Gomes-Silva et al., 2015), being the first report of *A. aurantiacum* for Pará state. *Inflatostereum glabrum*, *Cymatoderma caperatum* and *Stereopsis hiscens* also occur in the Amazon domain (Reid, 1965), but had not been registered in Pará yet. *Neodictyopus atlanticae* and *N. gugliottae* are part of a cryptic species of *Polyporus dictyopus* Mont. with distribution in the Atlantic Forest (Palacio et al., 2017), thus expanding its distribution to the Amazon. *Fomitiporia maxonii*, *Fuscoporia callimorpha* and *F. chryseus* are the only species of new Hymenochaetales records for Pará state, with records in the Atlantic Forest domains (Gomes-Silva &

Gibertoni, 2009; Baltazar & Gibertoni, 2006; Groposo et al., 2007; Westphalen & Silveira, 2013).

*Cerioporus mollis*, *Flavodon flavus*, *Fomes fomentarius*, *Lenzites betulina*, *Perenniporia ochroleuca*, *Neofavolus alveolaris* are species with wide occurrence in states that contemplate the Atlantic Forest and Caatinga biome (Baltazar & Gibertoni, 2009; Drechsler-Santos et al., 2009), of which the first five occur in the Brazilian Amazon (Gomes-Silva & Gibertoni, 2009).

*Fulvifomes imbricatus* is mainly distinguished by its imbricated basidiome and double context with a black cuticle, pores of 7–9 per mm and yellow basidiospores, ellipsoids 3.2–4.5 µm. The species examined macroscopically resembles *F. johnsonianus* (murrill) Y.C. Dai and *F. collinus* (Y.C. Dai & Nimedá) Y.C. Dai (Dai, 2010). They differ from *F. collinus* by presenting a substrate projection from 10 to 15 cm while *F. collinus* has a projection of a maximum of 5 cm, in addition it has 5–6 pores on the hymenial surface, whereas in *F. imbricatus* this variation is up to 9 pores/mm, in addition *F. collinus* does not present a double context.

## 5. Conclusion

The present study represents an increment to the knowledge of polypores in Pará state, and offer an important contribution to future studies, especially in support of the elaboration of a management and conservation plan for the areas where species occur, an important fact in view of the sustainable exploitation of the areas and tropical forest species.

We observed that the standardization of the sampling effort is a point that must be considered in studies with taxonomic approaches and fungal diversity, and even so, this type of study comes up against difficulties, which reaffirms the importance and need for resampling in the study areas.

## Acknowledgments

The authors thank PhD Genivaldo Alves who kindly identified some species of Hymenochaetales, the Universidade Federal do Oeste do Pará (UFOPA) for the infrastructure offered to carry out this work, ICMBio for the collection license, Eletrobrás Santarém for the support during the collections in the study area. The first author thanks the Postgraduate Program in Biodiversity and the Coordenação de Aperfeiçoamento de Pessoal de Nível Superior (CAPES) for the Master's degree scholarship.

## References

- Adarsh, C. K., Vidyasagan, K. & Ganesh, P. N. (2019). The diversity and distribution of Polypores (Basidiomycota: Aphyllophorales) in wet evergreen and shola forests of Silent Valley National Park, southern Western Ghats, India, with three new records. *Journal of Threatened Taxa*, 11(7), 13886-13909. <https://doi.org/10.11609/jot.3856.11.7.13886-13909>.
- Agrios, G. N. (2005). *Phytopathology*. Elsevier.
- Alvares, C. A., Stape, J. L., Sentelhas, P. C., De Moraes, G., Leonardo, J. & Sparovek, G. (2013). Köppen's climate classification map for Brazil. *Meteorologische Zeitschrift*, 22(6), 711-728. <https://doi.org/10.1127/0941-2948/2013/0507>.
- Baltazar, J. M. & Gibertoni, T. B. (2009). A checklist of the aphyllophoroid fungi (Basidiomycota) recorded from the Brazilian Atlantic Forest. *Mycotaxon*, 109(1), 439-442.
- Bekai, L. H., Smania, E. D. F. A., Silva Riehl, C. A. & Smania Jr, A. (2012). *Antrodia albida* (Fr.) Donk (higher Basidiomycetes) as a source of metabolites of biotechnological interest. *International Journal of Medicinal Mushrooms*, 14(2), 161-168. <https://doi.org/10.1615/IntJMedMushr.v14.i2.40>.
- Boddy, L. & Heilmann-Clausen, J. (2008). Basidiomycete Community Development in Temperate Angiosperm Wood. In L. Boddy, J. C. Frankland & P. Van West (Org.), *Ecology of saprotrophic Basidiomycetes* (pp. 211–236). London: Academic Press.
- Colwell, R. K., Mao, C. X. & Chang, J. (2004). Interpolating, extrapolating and comparing incidence-based species accumulation curves. *Ecology*, 85(10), 2717-2727. <https://doi.org/10.1890/03-0557>.
- Costanza, R., Groot, R., Sutton, P., Van Der Ploeg, S., Anderson, S. J., Kubiszewski, I. & Turner, R. K. (2014). Changes in the global value of ecosystem services. *Global Environmental Change*, 26, 152-158. <https://doi.org/10.1016/j.gloenvcha.2014.04.002>.

- Costa-Rezende, D. H., Gugliotta, A. M., Goes-Neto, A., Reck, M. A., Robledo, G. L. & Drechsler-Santos, E. R. (2016). *Amauroderma calcitum* sp. nov. and notes on taxonomy and distribution of *Amauroderma* species (Ganodermataceae). *Phytotaxa*, 244(2), 101-124. <https://doi.org/10.11646/phytotaxa.244.2.1>
- Dai, Y. (2010). Hymenochaetales in China. *Fungal Diversity*, 45(1), 131-343. <https://doi.org/10.1007/s13225-010-0066-9>.
- Dawson, S. K. & Jönsson, M. (2020). Just how big is intraspecific trait variation in basidiomycete wood fungal fruit bodies?. *Fungal Ecology*, 46, 100865.
- Deacon, J. W. (2006). *Fungal biology*. Blackwell Publishing, Malden, 380 pp.
- Domingues, M. S. D., Bermann, C. & Sidneide, M. S. (2014). A produção de soja no Brasil e sua relação com o desmatamento na Amazônia. *Revista Presença Geográfica*, 1(01), 32-47.
- Drechsler-Santos, E. R., Gibertoni, T. B., Góes-Neto, A. & Cavalcanti, M. A. Q. (2009). A re-evaluation of the lignocellulolytic Agaricomycetes from the Brazilian semi-arid region. *Mycotaxon*, 108, 241 - 244.
- Efron, B. & Tibshirani, R. (1993). *An Introduction to the Bootstrap*. Chapman and Hall, New York, 435 pp.
- Fidalgo, O. & Bononi, V. L. R. (1999). *Técnicas de coleta, preservação e herborização de material botânico*. 1 ed., Secretaria do Meio Ambiente, Rio de Janeiro.
- Furtado, J. S. (1981). Taxonomy of *Amauroderma* (Basidiomycetes, Polyporaceae). *Memoirs of the New York Botanical Garden*, 34, 1-109.
- Gibertoni, T. B. (2008). Polyporoid fungi (Agaricomycetes, Basidiomycota) in the Estação Científica Ferreira Penna (State of Pará, Brazilian Amazonia): diversity and ecological aspects. *Scientifica Acta*, 2(2), 70-74.
- Gibertoni, T. B., Medeiros, P. S., Soares, A. M., Gomes-Silva, A. C., Santos, P. J., ... & Ferre Savino, E. (2016). The distribution of polypore fungi in endemism centres in Brazilian Amazonia. *Fungal ecology*, 20, 1-6. <https://doi.org/10.1016/j.funeco.2015.09.012>.
- Gibertoni, T. B., Ryvardeen, L., Bernicchia, A. & Savino, E. (2013). Poroid fungi (Agaricomycetes, Basidiomycota) in the National Caxiuanã Forest. In P.L.B. Lisboa (Ed.). *Caxiuanã: paraíso ainda preservado*. (pp. 397-410). Pará: Museu Paraense Emílio Goeldi.
- Gomes-Silva, A. C. & Gibertoni, T. B. (2009). Checklist of the aphyllporaceous fungi (Agaricomycetes) of the Brazilian Amazonia. *Mycotaxon*, 108, 319-322.
- Gomes-Silva, A.C., Lima-Júnior, N., Malosso, E., Ryvardeen, L., Gibertoni, T., 2015. Delimitation of taxa in *Amauroderma* (Ganodermataceae, Polyporales) based in morphology and molecular phylogeny of Brazilian specimens. *Phytotaxa*, v. 227, (3), 201-228. <https://doi.org/10.11646/phytotaxa.00.0.0>.
- Gomes-Silva, A. C., Medeiros, P. S., Soares, A. M. S., Sotão, H. M. P., Ryvardeen, L. & Gibertoni, T. B. (2014). Two new species of *Rigidoporus* (Agaricomycetes) from Brazil and new records from the Brazilian Amazonia. *Phytotaxa*, 156(4), 191-200. <https://doi.org/10.11646/phytotaxa.00.0.0>.
- Groposo, C., Loguercio-Leite, C. & Góes-Neto, A. (2007). *Fuscoporia* (Basidiomycota, Hymenochaetales) in Southern Brazil. *Mycotaxon*, 101(1), 55-63.
- Hattori, T. (2005). Diversity of wood-inhabiting polypores in temperate forests with different vegetation types in Japan. *Fungal Diversity*, 18, 73-88.
- Hawksworth, D. L. & Lücking, R (2017). Fungal Diversity Revisited: 2.2 to 3.8 million Species. *Microbiology spectrum*, 5(4), 1-17.
- Hibbett, D. & Binder, M. (2002). Evolution of complex fruiting-body morphologies in homobasidiomycetes. *Proceedings of the Royal Society of London. Series B: Biological Sciences*, 269(1504), 1963-1969. <https://doi.org/10.1098/rspb.2002.2123>.
- Jati, D. A. & Silva, J. T. (2017). Estudos geo-hidrológicos da bacia do rio Curuá-Una, Santarém, Pará. Aplicação do modelo hidrológico de grandes bacias (MGB-IPH). *Revista Brasileira de Geografia Física*, 10(4), 1296-1311.
- Justo, A., Miettinen, O., Floudas, D., Ortiz-Santana, B., Sjökvist, E., Lindner, D., ... & Hibbett, D. S. (2017). A revised family-level classification of the Polyporales (Basidiomycota). *Fungal biology*, 121(9), 798-824.
- Kindt, R. & Coe, R. (2005). *Tree diversity analysis. A manual and software for common statistical methods for ecological and biodiversity studies*. World Agroforestry Centre, Kenya.
- Komonen, A., Kokkonen, M., Araujo, L. S., Halme, P. & Lopes-Andrade, C. (2018). Polypore Communities and Their Substrate Characteristics in Atlantic Forest Fragments in Southeast Brazil. *Tropical Conservation Science*, 11, 1940082918777118. <https://doi.org/10.1177/1940082918777118>.
- Kueppers, H. (1982). *Color Atlas: A Practical Guide for Color Mixing*. Barrons Educational Series Incorporated. New York.
- Lindblad, I. (2001). Diversity of poroid and some corticoid wood-inhabiting fungi along the rainfall gradient in tropical forests, Costa Rica. *Journal of Tropical Ecology*, 17(3), 353-369. <https://doi.org/10.1017/S0266467401001249>.
- Lomascolo, A., Uzan-Boukhris, E., Herpoël-Gimbert, I., Sigoillot, J. C. & Lesage-Meessen, L. (2011). Peculiarities of *Pycnoporus* species for applications in biotechnology. *Applied microbiology and biotechnology*, 92(6), 1129-1149. <https://doi.org/10.1007/s00253-011-3596-5>.
- Londsdale, D., Pautasso, M. & Holdenrieder, O. (2008). Wood-decaying fungi in the forest: conservation needs and management options. *European Journal of Forest Research*, 127(1), 1-22. <https://doi.org/10.1007/s10342-007-0182-6>.
- Maciel, M. J. M., Castro, E. S. A. & Ribeiro, H. C. T. (2010). Industrial and biotechnological applications of ligninolytic enzymes of the Basidiomycota: A review. *Electronic Journal of Biotechnology*, 13(6), 14-15. <https://doi.org/10.2225/vol13-issue6-fulltext-2>.

- Magurran, A. (2004). *Measuring biological diversity*. Blackwell Publishing, United Kingdom.
- Maia, L. C., Carvalho-Junior, A. A. D., Cavalcanti, L. D. H., Gugliotta, A. D. M., Drechsler-Santos, E. R., Santiago, A. L. D. A., ... & Silva, V. F. (2015). Diversity of Brazilian fungi. *Rodriguésia*, 66, 1033-1045. <https://doi.org/10.1590/2175-7860201566407>.
- Medeiros, P. S., Cattanio, J.H. & Sotão, H. M. P. (2015). Richness and relation of ligninolytic poroid fungi (Agaricomycetes) with the substrate in the Brazilian Amazon forest. *Boletim do Museu Paraense Emílio Goeldi-Ciências Naturais*, 10(3), 423-436. <https://doi.org/10.46357/bcnaturais.v10i3.475>
- Medeiros, P. S., Sotão, H. M. P., Gibertoni, T. B. & Cattanio, J. H. (2013). Fungos poroides (Agaricomycetes) no sítio do Programa de Biodiversidade da Amazônia (PPBio) em Caxiuana. In P. L. B. Lisboa (Ed.), *Caxiuana: paraíso ainda preservado*. (pp. 375-385) Pará: Museu Paraense Emílio Goeldi.
- Molina, R., Horton, T. R., Trappe, J. M. & Marcot, B. G. (2011). Addressing uncertainty: how to conserve and manage rare or little-known fungi. *Fungal Ecology*, 4(2), 134-146. <https://doi.org/10.1016/j.funeco.2010.06.003>.
- Núñez, M. (1995). *Polyporus* (Basidiomycotina) and related genera. *Synopsis Fungorum*, 10, 1-85.
- Núñez, M., 1996. Fructification of Polyporaceae s.l. (Basidiomycotina) along a gradient of altitude and humidity in the Guanacaste Conservation Area (Costa Rica). *Journal of Tropical Ecology*, v. 12, (6), 893 - 898. <https://doi.org/10.1017/S0266467400010154>.
- Oksanen, J., Blanchet, F. G., Kindt, R., Legendre, P., Minchin, P. R., O'hara, R. B., ... & Wagner, H. (2015). *Vegan: Community Ecology Package*. R package version 2.3-0, Vienna.
- Palacio, M., Robledo, G. L., Reck, M. A., Grassi, E., Góes-Neto, A., Drechsler-Santos, E. R. (2017). Decrypting the *Polyporus dictyopus* complex: Recovery of *Atroporus* Ryvardeen and segregation of *Neodictyopus* gen. nov. (Polyporales, Basidiomycota). *PLoS One*, 12(10), e0186183. <https://doi.org/10.1371/journal.pone.0186183>.
- R Core Team., 2012. *R: A language and environment for statistical computing*. R Foundation for Statistical Computing. Vienna, Austria.
- Raphael, M. G., Molina, R. & Molina, N. (2007). *Conservation of Rare or Little-Known Species-Biological*. DC Island Press, Washington.
- Reid, D. A. (1965). A monograph of the stipitate steroid fungi. *Beih. Nova Hedwigia*, 18, 1-388.
- Ritter, C. D., Mccrate, G., Nilsson, R. H., Fearnside, P. M., Palme, U. & Antonelli, A. (2017). Environmental impact assessment in Brazilian Amazonia: Challenges and prospects to assess biodiversity. *Biological Conservation*, 206, 161-168. <https://doi.org/10.1016/j.biocon.2016.12.031>.
- Ryvardeen, L. & Johansen, I. (1980). *A preliminary polypore flora of East Africa*. Fungiflora, Oslo, 636 p.
- Ryvardeen, L., 2004. *Neotropical Polypores: Part 1: Introduction, Ganodermataceae & Hymenochaetaceae*. Fungiflora, Oslo, 229 p.
- Ryvardeen, L., 2015. Neotropical Polypores part 2. Polyporaceae: *Abortiporus - Meripilus*. *Synopsis Fungorum*, 34, 229-443.
- Ryvardeen, L., 2016. Neotropical Polypores Part 3. Polyporaceae: *Obba - Wrightoporia*. *Synopsis Fungorum*, 36, 446-612.
- Santos, A. J. (2004). Estimativas de riqueza em espécies. In J. R. Cullen, R. Rudran & C. V. Pádua, (Eds.) *Métodos de estudos em biologia da conservação e manejo da vida silvestre*. (pp. 19-40), Paraná: Universidade Federal do Paraná.
- Medeiros, P. S., Sotão, H. M. P., Gibertoni, T. B. & Cattanio, J. H. (2013). Fungos poroides (Agaricomycetes) no sítio do Programa de Biodiversidade da Amazônia (PPBio) em Caxiuana. In P. L. B. Lisboa (Ed.) *Caxiuana: paraíso ainda preservado*. (pp. 375-385). Pará: Museu Paraense Emílio Goeldi.
- Schwarzmueller, F. & Kastner, T. (2022). Agricultural trade and its impacts on cropland use and the global loss of species habitat. *Sustainability Science*, 1-15.
- Singh, V., Shukla, S., & Singh, A. (2021). The principal factors responsible for biodiversity loss. *Open Journal of Plant Science*, 6(1), 011-014.
- Soares, A. M. S., Sotão, H. M. P., Medeiros, P. S. & Gibertoni, T. (2014). Riqueza de fungos poliporoides (Agaricomycetes, Basidiomycota) em uma floresta ombrófila densa no Amapá, Amazônia brasileira. *Boletim do Museu de Biologia Mello Leitão*, 35, 5-18.
- Sotão, H. M. P., Campos, E. L., Gugliotta, A. M. & Costa, S. P. S. E. C. (2003). Fungos macroscópicos: Basidiomycetes. In M. E. B. Fernandes (Ed.), *Os manguezais da costa norte brasileira*. (pp. 375-385). Maranhão: Fundação Rio Bacanga.
- Sotão, H. M. P., Gibertoni, T. B., Maziero, R., Baseia, I., Medeiros, O. S., Martins-Júnior, A. & Capelari, M. (2009). Fungos macroscópicos da Floresta nacional de Caxiuana, Pará, Brasil: Basidiomycota (Agaricomycetes). In P. L. B. Lisboa (Ed.), *Caxiuana: Desafios para conservação de uma Floresta Nacional na Amazônia*. (pp. 383-396). Pará: Museu Paraense Emílio Goeldi.
- Sotão, H. M. P., Hennen, J. F., Gugliotta, A. M., Melo, O. A. & Campos, E. L. (1997). Os fungos - Basidiomycotina. In P. L. B. Lisboa (Ed.), *Caxiuana*. (pp. 213-219). Pará: Museu Paraense Emílio Goeldi.
- Stephenson, P. J., Londoño-Murcia, M. C., Borges, P. A., Claassens, L., Frisch-Nwakanma, H., Ling, N., ... & Fumagalli, L. (2022). Measuring the Impact of Conservation: The Growing Importance of Monitoring Fauna, Flora and Funga. *Diversity*, 14(10), 824.
- Tabarelli, M., Da Silva, J. M. C. & Gascon, C. (2004). Forest fragmentation, synergisms and the impoverishment of neotropical forests. *Biodiversity & Conservation*, 13(7), 1419-1425. <https://doi.org/10.1023/B:BIOC.0000019398.36045.1b>.
- Tedersoo, L., Suvi, T., Beaver, K. & Saar, I., 2007. Ectomycorrhizas of *Coltricia* and *Coltriciella* (Hymenochaetales, Basidiomycota) on Caesalpiniaceae, Dipterocarpaceae and Myrtaceae in Seychelles. *Mycological Progress*, 6(2), 101-107. <https://doi.org/10.1007/s11557-007-0530-4>
- Teixeira, A.R., 1995. Método para estudo das hifas do basidiocarpo de fungos poliporáceos. Manual, n.6, Instituto de Botânica, São Paulo.

- Tsigain, F. T., Metsebing, B. P., Mossebo, D. C., Ryvardeen, L. R., Oba, R., Guifo, C., ... & Fokoua, U. L. (2022). Enzymatic Activities, Characteristics of Wood-Decay and Wood Substrate Specificity within Genera of Some Wood-Rotting Basidiomycetes from Cameroon and Tropical Africa. *European Journal of Biology and Biotechnology*, 3(1), 11-23.
- Urcelay, C. & Robledo, G. 2004. Community structure of polypores (Basidiomycota) in Andean alder wood in Argentina: Functional groups among wood decay fungi?. *Austral Ecology*, 29(4), 471-476. <https://doi.org/10.1111/j.1442-9993.2004.01387.x>
- Veloso, H. P., Rangel Filho, A. L. R. & Lima, J. C. A. (1991). *Classificação da vegetação brasileira adaptada a um sistema universal*. IBGE, Departamento de recursos naturais e estudos ambientais, Rio de Janeiro.
- Villela, R. & Bueno, R. S. (2017). A expansão do desmatamento no estado do Pará: População, dinâmicas territoriais e escalas de análise. *Anais*, 1-15.
- Westphalen, M. C. & Silveira, R. M. B. D. (2013). Resupinate Polypores from mixed ombrophilous forests in southern Brazil. *Mycotaxon*, 122, 111-122.
- Wijayawardene, N. N., Hyde, K. D., Al-Ani, L. K. T., Tedersoo, L., Haelewaters, D., Rajeshkumar, K. C., ... & Thines, M. (2020). Outline of Fungi and fungus-like taxa. *Mycosphere*, 11(1), 1060-1456. <https://doi.org/10.5943/mycosphere/11/1/8>.
- Wijayawardene, N. N., Hyde, K. D., Dai, D. Q., Sánchez-García, M., Goto, B. T., Saxena, R. K., ... & Thines, M. (2022). Outline of Fungi and fungus-like taxa-2021. *Mycosphere*, 13(1), 53-453. <https://doi.org/10.5943/mycosphere/13/1/2>.
- Willis, K. J. (2018). *State of the world's fungi 2018*, Report. Royal Botanic Gardens Kew, United Kingdom.
- Xavier, W. K. S., Sotão, H. M. P., Soares, A. M. S., Gibertoni, T. B., Rodrigues, F. J. & Ryvardeen, L. (2018). Richness of poroid Agaricomycetes from Serra do Navio, eastern Amazonia, Brazil, with a new record of *Oxyporus lacera* for Brazil. *Boletim Museu Paraense Emílio Goeldi*, 13(3), 303-315. <https://doi.org/10.46357/bcnaturais.v13i3.339>.
- Yamashita, S., Hattori, T., Ohkubo, T. & Nakashizuka, T. (2009). Spatial distribution of the basidiocarps of aphyllporaceous fungi in a tropical rainforest on Borneo Island, Malaysia. *Mycological Research*, 113(110), 1200-1207. <https://doi.org/10.1016/j.mycres.2009.08.004>.
- Yang, S., Limpens, J., Sterck, F. J., Sass-Klaassen, U., Cornelissen, J. H., Hefting, M., ... & Poorter, L. (2021). Dead wood diversity promotes fungal diversity. *Oikos*, 130(12), 2202-2216.